

IN THE UNITED STATES DISTRICT COURT
FOR THE SOUTHERN DISTRICT OF NEW YORK

ENDO PHARMACEUTICALS INC.,

Plaintiff,

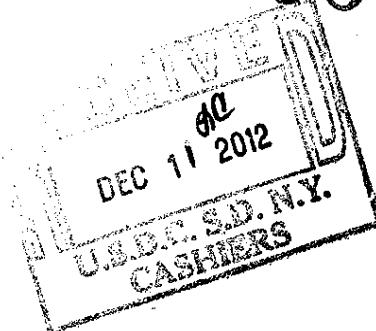
v.

ACTAVIS INC. and
ACTAVIS SOUTH ATLANTIC LLC,

Defendants.

C.A. No.

12 CV 8985



COMPLAINT

Plaintiff Endo Pharmaceuticals Inc. ("Endo") for its Complaint against Defendants Actavis Inc. and Actavis South Atlantic LLC (collectively "Defendants"), allege as follows:

PARTIES

1. Plaintiff Endo is a Delaware corporation, having its principal place of business at 100 Endo Boulevard, Chadds Ford, Pennsylvania 19317. Endo is a specialty pharmaceuticals company engaged in the research, development, sale and marketing of prescription pharmaceuticals used, among other things, to treat and manage pain. Endo markets and distributes OPANA® ER, an innovative opioid painkiller designed to be crush-resistant (alternatively referred to herein as "Opana ER CRF")

2. Upon information and belief, defendant Actavis Inc. is a corporation organized and existing under the laws of the State of Delaware, having its headquarters and principal place of business at 60 Columbia Road, Building B, Morristown, New Jersey 07960. Actavis Inc. is a pharmaceutical company engaged in the development, manufacture, sale and marketing of generic pharmaceuticals for sale and use throughout the United States, including in this judicial

district.

3. Upon information and belief, Actavis South Atlantic LLC (“ASA”) is a limited liability company, organized and existing under the laws of the State of Delaware, having its principal place of business at 13800 N.W. 2nd Street, Suite 190, Sunrise, Florida 33325. ASA is a pharmaceutical company engaged in the development, manufacture, sale and marketing of generic pharmaceuticals for sale and use throughout the United States, including in this judicial district.

4. Upon information and belief, Actavis Inc. controls and directs the operations of ASA, and ASA and Actavis Inc. have acted as each other’s alter ego, agent, and partner in the development, manufacturing, distribution, offer for sale, and sale in this judicial district of the infringing product at issue.— generic, non crush-resistant “Oxymorphone Hydrochloride Extended-Release Tablets CII” (“Actavis Generic Oxymorphone ER Tablets”).

NATURE OF ACTION

5. This is an action arising under the Patent Laws of the United States, 35 U.S.C. § 100, *et seq.*

JURISDICTION AND VENUE

6. This Court has jurisdiction over the subject matter of this action pursuant to 28 U.S.C. §§ 1331 and 1338(a) (patent infringement), and 28 U.S.C. §§ 2201 and 2202 (declaratory judgment).

7. Venue is proper in this district pursuant to 28 U.S.C. §§ 1391(b) and 1400(b).

8. This Court has personal jurisdiction over each of the Defendants by virtue of the fact that, *inter alia*, they have committed — or aided, abetted, planned, contributed to, or participated in the commission of — tortious conduct which will lead to foreseeable harm and

injury to Plaintiff in the State of New York. A substantial part of the events giving rise to Plaintiff's claims occurred in this judicial district. The infringing product at issue is being sold in this judicial district.

9. Defendants maintain continuous and systematic contacts with the State of New York and this District. Defendants market and sell pharmaceutical products through the United States, including the State of New York, and regularly, systematically, and currently transact business in the Southern District of New York, at least by making and shipping into this judicial district, or by offering to sell or selling, or causing others to offer to sell or sell, pharmaceutical products. Defendants derive substantial revenue from goods used or consumed or services rendered in this judicial district.

10. Upon information and belief, Defendants currently sell significant quantities of over forty (40) different generic drug products in the Southern District of New York. Those products include, for example, generic versions of Wellbutrin XL®, Xanax®, and Cardizem® CD. A list of generic products manufactured and sold by Defendants in the United States is provided by Actavis at <http://www.actavis.us/en/products/new.htm>.

11. Based on the facts and causes alleged herein, and for additional reasons to be developed through discovery, this Court has personal jurisdiction over the Defendants.

FACTUAL BACKGROUND

Endo's Opana ER CRF NDA

12. On December 12, 2011, FDA approved Endo's Supplemental New Drug Application ("sNDA") 201655, under § 505(b) of the Federal Food, Drug and Cosmetic Act, 21 U.S.C. § 355(b), for Opana ER CRF, which is designed to be a crush-resistant tablet that contains oxymorphone hydrochloride for the relief of pain.

13. Opana ER CRF is distributed and sold throughout the United States for relief of moderate to severe pain in patients requiring continuous around-the-clock opioid treatment for an extended period of time.

THE PATENTS

14. On December 14, 2010, the PTO duly and legally issued U.S. Patent No. 7,851,482 (the '482 Patent), entitled "Method For Making Analgesics" to Johnson Matthey Public Limited Company ("Johnson Matthey") as assignee. Jen-Sen Dung, Erno M. Keskeny, and James J. Mencel are named as inventors. A true and correct copy of the '482 Patent is attached as Exhibit A.

15. Endo has acquired full title to the '482 Patent, and is now the sole owner and assignee of the '482 Patent.

16. Information regarding the Endo '482 Patent was submitted to FDA for listing in its publication, the *Approved Drug Products with Therapeutic Equivalence Evaluations*, which is referred to as the "Orange Book." See 21 U.S.C. § 355(b)(1) and (c)(2). Pursuant to 21 C.F.R. § 314.53(e), FDA has listed the '482 Patent in the Orange Book with reference to NDA 201655.

17. On November 13, 2012, the PTO duly and legally issued U.S. Patent No. 8,309,122 (the '122 Patent), entitled "Oxymorphone Controlled Release Formulations" to Endo Pharmaceuticals, Inc. as assignee. Huai-Hung Kao, Anand R. Baichwal, Troy McCall, and David Lee are named as inventors. A true and correct copy of the '122 Patent is attached as Exhibit B.

18. Endo is the sole owner and assignee of the '122 Patent.

19. Information regarding the Endo '122 Patent has been submitted to FDA for listing in the Orange Book.

20. On December 11, 2012, the PTO duly and legally issued U.S. Patent No. 8,329,216 (the '216 Patent), entitled "Oxymorphone Controlled Release Formulations" to Endo Pharmaceuticals, Inc. as assignee. Huai-Hung Kao, Anand R. Baichwal, Troy McCall, and David Lee are named as inventors. A true and correct copy of the '216 Patent is attached as Exhibit C.

21. Endo is the sole owner and assignee of the '216 Patent.

22. Information regarding the Endo '216 Patent has been submitted to FDA for listing in the Orange Book.

23. Opana ER CRF is covered by one or more claims of each of the '482, '122 and '216 Patents.

DEFENDANTS' INFRINGING PRODUCT

24. On or about February 2008, ASA filed Abbreviated New Drug Application ("ANDA") No. 79-046 seeking approval to engage in the commercial manufacturing, use and sale of the Actavis's Generic Oxymorphone ER Tablets as a generic version of the original, non-crush-resistant formulation of Opana® ER (the "Discontinued Formulation").

25. In response, Endo filed suit against ASA for infringement of U.S. Patent No. 5,958,456 ("456 patent"). *See Endo Pharmaceuticals Inc., et. al v. Actavis South Atlantic LLC*, United States District Court, District of New Jersey, Dkt. Nos. 2:08-cv-03482-KSH-PS and 2:08-cv-01563-KSH-PS. Endo and ASA settled their infringement dispute in February 2009.

26. Although the parties' settlement agreement granted Actavis a license under the '456 patent to make and sell its Generic Oxymorphone ER Tablets, nothing in the agreement grants Defendants any license or other rights under the '482, '122 or '216 Patents.

27. Defendants currently make and sell 7.5 mg and 15 mg strengths of the Actavis's

Generic Oxymorphone ER Tablets.

28. Defendants' manufacture and sale of the Actavis Generic Oxymorphone ER Tablets has caused Endo to suffer harm, including without limitation, irreparable injury to its business reputation and goodwill, lost sales of Opana® ER CRF, the loss of the benefit of its investment in developing the reformulated crush-resistant version of Opana® ER, and price erosion for Opana® ER CRF.

COUNT I: INFRINGEMENT OF THE '482 PATENT

29. Endo incorporates each of paragraphs 1-28 above as if set forth fully herein.

30. Defendants' commercial manufacture, sale, and offer for sale of the Actavis Generic Oxymorphone ER Tablets constitutes an infringement of the '482 Patent under 35 U.S.C. § 271(a)-(c).

31. Upon information and belief, Defendants are aware of the existence of the '482 Patent, and are aware that the commercial manufacture, sale, and offer for sale of filing of the Actavis Generic Oxymorphone ER Tablets constitutes infringement of the '482 Patent. Defendants' infringement is willful.

COUNT II: INFRINGEMENT OF THE '122 PATENT

32. Plaintiffs incorporate each of paragraphs 1-31 above as if set forth fully herein.

33. Defendants' commercial manufacture, sale, and offer for sale of the Actavis Generic Oxymorphone ER Tablets constitutes an infringement of the '122 Patent under 35 U.S.C. § 271(a)-(c).

34. Upon information and belief, Defendants are aware of the existence of the '122 Patent, and are aware that the commercial manufacture, sale, and offer for sale of filing of the Actavis Generic Oxymorphone ER Tablets constitutes infringement of the '122 Patent.

Defendants' infringement is willful.

COUNT III: INFRINGEMENT OF THE '216 PATENT

35. Endo incorporates each of paragraphs 1-34 above as if set forth fully herein.
36. Defendants' commercial manufacture, sale, and offer for sale of the Actavis Generic Oxymorphone ER Tablets constitutes an infringement of the '216 Patent under 35 U.S.C. § 271(a)-(c).
37. Upon information and belief, Defendants were aware of the pending issuance of the '216 Patent, and were aware that the commercial manufacture, sale, and offer for sale of filing of the Actavis Generic Oxymorphone ER Tablets would constitute infringement of the patent upon the '216 Patent's issuance. Defendants' infringement is willful.

PRAYER FOR RELIEF

WHEREFORE, Plaintiffs Endo respectfully requests the following relief:

- A. A judgment that Defendants have infringed and are infringing the '482 Patent;
- B. A judgment that Defendants have infringed and are infringing the '122 Patent;
- C. A judgment that Defendants have infringed and are infringing the '216 Patent;
- D. A permanent injunction, pursuant to 35 U.S.C. § 271(e)(4)(B), restraining and enjoining Defendants, their officers, agents, servants and employees, and those persons in active concert or participation with any of them, from infringement of the '482, '122, and '216 Patents, for the full terms thereof, including any extensions;
- E. An order that damages or other monetary relief be awarded to Endo because of Defendants' engaging in the commercial manufacture, use, offer to sell, sale, distribution or importation of the Actavis Generic Oxymorphone ER Tablets, or in inducing such conduct by others, prior to the expiration of the '482, '122, and '216 Patents, and any additional period of

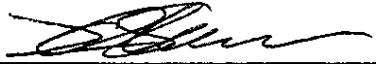
exclusivity to which Plaintiffs are or become entitled, and that any such damages or monetary relief be trebled and awarded to Endo with prejudgment interest;

F. A declaration that this is an exceptional case and an award of reasonable attorneys' fees pursuant to 35 U.S.C. § 285;

G. Reasonable attorneys' fees, filing fees, and reasonable costs of suit incurred by Endo in this action; and

H. Such other and further relief as the Court may deem just and proper.

Dated: December 11, 2012

By: 

Joshua I. Sherman
DECHERT LLP
1095 Avenue of the Americas
New York, NY 10036
(212) 698-3500
joshua.sherman@dechert.com

ATTORNEYS FOR PLAINTIFF
ENDO PHARMACEUTICALS INC.

Of Counsel:

Martin J. Black
Robert D. Rhoad
DECHERT LLP
Cira Centre
2929 Arch Street
Philadelphia, PA 19104
(215) 994-4000

Ann M. Caviani Pease
Jonathan D. Loeb
DECHERT LLP
2440 W. El Camino Real
Suite 700
Mountain View, CA 94040
(650) 813-4800

ATTORNEYS FOR PLAINTIFF
ENDO PHARMACEUTICALS INC.

Exhibit A



US007851482B2

(12) United States Patent
Dung et al.(10) Patent No.: US 7,851,482 B2
(45) Date of Patent: Dec. 14, 2010

(54) METHOD FOR MAKING ANALGESICS

6,403,798 B2	6/2002	Chiu et al.
6,723,894 B2	4/2004	Fist et al.
6,864,370 B1	3/2005	Lin et al.
6,949,645 B1	9/2005	Francis
7,071,336 B2	7/2006	Francis et al.
7,129,248 B2	10/2006	Chapman et al.
7,153,966 B2	12/2006	Casner et al.
2002/0045755 A1	4/2002	Coop et al.
2003/0129230 A1	7/2003	Baichwal et al.
2003/0129234 A1	7/2003	Baichwal et al.
2003/0157167 A1	8/2003	Kao et al.
2006/009479 A1	1/2006	Bailey et al.
2006/0173029 A1	8/2006	Chapman et al.
2008/0045716 A1	2/2008	Smith et al.
2008/0125592 A1	5/2008	Huang
2008/0312442 A1	12/2008	Buehler et al.

- (75) Inventors: **Jen-Sen Dung**, Boothwyn, PA (US); **Erno M. Keskeny**, Wilmington, DE (US); **James J. Mencel**, North Wales, PA (US)
- (73) Assignee: **Johnson Matthey Public Limited Company**, London (GB)
- (*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 646 days.

(21) Appl. No.: 11/866,840

(22) Filed: Oct. 3, 2007

(65) Prior Publication Data

US 2008/0146601 A1 Jun. 19, 2008

(30) Foreign Application Priority Data

Dec. 14, 2006 (GB) 0624880.1

FOREIGN PATENT DOCUMENTS

EP	0 359 647	3/1990
WO	WO 99/02529	1/1999
WO	WO 01/29048	4/2001
WO	WO 2005/028483	3/2005
WO	WO 2007/103105 A2 *	9/2007

(51) Int. Cl.

A61K 31/485 (2006.01)
C07D 489/04 (2006.01)(52) U.S. Cl. 514/282; 546/45; 546/44
(58) Field of Classification Search 514/282;

546/45, 44

See application file for complete search history.

(56) References Cited

U.S. PATENT DOCUMENTS

3,332,950 A	7/1967	Blumberg et al.
3,433,791 A	3/1969	Bentley
3,812,132 A	5/1974	Grew et al.
3,845,770 A	11/1974	Theeuwes et al.
3,916,899 A	11/1975	Theeuwes et al.
4,063,064 A	12/1977	Saunders et al.
4,088,864 A	5/1978	Theeuwes et al.
4,200,098 A	4/1980	Ayer et al.
4,285,987 A	8/1981	Ayer et al.
4,861,598 A	8/1989	Oshlack
4,957,681 A	9/1990	Klimesch et al.
5,071,985 A	12/1991	Andre et al.
5,215,758 A	6/1993	Krishnamurthy
5,273,760 A	12/1993	Oshlack et al.
5,286,493 A	2/1994	Oshlack et al.
5,324,351 A	6/1994	Oshlack et al.
5,356,467 A	10/1994	Oshlack et al.
5,472,712 A	12/1995	Oshlack et al.
5,869,669 A	2/1999	Huang et al.
5,922,876 A	7/1999	Huang et al.
5,948,788 A	9/1999	Huang et al.
5,952,495 A	9/1999	Huang et al.
5,981,751 A	11/1999	Mudryk et al.
6,008,354 A	12/1999	Huang et al.
6,008,355 A	12/1999	Huang et al.
6,013,796 A	1/2000	Huang et al.
6,177,567 B1	1/2001	Chiu et al.
6,262,266 B1	7/2001	Chiu et al.
6,291,675 B1	9/2001	Coop et al.
6,365,742 B1	4/2002	Mudryk et al.
6,395,900 B1	5/2002	Coop et al.

OTHER PUBLICATIONS

Andrew Coop et al., "L-Selectride as a General Reagent for the O-Demethylation and N-Decarbomethoxylation of Opium Alkaloids and Derivatives," *J. Org. Chem.*, 1998, 63 (13), pp. 4392-4396.

Ulrich Weiss, Derivatives of Morphine. II. Demethylation of 14-hydroxycodeinone. 14-Hydroxymorphinone and 8,14-Dihydroxydihydromorphinone, *J. Org. Chem.*, 1957, 22 (11), pp. 1505-1508.

U.S. Appl. No. 12/446,171 which is a National Stage of PCT/US07/68009 to Bao-Shan Huang, filed May 2, 2007 and entitled "Process for Preparing Oxymorphone".

U.S. Appl. No. 12/446,172 which is the National Stage of PCT/US07/81513 to Bao-Shan Huang, filed Oct. 16, 2007 and entitled "Process for Preparing Oxymorphone, Naltrexone, and Buprenorphine".

Andre et al., "O-Demethylation of Opioid Derivatives with Methane Sulfonic Acid / Methionine: Application to the Synthesis of Naloxone and Analogues," *Synthetic Communications*, vol. 22, No. 16, pp. 2313-2327 (1992).

Hosztai et al., "Reactions of Azodicarboxylic Esters with Amines," *Scientia Pharmaceutica*, vol. 55, pp. 61-75 (1987).

Marton et al., "Herstellung von 6, 14-Ethenomorphinan-Derivaten," *Monatshfte für Chemie*, vol. 125, pp. 1229-1239 (1994).

* cited by examiner

Primary Examiner—Charanjit S Aulakh
(74) Attorney, Agent, or Firm—RatnerPrestia

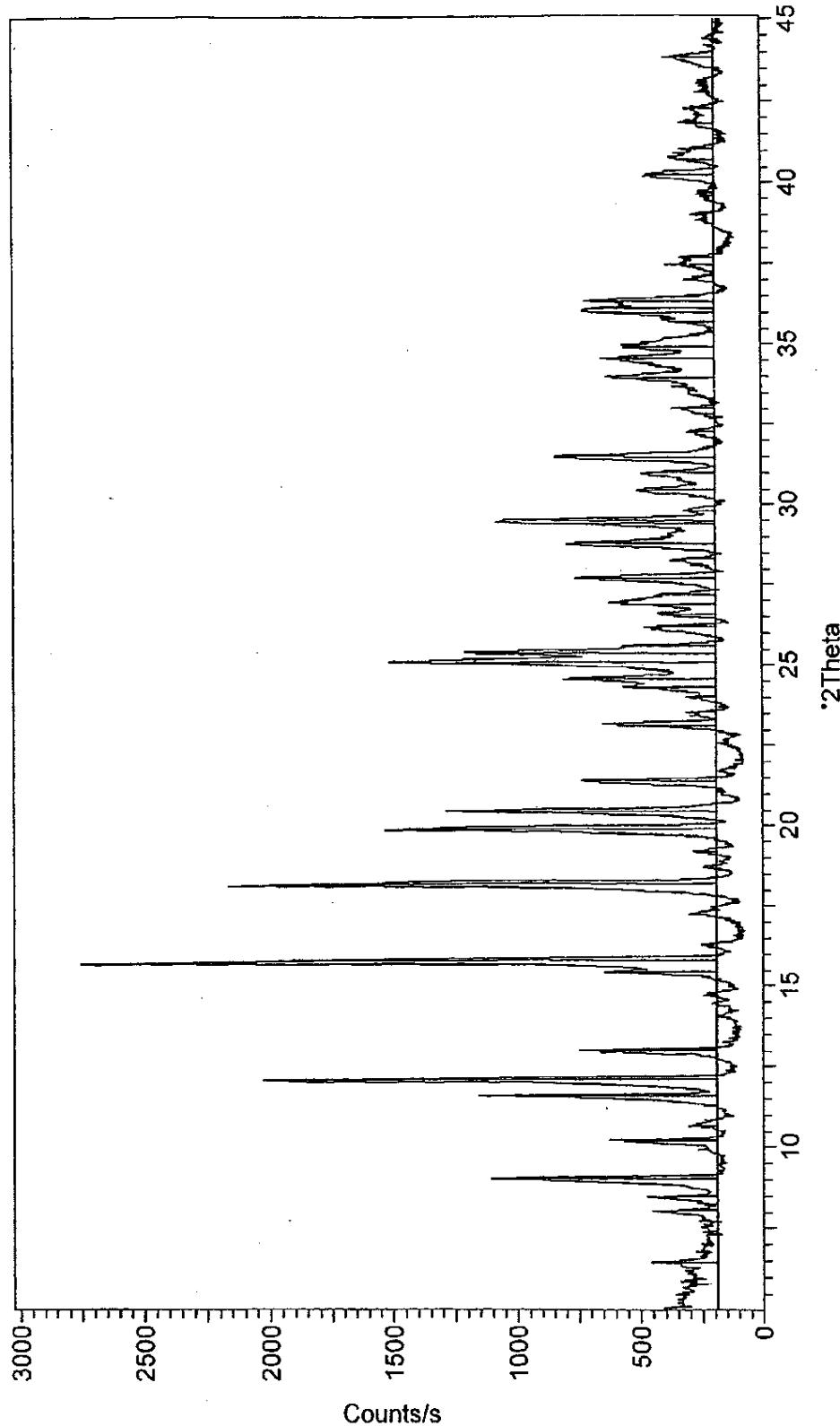
(57) ABSTRACT

Improved analgesic oxymorphone hydrochloride contains less than 10 ppm of alpha, beta unsaturated ketones and pharmaceutical preparations comprising such oxymorphone hydrochloride. The oxymorphone hydrochloride is produced by reducing a starting material oxymorphone hydrochloride using gaseous hydrogen and under specified acidity, solvent system and temperature conditions. A specific polymorph of oxymorphone hydrochloride may be obtained by hydration.

U.S. Patent

Dec. 14, 2010

US 7,851,482 B2



US 7,851,482 B2

1**METHOD FOR MAKING ANALGESICS****FIELD OF THE INVENTION**

This invention concerns an improved method for making analgesics, more especially for making the opiate oxymorphone as its hydrochloride.

BACKGROUND OF THE INVENTION

Oxymorphone, generally administered in the form of its hydrochloride salt, is a potent semi-synthetic opiate analgesic, for the relief of moderate to severe pain, and has been approved for use since 1959. It can be administered as an injectable solution, suppository, tablet or extended release tablet. It is desirable to develop high purity forms of oxymorphone and a method for its synthesis.

Several methods for synthesising oxymorphone from compounds isolated from the opium poppy or compounds derived therefrom are known, for example, starting from morphine, thebaine, or from oxycodone. There remains the need for methods which permit the formation of oxymorphone with low contamination of alpha, beta unsaturated ketones. The present invention provides an improved oxymorphone product and a method for producing such oxymorphone.

U.S. Pat. No. 7,129,248 claims a process for producing oxycodone hydrochloride with less than 25 ppm of 14-hydroxycodeinone, by hydrogenating oxycodone having greater than 100 ppm 14-hydroxycodeinone. The synthetic route to oxycodone taught in US'248 starts from thebaine and produces 14-hydroxycodeinone as an intermediate product and 8,14-dihydroxy-7,8-dihydrocodeinone as a by-product resulting from over-oxidation of thebaine. During conversion of oxycodone free base to the hydrogen chloride salt, the by-product may undergo acid-catalysed dehydration and be converted into 14-hydroxycodeinone. Thus the final oxycodone hydrogen chloride salt contains unreacted 14-hydroxycodeinone as well as 14-hydroxycodeinone derived from the by-product 8,14-dihydroxy-7,8-dihydrocodeinone. A hydrogenation step is claimed to reduce contents of 14-hydroxycodeinone from at least 100 ppm to less than 25 ppm.

SUMMARY OF THE INVENTION

The present invention provides an oxymorphone hydrochloride product containing less than 10 ppm of alpha, beta unsaturated ketones.

The invention also provides a method of purifying oxymorphone hydrochloride to yield an oxymorphone hydrochloride product containing less than 10 ppm of alpha, beta unsaturated ketones, which method comprises reducing a starting material oxymorphone hydrochloride in a strongly acid water and alcohol solvent, using gaseous hydrogen at a temperature in the range from 60 to 70° C. Reduction is suitably carried out for a period of at least 20 hours, but in another embodiment, reduction is carried out for 1 to 20 hours.

BRIEF DESCRIPTION OF THE DRAWINGS

The invention will be described below with reference to the drawing, in which:

FIG. 1 is the Powder X-Ray Diffraction pattern collected for a hydrated oxymorphone hydrochloride product made according to Example 3.2D.

DETAILED DESCRIPTION OF THE INVENTION

Preferably, the solvent is ethanol/water, although other water miscible alcohols, such as isopropanol and n-propanol,

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may be used. The reaction medium is very acidic, preferably by incorporating at least two equivalents of hydrochloric acid. A pH of less than 1 is desirable.

The reaction temperature is most preferably maintained at about 65° C. Hydrogen is conveniently supplied to the reaction vessel at 2.41 bar pressure.

The method of the invention has been able to reduce starting material oxymorphone hydrochloride having very high (of the order of 0.3 to 0.5%, or 3,000 to 5,000 ppm) content of alpha, beta unsaturated ketones to less than 10 ppm, and in many cases to undetectable levels (by HPLC).

The starting material oxymorphone hydrochloride may be an isolated or non-isolated material. Desirably, it has been obtained by the formation of the hydrogen chloride salt by heating oxymorphone free base in the presence of hydrochloric acid and an alcohol/water reaction medium. Suitable temperatures are 60-70° C. It can be seen that the reaction medium is ideal for the reduction of the method of the invention, so that it is generally not necessary to isolate the oxymorphone hydrochloride. However, the starting material oxymorphone hydrochloride may be isolated from the reaction medium or may be from another source.

The oxymorphone free base is itself preferably prepared by a reduction of 14-hydroxymorphinone. This may be carried out in a single- or two-stage process. The reduction is preferably carried out in acetic acid using gaseous hydrogen and a palladium on carbon catalyst. Preferred temperatures are of the order of 30° C. The base is precipitated by adding aqueous ammonia (NH₄OH).

This reduction may be in the presence of the reaction medium to which is added dichloromethane in methanol, Florasil and n-propanol.

The 14-hydroxymorphinone itself is most suitably prepared by hydroxylation of oripavine, using hydrogen peroxide in the presence of formic acid.

Oripavine is a known compound, which is extractable from poppy straw. The strain developed in Tasmania to be a high-Thebaine-yielding strain also produces higher than normal levels of oripavine.

The process of the invention is highly flexible, permitting many reaction steps to be carried out without isolation of intermediate products, whilst still retaining high (of the order of 50%) overall yields from oripavine, as well as remarkably high purity. Under favourable conditions, the presence of alpha, beta unsaturated ketones is undetectable by conventional means such as HPLC, but the skilled person can readily achieve less than 10 ppm contamination. The process of the invention has been successfully carried out at kilogram scale.

The oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones can be incorporated into pharmaceutical dosage forms, e.g., by admixtures of the oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones with conventional excipients, i.e., pharmaceutically acceptable organic or inorganic carrier substances. For oral formulations, the dosage forms can provide a sustained release of the active component. Suitable pharmaceutically acceptable carriers include but are not limited to, alcohols, gum arabic, vegetable oils, benzyl alcohols, polyethylene glycols, gelate, carbohydrates such as lactose, amylose or starch, magnesium stearate, talc, silicic acid, viscous paraffin, perfume oil, fatty acid monoglycerides and diglycerides, pentaerythritol fatty acid esters, hydroxy-methylcellulose, polyvinylpyrrolidone, etc. The pharmaceutical preparations can be sterilized and if desired mixed with auxiliary agents, e.g., lubricants, disintegrants, preservatives, stabilizers, wetting agents, emulsifiers, salts for influencing osmotic pressure buffers, colouring, flavouring and/or aro-

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matic substances and the like. The compositions intended for oral use may be prepared according to any method known in the art and such compositions may contain one or more agents selected from the group consisting of inert, non-toxic pharmaceutically acceptable excipients that are suitable for the manufacture of tablets. Such excipients include, for example an inert diluent such as lactose; granulating and disintegrating agents such as cornstarch; binding agents such as starch; and lubricating agents such as magnesium stearate. The tablets may be uncoated or they may be coated by known techniques for elegance or to delay release of the active ingredients. Formulations for oral use may also be presented as hard gelatin capsules wherein the active ingredient is mixed with an inert diluent. The oral dosage forms of the present invention may be in the form of tablets (sustained release and/or immediate release), troches, lozenges, powders or granules, hard or soft capsules, microparticles (e.g., microcapsules, microspheres and the like), buccal tablets, solutions, suspensions, etc.

In certain embodiments, the present invention provides for a method of treating pain by administering to a human patient the dosage forms described herein.

When the dosage form is oral, the dosage form of the present invention contains from about 1 mg to about 40 mg of oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones. Particularly preferred dosages are about 5 mg, about 10 mg, about 20 mg or about 40 mg however other dosages may be used as well. The oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones can also be formulated with suitable pharmaceutically acceptable excipients to provide a sustained release of having less than 10 ppm of alpha, beta unsaturated ketones. Such formulations can be prepared in accordance with US 2003/129230 A1, US 2003/129234 A1 and US 2003/157167 A1.

The oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones can be formulated as a sustained release oral formulation in any suitable tablet, coated tablet or multiparticulate formulation known to those skilled in the art. The sustained release dosage form may include a sustained release material that is incorporated into a matrix along with the oxymorphone salt thereof.

The sustained release dosage form may optionally comprise particles containing oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones. In certain embodiments, the particles have a diameter from about 0.1 mm to about 2.5 mm, preferably from about 0.5 mm to about 2 mm. Preferably, the particles are film coated with a material that permits release of the active at a sustained rate in an aqueous medium. The film coat is chosen so as to achieve, in combination with the other stated properties, desired release properties. The sustained release coating formulations of the present invention should preferably be capable of producing a strong, continuous film that is smooth and elegant, capable of supporting pigments and other coating additives, non-toxic, inert, and tack-free.

Coated Beads

In certain embodiments of the present invention a hydrophobic material is used to coat inert pharmaceutical beads such as mu pariel 18/20 beads, and a plurality of the resultant solid sustained release beads may thereafter be placed in a gelatin capsule in an amount sufficient to provide an effective sustained release dose when ingested and contacted by an environmental fluid, e.g., gastric fluid or dissolution media.

The sustained release bead formulations of the present invention slowly release the active component of the present

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invention, e.g., when ingested and exposed to gastric fluids, and then to intestinal fluids. The sustained release profile of the formulations of the invention can be altered, for example, by varying the amount of overcoating with the hydrophobic material, altering the manner in which a plasticiser is added to the hydrophobic material, by varying the amount of plasticiser relative to hydrophobic material, by the inclusion of additional ingredients or excipients, by altering the method of manufacture, etc. The dissolution profile of the ultimate product may also be modified, for example, by increasing or decreasing the thickness of the retardant coating.

Spheroids or beads coated with the agent(s) of the present are prepared, e.g., by dissolving the agent(s) in water and then spraying the solution onto a substrate, for example, mu pariel 18/20 beads, using a Wurster insert. Optionally, additional ingredients are also added prior to coating the beads in order to assist the binding of the active to the beads, and/or to color the solution, etc. For example, a product that includes hydroxypropylmethylcellulose, etc with or without colorant (e.g., Opadry™, commercially available from Colorcon, Inc.) may be added to the solution and the solution mixed (e.g., for about 1 hour) prior to application of the same onto the beads. The resultant coated substrate, in these example beads, may then be optionally overcoated with a barrier agent, to separate the active component(s) from the hydrophobic sustained release coating. An example of a suitable barrier agent is one which comprises hydroxypropylmethylcellulose. However, any film-former known in the art may be used. It is preferred that the barrier agent does not affect the dissolution rate of the final product.

The beads may then be overcoated with an aqueous dispersion of the hydrophobic material. The aqueous dispersion of hydrophobic material preferably further includes an effective amount of plasticiser, e.g. triethyl citrate. Pre-formulated aqueous dispersions of ethylcellulose, such as Aquacoat™ or Surelease™, may be used. If Surelease™ is used, it is not necessary to separately add a plasticiser. Alternatively, pre-formulated aqueous dispersions of acrylic polymers such as Eudragit™ can be used.

The coating solutions of the present invention preferably contain, in addition to the film-former, plasticiser, and solvent system (i.e., water), a colorant to provide elegance and product distinction. Colour may be added to the solution of the therapeutically active agent instead, or in addition to the aqueous dispersion of hydrophobic material. For example, colour may be added to Aquacoat™ via the use of alcohol or propylene glycol based colour dispersions, milled aluminium lakes and opacifiers such as titanium dioxide by adding colour with shear to water soluble polymer solution and then using low shear to the plasticised Aquacoat™. Alternatively, any suitable method of providing colour to the formulations of the present invention may be used. Suitable ingredients for providing colour to the formulation when an aqueous dispersion of an acrylic polymer is used include titanium dioxide and colour pigments, such as iron oxide pigments. The incorporation of pigments, may, however, increase the retard effect of the coating.

Plasticised hydrophobic material may be applied onto the substrate comprising the agent(s) by spraying using any suitable spray equipment known in the art. In a preferred method, a Wurster fluidised-bed system is used in which an air jet, injected from underneath, fluidizes the core material and effects drying while the acrylic polymer coating is sprayed on. A sufficient amount of the hydrophobic material to obtain a predetermined sustained release of the agent(s) when the coated substrate is exposed to aqueous solutions, e.g. gastric fluid, may be applied. After coating with the hydrophobic

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material, a further overcoat of a film-former, such as Opadry™, is optionally applied to the beads. This overcoat is provided, if at all, in order to substantially reduce agglomeration of the beads.

The release of the agent(s) from the sustained release formulation of the present invention can be further influenced, i.e., adjusted to a desired rate, by the addition of one or more release-modifying agents, or by providing one or more passageways through the coating. The ratio of hydrophobic material to water soluble material is determined by, among other factors, the release rate required and the solubility characteristics of the materials selected.

The release-modifying agents, which function as pore-formers may be organic or inorganic, and include materials that can be dissolved, extracted or leached from the coating in an environment of use. The pore-formers may comprise one or more hydrophilic materials such as hydroxypropylmethylcellulose.

The sustained release coatings of the present invention can also include erosion-promoting agents such as starch and gums.

The sustained release coatings of the present invention can also include materials useful for making microporous lamina in the environment of use, such as polycarbonates comprised of linear polyesters of carbonic acid in which carbonate groups reoccur in the polymer chain.

The release-modifying agent may also comprise a semi-permeable polymer.

In certain preferred embodiments, the release-modifying agent is selected from hydroxypropylmethylcellulose, lactose, metal stearates, and mixtures of any of the foregoing.

The sustained release coatings of the present invention may also include an exit means comprising at least one passageway, orifice, or the like. The passageway may be formed by such methods as those disclosed in U.S. Pat. No. 3,845,770, U.S. Pat. No. 3,916,899, U.S. Pat. No. 4,063,064 and U.S. Pat. No. 4,088,864.

Matrix Formulations

In other embodiments of the present invention, the sustained release formulation is achieved via a matrix optionally having a sustained release coating as set forth herein. The materials suitable for inclusion in a sustained release matrix may depend on the method used to form the matrix.

For example, a matrix in addition to the oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones may include: hydrophilic and/or hydrophobic materials, such as gums, cellulose ethers, acrylic resins, protein derived materials. The list is not meant to be exclusive, any pharmaceutically acceptable hydrophobic material or hydrophilic material which is capable of imparting sustained release of the agent(s) and which melts (or softens to the extent necessary to be extruded) may be used in accordance with the present invention.

Digestible, long chain (C_8-C_{50} , especially $C_{12}-C_{40}$), substituted or unsubstituted hydrocarbons, such as fatty acids, fatty alcohols, glyceryl esters of fatty acids, mineral and vegetable oils and waxes, and stearyl alcohol; and polyalkylene glycols. Of these polymers, acrylic polymers, especially Eudragit™ RSPO—the cellulose ethers, especially hydroxyalkylcelluloses and carboxyalkylcelluloses, are preferred. The oral dosage form may contain between 1% and 80% (by weight) of at least one hydrophilic or hydrophobic material.

When the hydrophobic material is a hydrocarbon, the hydrocarbon preferably has a melting point of between 25° C. and 90° C. Of the long chain hydrocarbon materials, fatty

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(aliphatic) alcohols are preferred. The oral dosage form may contain up to 60% (by weight) of at least one digestible, long chain hydrocarbon.

Preferably, the oral dosage form contains up to 60% (by weight) of at least one polyalkylene glycol.

The hydrophobic material is preferably selected from the group consisting of alkylcelluloses, acrylic and methacrylic acid polymers and copolymers, shellac, zein, hydrogenated castor oil, hydrogenated vegetable oil, or mixtures thereof. In certain preferred embodiments of the present invention, the hydrophobic material is a pharmaceutically acceptable acrylic polymer, including but not limited to acrylic acid and methacrylic acid copolymers, methyl methacrylate, methyl methacrylate copolymers, ethoxyethyl methacrylates, cyanoethyl methacrylate, aminoalkyl methacrylate copolymer, poly(acrylic acid), poly(methacrylic acid), methacrylic acid alkylamine copolymer, poly(methyl methacrylate), poly(methacrylic acid) (anhydride), polymethacrylate, polyacrylamide, poly(methacrylic acid anhydride), and glycidyl methacrylate copolymers. In other embodiments, the hydrophobic material is selected from materials such as hydroxyalkylcelluloses such as hydroxypropylmethylcellulose and mixtures of the foregoing.

Preferred hydrophobic materials are water-insoluble with more or less pronounced hydrophilic and/or hydrophobic trends. Preferably, the hydrophobic materials useful in the invention have a melting point from about 25° C. to about 200° C., preferably from about 45° C. to about 90° C. Specifically, the hydrophobic material may comprise natural or synthetic waxes, fatty alcohols (such as lauryl, myristyl, stearyl, cetyl or preferably cetostearyl alcohol), fatty acids, including but not limited to fatty acid esters, fatty acid glycerides (mono-, di-, and tri-glycerides), hydrogenated fats, hydrocarbons, normal waxes, stearic acid, stearyl alcohol and hydrophobic and hydrophilic materials having hydrocarbon backbones. Suitable waxes include, for example, beeswax, glycowax, castor wax and carnauba wax. For the purposes of the present invention, a wax-like substance is defined as any material that is normally solid at room temperature and has a melting point of from about 25° C. to about 100° C.

Suitable hydrophobic materials which may be used in accordance with the present invention include digestible, long chain (C_8-C_{50} , especially $C_{12}-C_{40}$), substituted or unsubstituted hydrocarbons, such as fatty acids, fatty alcohols, glyceryl esters of fatty acids, mineral and vegetable oils and natural and synthetic waxes. Hydrocarbons having a melting point of between 25° C. and 90° C. are preferred. Of the long chain hydrocarbon materials, fatty (aliphatic) alcohols are preferred in certain embodiments. The oral dosage form may contain up to 60% (by weight) of at least one digestible, long chain hydrocarbon. Preferably, a combination of two or more hydrophobic materials are included in the matrix formulations. If an additional hydrophobic material is included, it is preferably selected from natural and synthetic waxes, fatty acids, fatty alcohols, and mixtures of the same. Examples include beeswax, carnauba wax, stearic acid and stearyl alcohol. This list is not meant to be exclusive.

One particular suitable matrix comprises at least one water soluble hydroxyalkyl cellulose, at least one $C_{12}-C_{36}$, preferably $C_{14}-C_{22}$, aliphatic alcohol and, optionally, at least one polyalkylene glycol. The at least one hydroxyalkyl cellulose is preferably a hydroxy (C_1 to C_6) alkyl cellulose, such as hydroxypropylcellulose, hydroxypropyl-methylcellulose and, especially, hydroxyethylcellulose. The amount of the at least one hydroxyalkyl cellulose in the present oral dosage form will be determined, inter alia, by the precise rate of oxymorphone hydrochloride release required. The at least

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one aliphatic alcohol may be, for example, lauryl alcohol, myristyl alcohol or stearyl alcohol. In particularly preferred embodiments of the present oral dosage form, however, the at least one aliphatic alcohol is cetyl alcohol or cetostearyl alcohol. The amount of the at least one aliphatic alcohol in the present oral dosage form will be determined, as above, by the precise rate of opioidoxymorphone release required. It will also depend on whether at least one polyalkylene glycol is present in or absent from the oral dosage form. In the absence of at least one polyalkylene glycol, the oral dosage form preferably contains between 20% and 50% (by wt) of the at least one aliphatic alcohol. When at least one polyalkylene glycol is present in the oral dosage form, then the combined weight of the at least one aliphatic alcohol and the at least one polyalkylene glycol preferably constitutes between 20% and 50% (by wt) of the total dosage.

In one embodiment, the ratio of, e.g., the at least one hydroxyalkyl cellulose or acrylic resin to the at least one aliphatic alcohol/polyalkylene glycol determines, to a (w/w) of the at least one hydroxyalkyl cellulose to the at least one aliphatic alcohol/polyalkylene glycol of between 1:2 and 1:4 is preferred, with a ratio of between 1:3 and 1:4 being particularly preferred.

The at least one polyalkylene glycol may be, for example, polypropylene glycol or, preferably, polyethylene glycol. The number average molecular weight of the at least one polyalkylene glycol is preferably between 1,000 and 15,000 especially between 1,500 and 12,000.

Another suitable sustained release matrix would comprise an alkylcellulose (especially ethyl cellulose), a C₁₂ to C₃₆ aliphatic alcohol and, optionally, a polyalkylene glycol.

In another preferred embodiment, the matrix includes a pharmaceutically acceptable combination of at least two hydrophobic materials.

In addition to the above ingredients, a sustained release matrix may also contain suitable quantities of other materials, e.g. diluents, lubricants, binders, granulating aids, colorants, flavorants and glidants that are conventional in the pharmaceutical art.

Matrix—Particulates

In order to facilitate the preparation of a solid, sustained release, oral dosage form according to this invention, any method of preparing a matrix formulation known to those skilled in the art may be used. For example incorporation in the matrix may be effected, for example, by (a) forming granules comprising at least one water soluble hydroxyalkyl cellulose, and the oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones; (b) mixing the hydroxyalkyl cellulose containing granules with at least one C₁₂ to C₃₆ aliphatic alcohol; and (c) optionally, compressing and shaping the granules. Preferably, the granules are formed by wet granulating the hydroxalkyl cellulose granules with water.

In yet other alternative embodiments, a spheronizing agent, together with the active component can be spheronized to form spheroids. Microcrystalline cellulose is a preferred spheronizing agent. A suitable microcrystalline cellulose is, for example, the material sold as Avicel PH 101 (Trade Mark, FMC Corporation). In such embodiments, in addition to the active ingredient and spheronizing agent, the spheroids may also contain a binder. Suitable binders, such as low viscosity, water soluble polymers, will be well known to those skilled in the pharmaceutical art. However, water soluble hydroxy lower alkyl cellulose, such as hydroxypropyl-cellulose, are preferred. Additionally (or alternatively) the spheroids may contain a water insoluble polymer, especially an acrylic poly-

mer, an acrylic copolymer, such as a methacrylic acid-ethyl acrylate copolymer, or ethyl cellulose. In such embodiments, the sustained release coating will generally include a hydrophobic material such as (a) a wax, either alone or in admixture with a fatty alcohol; or (b) shellac or zein.

Melt Extrusion Matrix

Sustained release matrices can also be prepared via melt-granulation or melt-extrusion techniques. Generally, melt-granulation techniques involve melting a normally solid hydrophobic material, e.g. a wax, and incorporating a powdered drug therein. To obtain a sustained release dosage form, it may be necessary to incorporate an additional hydrophobic substance, e.g. ethylcellulose or a water-insoluble acrylic polymer, into the molten wax hydrophobic material. Examples of sustained release formulations prepared via melt-granulation techniques are found in U.S. Pat. No. 4,861,598.

The additional hydrophobic material may comprise one or more water-insoluble wax-like thermoplastic substances possibly mixed with one or more wax-like thermoplastic substances being less hydrophobic than said one or more water-insoluble wax-like substances. In order to achieve constant release, the individual wax-like substances in the formulation should be substantially non-degradable and insoluble in gastrointestinal fluids during the initial release phases. Useful water-insoluble wax-like substances may be those with a water-solubility that is lower than about 1:5,000 (w/w).

In addition to the above ingredients, a sustained release matrix may also contain suitable quantities of other materials, e.g., diluents, lubricants, binders, granulating aids, colorants, flavorants and glidants that are conventional in the pharmaceutical art. The quantities of these additional materials will be sufficient to provide the desired effect to the desired formulation.

In addition to the above ingredients, a sustained release matrix incorporating melt-extruded multiparticulates may also contain suitable quantities of other materials, e.g. diluents, lubricants, binders, granulating aids, colourants, flavorants and glidants that are conventional in the pharmaceutical art in amounts up to about 50% by weight of the particulate if desired.

Specific examples of pharmaceutically acceptable carriers and excipients that may be used to formulate oral dosage forms are described in the Handbook of Pharmaceutical Excipients, American Pharmaceutical Association (1986).

Melt Extrusion Multiparticulates

The preparation of a suitable melt-extruded matrix according to the present invention may, for example, include the steps of blending the oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones together with at least one hydrophobic material and preferably the additional hydrophobic material to obtain a homogeneous mixture. The homogeneous mixture is then heated to a temperature sufficient to at least soften the mixture sufficiently to extrude the same. The resulting homogeneous mixture is then extruded to form strands. The extrudate is preferably cooled and cut into multiparticulates by any means known in the art. The strands are cooled and cut into multiparticulates. The multiparticulates are then divided into unit doses. The extrudate preferably has a diameter of from about 0.1 mm to about 5 mm and provides sustained release of the therapeutically active agent for a time period of from about 8 hours to about 24 hours.

An optional process for preparing the melt extrusions of the present invention includes directly metering into an extruder a hydrophobic material, the oxymorphone hydrochloride

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having less than 10 ppm of alpha, beta unsaturated ketones, and an optional binder, heating the homogenous mixture; extruding the homogenous mixture to thereby form strands; cooling the strands containing the homogeneous mixture; cutting the strands into particles having a size from about 0.1 mm to about 12 mm; and dividing said particles into unit doses. In this aspect of the invention, a relatively continuous manufacturing procedure is realized.

The diameter of the extruder aperture or exit port can also be adjusted to vary the thickness of the extruded strands. Furthermore, the exit part of the extruder need not be round; it can be oblong, rectangular, etc. The exiting strands can be reduced to particles using a hot wire cutter, guillotine, etc.

The melt extruded multiparticulate system can be, for example, in the form of granules, spheroids or pellets depending upon the extruder exit orifice. For the purposes of the present invention, the terms "melt-extruded multiparticulate(s)" and "melt-extruded multiparticulate system(s)" and "melt-extruded particles" shall refer to a plurality of units, preferably within a range of similar size and/or shape and containing one or more active agents and one or more excipients, preferably including a hydrophobic material as described herein. In this regard, the melt-extruded multiparticulates will be of a range of from about 0.1 mm to about 12 mm in length and have a diameter of from about 0.1 mm to about 5 mm. In addition, it is to be understood that the melt-extruded multiparticulates can be any geometrical shape within this size range. Alternatively, the extrudate may simply be cut into desired lengths and divided into unit doses of the therapeutically active agent without the need of a spheronization step.

In one preferred embodiment, oral dosage forms are prepared to include an effective amount of melt-extruded multiparticulates within a capsule. For example, a plurality of the melt-extruded multiparticulates may be placed in a gelatin capsule in an amount sufficient to provide an effective sustained release dose when ingested and contacted by gastric fluid.

In another preferred embodiment, a suitable amount of the multiparticulate extrudate is compressed into an oral tablet using conventional tabletting equipment using standard techniques. Techniques and compositions for making tablets (compressed and moulded), capsules (hard and soft gelatin) and pills are also described in Remington's Pharmaceutical Sciences, (Arthur Osol, editor), 1553-1593 (1980).

In yet another preferred embodiment, the extrudate can be shaped into tablets as set forth in U.S. Pat. No. 4,957,681, described in additional detail above.

Optionally, the sustained release melt-extruded multiparticulate systems or tablets can be coated, or the gelatin capsule containing the multiparticulates can be further coated, with a sustained release coating such as the sustained release coatings described above. Such coatings preferably include a sufficient amount of hydrophobic material to obtain a weight gain level from about 2% to about 30%, although the overcoat may be greater depending upon the desired release rate, among other things.

The melt-extruded unit dosage forms of the present invention may further include combinations of melt-extruded particles before being encapsulated. Furthermore, the unit dosage forms can also include an amount of an immediate release agent for prompt release. The immediate release agent may be incorporated, e.g., as separate pellets within a gelatin capsule, or may be coated on the surface of the multiparticulates after preparation of the dosage forms (e.g., sustained release coating or matrix-based). The unit dosage forms of the present

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invention may also contain a combination of sustained release beads and matrix multiparticulates to achieve a desired effect.

The sustained release formulations of the present invention preferably slowly release the agent(s), e.g. when ingested and exposed to gastric fluids, and then to intestinal fluids. The sustained release profile of the melt-extruded formulations of the invention can be altered, for example, by varying the amount of retardant, i.e., hydrophobic material, by varying the amount of plasticiser relative to hydrophobic material, by the inclusion of additional ingredients or excipients, by altering the method of manufacture, etc.

In other embodiments of the invention, the melt extruded material is prepared without the inclusion of the oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones, which can be added thereafter to the extrudate. Such formulations typically will have the agents blended together with the extruded matrix material, and then the mixture would be tableted in order to provide a slow release formulation.

20 Coatings

The dosage forms of the present invention may optionally be coated with one or more materials suitable for the regulation of release or for the protection of the formulation. In one embodiment, coatings are provided to permit either pH-dependent or pH-independent release. A pH-dependent coating serves to release the active in desired areas of the gastro-intestinal (GI) tract, e.g. the stomach or small intestine, such that an absorption profile is provided which is capable of providing at least about eight hours and preferably about twelve hours to up to about twenty-four hours of analgesia to a patient. When a pH-independent coating is desired, the coating is designed to achieve optimal release regardless of pH-changes in the environmental fluid, e.g., the GI tract. It is also possible to formulate compositions that release a portion of the dose in one desired area of the GI tract, e.g., the stomach, and release the remainder of the dose in another area of the GI tract, e.g., the small intestine.

Formulations according to the invention that utilize pH-dependent coatings to obtain formulations may also impart a repeat-action effect whereby unprotected drug is coated over the enteric coat and is released in the stomach, while the remainder, being protected by the enteric coating, is released further down the gastrointestinal tract. Coatings which are pH-dependent may be used in accordance with the present invention include shellac, cellulose acetate phthalate (CAP), polyvinyl acetate phthalate (PVAP), hydroxypropylmethylcellulose phthalate, and methacrylic acid ester copolymers, zein, and the like.

In certain preferred embodiments, the substrate (e.g., tablet core bead, matrix particle) containing the oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones thereof is coated with a hydrophobic material selected from (i) an alkylcellulose; (ii) an acrylic polymer; or (iii) mixtures thereof. The coating may be applied in the form of an organic or aqueous solution or dispersion. The coating may be applied to obtain a weight gain from about 2% to about 25% of the substrate in order to obtain a desired sustained release profile. Coatings derived from aqueous dispersions are described in detail U.S. Pat. No. 5,273,760, U.S. Pat. No. 5,286,493, U.S. Pat. No. 5,324,351, U.S. Pat. No. 5,356,467, and U.S. Pat. No. 5,472,712.

Alkylcellulose Polymers

Cellulosic materials and polymers, including alkylcelluloses, provide hydrophobic materials well suited for coating the beads according to the invention. Simply by way of example, one preferred alkylcellulosic polymer is ethylcellulose,

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although the artisan will appreciate that other cellulose and/or alkylcellulose polymers may be readily employed, singly or in any combination, as all or part of a hydrophobic coating according to the invention.

Acrylic Polymers

In other preferred embodiments of the present invention, the hydrophobic material comprising the sustained release coating is a pharmaceutically acceptable acrylic polymer, including but not limited to acrylic acid and methacrylic acid copolymers, methyl methacrylate copolymers, ethoxyethyl methacrylates, cyanoethyl methacrylate, poly(acrylic acid), poly(methacrylic acid), methacrylic acid alkylamide copolymer, poly(methyl methacrylate), polymethacrylate, poly(methyl methacrylate) copolymer, polyacrylamide, aminoalkyl methacrylate copolymer, poly(methacrylic acid anhydride), and glycidyl methacrylate copolymers.

In certain preferred embodiments, the acrylic polymer is comprised of one or more ammonio methacrylate copolymers. Ammonio methacrylate copolymers are well known in the art, and are described as fully polymerised copolymers of acrylic and methacrylic acid esters with a low content of quaternary ammonium groups.

In order to obtain a desirable dissolution profile, it may be necessary to incorporate two or more ammonio methacrylate copolymers having differing physical properties, such as different molar ratios of the quaternary ammonium groups to the neutral (meth)acrylic esters.

Certain methacrylic acid ester-type polymers are useful for preparing pH-dependent coatings, which may be used in accordance with the present invention. For example, there are a family of copolymers synthesized from diethylaminoethyl methacrylate and other neutral methacrylic esters, also known as methacrylic acid copolymer or polymeric methacrylates, commercially available as EudragitTM from Rohm Tech, Inc. There are several different types of EudragitTM, for example EudragitTM E is an example of a methacrylic acid copolymer that swells and dissolves in acidic media. EudragitTM L is a methacrylic acid copolymer which does not swell at about pH<5.7 and is soluble at about pH>6. EudragitTM S does not swell at about pH<6.5 and is soluble at about pH>7. EudragitTM RL and EudragitTM RS are water swellable, and the amount of water absorbed by these polymers is pH-dependent, however, dosage forms coated with EudragitTM RL and RS are pH-independent.

In certain preferred embodiments, the acrylic coating comprises a mixture of two acrylic resin lacquers commercially available from Rohm Pharma under the Tradenames EudragitTM RL30D and EudragitTM RS30D, respectively. EudragitTM RL30D and EudragitTM RS30D are copolymers of acrylic and methacrylic esters with a low content of quaternary ammonium groups, the molar ratio of ammonium groups to the remaining neutral (meth)acrylic esters being 1:20 in EudragitTM RL30D and 1:40 in EudragitTM RS30D. The mean molecular weight is about 150,000. The code designations RL (high permeability) and RS (low permeability) refer to the permeability properties of these agents. EudragitTM RL/RS mixtures are insoluble in water and in digestive fluids. However, coatings formed from the same are swellable and permeable in aqueous solutions and digestive fluids.

The EudragitTM RL/RS dispersions of the present invention may be mixed together in any desired ratio in order to ultimately obtain a sustained release formulation having a desirable dissolution profile. Desirable sustained release formulations may be obtained, for instance, from a retardant coating derived from 100% EudragitTM RL, 50% EudragitTM RL and

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50% EudragitTM RS, or 10% EudragitTM RL and 90% EudragitTM RS. Of course, one skilled in the art will recognize that other acrylic polymers may also be used, such as, for example, EudragitTM L.

Plasticizers

In embodiments of the present invention where the coating comprises an aqueous dispersion of a hydrophobic material, the inclusion of an effective amount of a plasticiser in the aqueous dispersion of hydrophobic material will further improve the physical properties of the sustained release coating. For example, because ethyl-cellulose has a relatively high glass transition temperature and does not form flexible films under normal coating conditions, it is preferable to incorporate a plasticiser into an ethylcellulose coating containing sustained release coating before using the same as a coating material. Generally, the amount of plasticiser included in a coating solution is based on the concentration of the film-former, e.g., most often from about 1 wt % to about 50 wt % of the film-former. Concentration of the plasticiser, however, can only be properly determined after careful experimentation with the particular coating solution and method of application.

Examples of suitable plasticizers for ethylcellulose include water insoluble plasticizers such as dibutyl sebacate, diethyl phthalate, triethyl citrate, tributyl citrate, and triacetin, although it is possible that other water-insoluble plasticizers (such as acetylated monoglycerides, phthalate esters, castor oil, etc.) may be used. Triethyl citrate is an especially preferred plasticiser for the aqueous dispersions of ethyl cellulose of the present invention.

Examples of suitable plasticizers for the acrylic polymers of the present invention include, but are not limited to citric acid esters such as triethyl citrate, tributyl citrate, dibutyl phthalate, and possibly 1,2-propylene glycol. Other plasticizers that have proved to be suitable for enhancing the elasticity of the films formed from acrylic films such as EudragitTM RL/RS lacquer solutions include polyethylene glycols, propylene glycol, diethyl phthalate, castor oil, and triacetin. Triethyl citrate is an especially preferred plasticiser for the aqueous dispersions of ethyl cellulose of the present invention.

The addition of a small amount of talc may also help reduce the tendency of the aqueous dispersion to stick during processing, and may act as a polishing agent.

45 Sustained Release Osmotic Dosage Form

Sustained release dosage forms according to the present invention may also be prepared as osmotic dosage formulations. The osmotic dosage forms preferably include a bilayer core comprising a drug layer (containing the oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones) and a delivery or push layer, wherein the bilayer core is surrounded by a semipermeable wall and optionally having at least one passageway disposed therein.

The expression "passageway" as used for the purpose of this invention, includes aperture, orifice, bore, pore, porous element through which oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones can be pumped, diffuse or migrate through a fibre, capillary tube, porous overlay, porous insert, microporous member, or porous composition. The passageway can also include a compound that erodes or is leached from the wall in the fluid environment of use to produce at least one passageway. Representative compounds for forming a passageway include erodible poly(glycolic) acid, or poly(lactic) acid in the wall; a gelatinous filament; a water-removable poly(vinyl alcohol); leachable compounds such as fluid-removable pore-forming polysaccharides, acids, salts or oxides. A passageway can be

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formed by leaching a compound from the wall, such as sorbitol, sucrose, lactose, maltose, or fructose, to form a sustained-release dimensional pore-passageway. The dosage form can be manufactured with one or more passageways in spaced-apart relation on one or more surfaces of the dosage form. A passageway and equipment for forming a passageway are disclosed in U.S. Pat. No. 3,845,770, U.S. Pat. No. 3,916,899, U.S. Pat. No. 4,063,064 and U.S. Pat. No. 4,088,864. Passageways comprising sustained-release dimensions sized, shaped and adapted as a releasing-pore formed by aqueous leaching to provide a releasing-pore of a sustained-release rate are disclosed in U.S. Pat. No. 4,200,098 and U.S. Pat. No. 4,285,987.

In certain embodiments the drug layer may also comprise at least one polymer hydrogel. The polymer hydrogel may have an average molecular weight of between about 500 and about 6,000,000. Examples of polymer hydrogels include but are not limited to a maltodextrin polymer comprising the formula $(C_6H_{12}O_5)_nH_2O$, wherein n is 3 to 7,500, and the maltodextrin polymer comprises a 500 to 1,250,000 number-average molecular weight; a poly(alkylene oxide) represented by, e.g., a poly(ethylene oxide) and a poly(propylene oxide) having a 50,000 to 750,000 weight-average molecular weight, and more specifically represented by a poly(ethylene oxide) of at least one of 100,000, 200,000, 300,000 or 400,000 weight-average molecular weights; an alkali carboxyalkylcellulose, wherein the alkali is sodium or potassium, the alkyl is methyl, ethyl, propyl, or butyl of 10,000 to 175,000 weight-average molecular weight; and a copolymer of ethylene-acrylic acid, including methacrylic and ethacrylic acid of 10,000 to 500,000 number-average molecular weight.

In certain embodiments of the present invention, the delivery or push layer comprises an osmopolymer. Examples of an osmopolymer include but are not limited to a member selected from the group consisting of a polyalkylene oxide and a carboxyalkylcellulose. The polyalkylene oxide possesses a 1,000,000 to 10,000,000 weight-average molecular weight. The polyalkylene oxide may be a member selected from the group consisting of polymethylene oxide, polyethylene oxide, polypropylene oxide, polyethylene oxide having a 1,000,000 average molecular weight, polyethylene oxide comprising a 5,000,000 average molecular weight, polyethylene oxide comprising a 7,000,000 average molecular weight, cross-linked polymethylene oxide possessing a 1,000,000 average molecular weight, and polypropylene oxide of 1,200,000 average molecular weight. Typical osmopolymer carboxyalkylcellulose comprises a member selected from the group consisting of alkali carboxyalkylcellulose, sodium carboxymethylcellulose, potassium carboxymethylcellulose, sodium carboxyethylcellulose, lithium carboxymethylcellulose, sodium carboxyethyl-cellulose, carboxyalkylhydroxyalkylcellulose, carboxymethylhydroxyethyl cellulose, carboxyethylhydroxyethylcellulose and carboxymethylhydroxypropylcellulose. The osmopolymers used for the displacement layer exhibit an osmotic pressure gradient across the semipermeable wall. The osmopolymers imbibe fluid into dosage form, thereby swelling and expanding as an osmotic hydrogel (also known as an osmogel), whereby they push the oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones thereof from the osmotic dosage form.

The push layer may also include one or more osmotically effective compounds also known as osmagents and as osmotically effective solutes. They imbibe an environmental fluid, for example, from the gastrointestinal tract, into dosage form and contribute to the delivery kinetics of the displacement layer. Examples of osmotically active compounds comprise a

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member selected from the group consisting of osmotic salts and osmotic carbohydrates. Examples of specific osmagents include but are not limited to sodium chloride, potassium chloride, magnesium sulphate, lithium phosphate, lithium chloride, sodium phosphate, potassium sulphate, sodium sulphate, potassium phosphate, glucose, fructose and maltose.

The push layer may optionally include a hydroxypropylalkylcellulose possessing a 9,000 to 450,000 number-average molecular weight. The hydroxypropylalkyl-cellulose is represented by a member selected from the group consisting of hydroxypropylmethylcellulose, hydroxypropylethylcellulose, hydroxypropylisopropyl cellulose, hydroxypropylbutylcellulose, and hydroxypropylpentylcellulose.

The push layer optionally may comprise a non-toxic colorant or dye. Examples of colourants or dyes include but are not limited to Food and Drug Administration Colourants (FD&C), such as FD&C No. 1 blue dye, FD&C No. 4 red dye, red ferric oxide, yellow ferric oxide, titanium dioxide, carbon black, and indigo.

The push layer may also optionally comprise an antioxidant to inhibit the oxidation of ingredients. Some examples of antioxidants include but are not limited to a member selected from the group consisting of ascorbic acid, ascorbyl palmitate, butylated hydroxyanisole, a mixture of 2 and 3 tertiary-butyl-4-hydroxyanisole, butylated hydroxytoluene, sodium isoascorbate, dihydroguaric acid, potassium sorbate, sodium bisulfate, sodium metabisulfate, sorbic acid, potassium ascorbate, vitamin E, 4-chloro-2,6-ditertiary butylphenol, alphatocopherol, and propylgallate.

In certain alternative embodiments, the dosage form comprises a homogenous core comprising oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones, a pharmaceutically acceptable polymer (e.g., polyethylene oxide), optionally a disintegrant (e.g., polyvinylpyrrolidone), optionally an absorption enhancer (e.g., a fatty acid, a surfactant, a chelating agent, a bile salt, etc). The homogenous core is surrounded by a semipermeable wall having a passageway (as defined above) for the release of the oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones.

In certain embodiments, the semipermeable wall comprises a member selected from the group consisting of a cellulose ester polymer, a cellulose ether polymer and a cellulose ester-ether polymer. Representative wall polymers comprise a member selected from the group consisting of cellulose acylate, cellulose diacylate, cellulose triacylate, cellulose acetate, cellulose diacetate, cellulose triacetate, mono-, di- and tricellulose alkenylates, and mono-, di- and tricellulose alkinylates. The poly(cellulose) used for the present invention comprises a number-average molecular weight of 20,000 to 7,500,000.

Additional semipermeable polymers for the purpose of this invention comprise acetaldehyde dimethylcellulose acetate, cellulose acetate ethylcarbamate, cellulose acetate methylcarbamate, cellulose diacetate, propylcarbamate, cellulose acetate diethylaminoacetate; semipermeable polyamide; semipermeable polyurethane; semipermeable sulfonated polystyrene; semipermeable cross-linked polymer formed by the coprecipitation of a polyanion and a polycation, semipermeable crosslinked polystyrenes, semipermeable cross-linked poly(sodium styrene sulfonate), semipermeable crosslinked poly(vinylbenzyltrimethyl ammonium chloride) and semipermeable polymers possessing a fluid permeability of 2.5×10^{-8} to 2.5×10^{-2} ($\text{cm}^2/\text{hr atm}$) expressed per atmosphere of hydrostatic or osmotic pressure difference across the semipermeable wall. Other polymers useful in the present

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invention are known in the art including those in *Handbook of Common Polymers*, Scott, J. R. and W. J. Roff, 1971, CRC Press, Cleveland, Ohio.

In certain embodiments, preferably the semipermeable wall is nontoxic, inert, and it maintains its physical and chemical integrity during the dispensing life of the drug. In certain embodiments, the dosage form comprises a binder. An example of a binder includes, but is not limited to a therapeutically acceptable vinyl polymer having a 5,000 to 350,000 viscosity-average molecular weight, represented by a member selected from the group consisting of poly-n-vinylamide, poly-n-vinylacetamide, poly(vinyl pyrrolidone), also known as poly-n-vinylpyrrolidone, poly-n-vinylcaprolactone, poly-n-vinyl-5-methyl-2-pyrrolidone, and poly-n-vinyl-pyrrolidone copolymers with a member selected from the group consisting of vinyl acetate, vinyl alcohol, vinyl chloride, vinyl fluoride, vinyl butyrate, vinyl laurate, and vinyl stearate. Other binders include for example, acacia, starch, gelatin, and hydroxypropylalkylcellulose of 9,200 to 250,000 average molecular weight.

In certain embodiments, the dosage form comprises a lubricant, which may be used during the manufacture of the dosage form to prevent sticking to die wall or punch faces. Examples of lubricants include but are not limited to magnesium stearate, sodium stearate, stearic acid, calcium stearate, magnesium oleate, oleic acid, potassium oleate, caprylic acid, sodium stearyl fumarate, and magnesium palmitate.

In certain preferred embodiments, the present invention includes a therapeutic composition comprising an amount of oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones equivalent to 10 to 40 mg oxymorphone hydrochloride, 25 mg to 500 mg of poly(alkylene oxide) having a 150,000 to 500,000 average molecular weight, 1 mg to 50 mg of polyvinylpyrrolidone having a 40,000 average molecular weight, and 0 mg to about 7.5 mg of a lubricant.

Suppositories

The sustained release formulations of the present invention may be formulated as a pharmaceutical suppository for rectal administration comprising a suitable suppository base, and oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones. Preparation of sustained release suppository formulations is described in, e.g., U.S. Pat. No. 5,215,758.

Prior to absorption, the drug must be in solution. In the case of suppositories, solution must be preceded by dissolution of the suppository base, or the melting of the base and subsequent partition of the drug from the suppository base into the rectal fluid. The absorption of the drug into the body may be altered by the suppository base. Thus, the particular suppository base to be used in conjunction with a particular drug must be chosen giving consideration to the physical properties of the drug. For example, lipid-soluble drugs will not partition readily into the rectal fluid, but drugs that are only slightly soluble in the lipid base will partition readily into the rectal fluid.

Among the different factors affecting the dissolution time (or release rate) of the drugs are the surface area of the drug substance presented to the dissolution solvent medium, the pH of the solution, the solubility of the substance in the specific solvent medium, and the driving forces of the saturation concentration of dissolved materials in the solvent medium. Generally, factors affecting the absorption of drugs from suppositories administered rectally include suppository vehicle, absorption site pH, drug pKa, degree of ionisation, and lipid solubility.

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The suppository base chosen should be compatible with the active of the present invention. Further, the suppository base is preferably non-toxic and non-irritating to mucous membranes, melts or dissolves in rectal fluids, and is stable during storage.

In certain preferred embodiments of the present invention for both water-soluble and water-insoluble drugs, the suppository base comprises a fatty acid wax selected from the group consisting of mono-, di- and triglycerides of saturated, natural fatty acids of the chain length C₁₂ to C₁₈.

In preparing the suppositories of the present invention other excipients may be used. For example, a wax may be used to form the proper shape for administration via the rectal route. This system can also be used without wax, but with the addition of diluent filled in a gelatin capsule for both rectal and oral administration.

Examples of suitable commercially available mono-, di- and triglycerides include saturated natural fatty acids of the 20 12-18 carbon atom chain sold under the trade name Novata™ (types AB, AB, B, BC, BD, BBC, E, BCF, C, D and 299), manufactured by Henkel, and Witepsol™ (types H5, H12, H15, H175, H185, H19, H32, H35, H39, H42, W25, W31, W35, W45, S55, S58, E75, E76 and E85), manufactured by 25 Dynamit Nobel.

Other pharmaceutically acceptable suppository bases may be substituted in whole or in part for the above-mentioned mono-, di- and triglycerides. The amount of base in the suppository is determined by the size (i.e. actual weight) of the dosage form, the amount of base (e.g., alginate) and drug used. Generally, the amount of suppository base is from about 30 20% to about 90% by weight of the total weight of the suppository. Preferably, the amount of suppository base in the suppository is from about 65% to about 80%, by weight of the total weight of the suppository.

Additional Embodiments

40 The oxymorphone hydrochloride having less than 10 ppm of alpha, beta unsaturated ketones may be used as a substitute for the oxymorphone hydrochloride in any existing commercial product such as, e.g., Opana™, Opana ER™ and Numorphan™. Such formulations are listed in the FDA Orange Book.

EXAMPLES

50 The invention will now be illustrated by the following examples, showing the synthesis of the high purity oxymorphone, starting from oripavine.

FIG. 1 is the Powder X-Ray Diffraction pattern collected 55 for a hydrated oxymorphone hydrochloride product made according to Example 3.2D.

Example 1.1A

Hydroxylation of Oripavine to
14-hydroxymorphinone

1 kg oripavine is added with stirring to a reaction vessel containing 2.76 kg of formic acid and 0.53 kg water, and 60 stirring is continued until the oripavine is completely dissolved, and the temperature remains in the range 20-30° C. Subsequently, 0.36 kg of 35 wt % hydrogen peroxide solution

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is added, and the reaction mixture is stirred for three hours or more, whilst maintaining the temperature in the range 20-35° C. The reaction vessel is cooled to 10° C. and 7.12 litres of dilute ammonium hydroxide is added slowly, whilst maintaining the reaction mixture below 40° C. If necessary, the pH of the reaction mixture is adjusted to the range 8 to 10, with more dilute ammonium hydroxide solution or hydrochloric acid as appropriate, and stirring is continued for 3-5 hours.

A precipitate of product 14-hydroxymorphinone is formed and filtered off. The precipitate is washed with water until colourless and then dried to a damp cake and collected for the next stage.

Example 1.1B

Formation of Oxymorphone Base

A hydrogenation vessel is charged with kg litre water and 0.73 kg acetic acid before adding 1 kg of 14-hydroxymorphinone prepared as in Example 1.1A and the mixture stirred until clear. 40 g of wet 10% Pd on carbon catalyst is added under a stream of nitrogen, and hydrogen supplied at 35-40 psi (2.41-2.76 bar). The temperature is maintained at 30±5° C. until hydrogen uptake stops, then the vessel is maintained at 35-40 psi (2.41-2.76 bar) and 30±5° C. for 3-4 hours. The reaction vessel is cooled to less than 25° C. and a sample subjected to HPLC to check for 14-hydroxymorphinone. If the 14-hydroxymorphinone area detected by HPLC is >0.1%, the hydrogenation is repeated.

Once it is assessed that the reaction is complete, the catalyst is filtered off, the pH of the filtrate is adjusted to pH 9 using ammonium hydroxide solution, the product precipitates and is isolated by filtration and dried under vacuum. The product is dissolved in dichloromethane/methanol (9:1 v/v) and slurried in florisil, filtered, and the filtrate is distilled to exchange to n-propanol. The n-propanol mixture is cooled and the product precipitates and is collected by filtration in 66% yield. A sample of product is tested by HPLC for alpha, beta unsaturated ketones, and is found to contain 0.51% by area measurement.

Example 1.1C

Formation of Highly Pure Oxymorphone Hydrochloride

A reaction vessel is charged with 1 kg of oxymorphone base, prepared as in Example 1.1B, together with 2.05 kg of absolute alcohol and 0.66 kg of water. The mixture is heated to 60±2° C. and stirred to form a slurry. A hydrochloric acid solution prepared from 0.66 kg concentrated hydrochloric acid, 0.24 kg of water and 0.31 kg of absolute alcohol is added to the oxymorphone base slurry and the pH checked to ensure that it is <1.0. 40 g of 10% Pd on carbon catalyst water-wet paste is added under a stream of nitrogen to the reaction mixture and the mixture is hydrogenated at 35±5 psi (2.41 bar) for 20 hours whilst maintaining a temperature of 65±3° C. The reaction mixture is filtered whilst hot through Celite and a 0.2 µm polish filter. The filtrate is cooled to 0-5° C. over 2-3 hours, and stirred for a further 2 hours to form oxymorphone hydrochloride as a precipitate. The precipitate is washed with absolute alcohol then dried. Yield is 80%.

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A sample of the product is tested by HPLC for the presence of alpha, beta unsaturated ketones, and is found to contain 6.2 ppm.

Example 1.2A

Hydroxylation of Oripavine to 14-hydroxymorphinone

40 g of Oripavine is added with stirring to a reaction vessel containing 30 g of water and 85 g of formic acid, and stirring continued until oripavine is completely dissolved. The temperature remains in the range 20-30° C. Subsequently, 17.72 g of 30 wt % hydrogen peroxide solution is added, and the reaction mixture is stirred for three hours or more, whilst maintaining the temperature in the range 20-35° C. The reaction mixture is cooled to <20° C. and 335 mL of dilute ammonium hydroxide is added slowly, whilst maintaining the reaction mixture below 32° C. If necessary, the pH of the reaction mixture is adjusted to 9.0, with more dilute ammonium hydroxide solution or hydrochloric acid as appropriate, and stirring is continued for 2 hours at 20 C and 2 hours at 4-5° C.

A precipitate of 14-hydroxymorphinone is formed and filtered off. The precipitate is washed with water and then dried to a damp cake and collected for the next stage.

Example 1.2B

Formation of Oxymorphone Base

A hydrogenation vessel is charged with 148 g of water, 90.6 g of acetic acid, and 250 g of damp 14-hydroxymorphinone (48% water content), prepared as in Example 1.2A. The mixture is stirred until clear then 1.34 g of 10% Pd on carbon catalyst (dry weight) in the form of a paste is added under a stream of nitrogen. The hydrogenation vessel is flushed with nitrogen and hydrogen respectively, and then the reaction mixture is hydrogenated at 30° C. and 35 psi (2.41 bar) for 5 hours. An in process test by HPLC indicates an 14-hydroxymorphinone area of 0.07%.

Once it is assessed that the reaction is complete, the catalyst is filtered off through a pad of celite, and the celite cake is washed with 25 mL water. The filtrate is cooled to 0-5° C. and the pH is adjusted to 9.5±0.5 with 1:1 mixture (V/V) of concentrated ammonium hydroxide and water. The precipitate is stirred at 0-5° C. for one hour and isolated by filtration. The crude product is dried in vacuum oven at 50° C. to afford 113 g (86.9% yield) of light beige solid. A sample of product is tested by HPLC for alpha, beta unsaturated ketone, and is found to contain 0.27% by area measurement.

113 g of crude oxymorphone base is taken up in 1.13 L of dichloromethane/methanol (9:1, v/v). 113 g of florisil is added to the solution and the mixture is stirred for 12 hours. The mixture is filtered through a pad of 113 g of florisil, and the florisil cake is rinsed with 120 mL of dichloromethane/methanol. The solvent is removed by distillation and then switched to n-propanol. The batch is cooled to 0-5° C. and stirred for 1 hour to precipitate the oxymorphone base, which is filtered off, washed with cold n-propanol, and dried in a vacuum oven to afford 67.2 g (59.47%) of white solids.

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A sample of product is tested by HPLC for alpha, beta unsaturated ketones, and is found to contain 0.027% by area measurement.

Example 1.2C

Formation of Highly Pure Oxymorphone Hydrochloride

A reaction vessel is charged with 50.1 g of oxymorphone base, prepared as in Example 1.2B, together with 120 g of absolute alcohol. The mixture is heated to 60±2° C. and stirred to form a slurry. A hydrochloric acid solution prepared from 32.7 g concentrated hydrochloric acid and 33.6 g of water is added to the oxymorphone base slurry and the pH is checked to ensure that it is <1.0. 2.0 g of 10% Pd on carbon catalyst water-wet paste is added under a stream of nitrogen to the reaction mixture and the mixture is hydrogenated at 35 psi (2.41 bar) for 20 hours whilst maintaining a temperature of 65° C. The reaction mixture is filtered whilst hot through Celite. The filtrate is cooled to 0-5° C. over 2-3 hours, and stirred for a further 2 hours to form oxymorphone hydrochloride as a precipitate. The precipitate is filtered off, washed with absolute alcohol and then dried to afford white crystals in 77% yield.

A sample of the product is tested by HPLC for the presence of alpha, beta unsaturated ketones, and is found to contain 1.1 ppm.

The above method may be varied by the skilled person whilst still maintaining excellent purity of the product oxymorphone hydrochloride, and examples of such variations follow.

Example 2.1B

Reduction of 14-hydroxymorphinone to Oxymorphone Base

A hydrogenation vessel is charged with 2.5 kg of water and 0.73 kg of acetic acid and 1 kg of 14-hydroxymorphinone is added to the vessel. The reaction mixture is stirred until a clear solution is obtained before 40 g of wet 10% Pd on carbon catalyst is added under a stream of nitrogen. Hydrogen is supplied at 35-40 psi (2.41-2.76 bar). The temperature is maintained at 30±5° C. until hydrogen uptake stops, then the vessel is maintained at 35-40 psi (2.41-2.76 bar) and 30±5° C. for 3-4 hours. The reaction vessel is cooled to less than 25° C. and a sample subjected to HPLC to check for 14-hydroxymorphinone. If the 14-hydroxymorphinone area detected by HPLC is >0.1%, the hydrogenation is repeated.

Once it is assessed that the reaction is complete, the catalyst is filtered off, dichloromethane/methanol (9:1 v/v) is added to the filtrate and the mixture is adjusted to pH 9-10 by adding ammonium hydroxide solution. The dichloromethane/methanol phase is separate, slurried in Florisil, filtered, and the filtrate is distilled to exchange to n-propanol. The n-propanol mixture is cooled and the product precipitates and is collected by filtration in 73% yield. A sample of product is tested by HPLC for alpha, beta unsaturated ketones, and is found to contain 0.32% by area.

Example 2.2B

Reduction of 14-hydroxymorphinone to Oxymorphone Base

A hydrogenation vessel is charged with 35 g of water, 17 g of acetic acid and 38.08 g of 14-hydroxymorphinone, pre-

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pared in Example 1.2A. The reaction mixture is stirred until a clear solution is obtained before 1.8 g of wet 5% Pd on carbon catalyst is added under a stream of nitrogen. Hydrogen is supplied at 35-40 psi (2.41-2.76 bar). The temperature is maintained at 30±5° C. until hydrogen uptake stops, then the vessel is maintained at 35-40 psi (2.41-2.76 bar) and 30±5° C. for 4 hours. The reaction vessel is cooled to less than 25° C., and a sample is analyzed by HPLC to check for 14-hydroxymorphinone. The 14-hydroxymorphinone area detected by HPLC is 0.26%.

Once it is assessed that the reaction is complete, the catalyst is filtered off and the cake is washed with 15 mL of water. 180 mL of dichloromethane/methanol (9:1, v/v) are added to the filtrate and the pH of the mixture is adjusted to pH 9-10 by adding concentrated ammonium hydroxide. The dichloromethane/methanol layer is separated and purified by slurrying with ca. 20 g Florisil. The slurry is filtered and the filtrate is distilled to exchange into n-propanol, and the mixture is cooled to 0-5° C. and stirred for 1-2 hours to precipitate oxymorphone base, which is isolated by filtration. The oxymorphone base is then slurried from n-propanol providing product in 74% yield. A sample of product is tested by HPLC for alpha, beta unsaturated ketones, and is found to contain 0.32% by area.

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Example 2.2C

Formation of Highly Pure Oxymorphone Hydrochloride

A reaction vessel is charged with 2.5 g of oxymorphone base, prepared as in Example 2.2B, together with 7.5 mL of absolute alcohol, 2.5 g of water and 1.66 g of concentrated hydrochloric acid. The mixture is heated to 50-60° C. and a solution results. The pH is checked to ensure that it is <1.0. 0.111 g of 10% Pd on carbon catalyst water-wet paste is added under a stream of nitrogen to the reaction mixture and the mixture is hydrogenated at 35±5 psi (2.41 bar) for 21 hours whilst maintaining a temperature of 65±3° C. The reaction mixture is filtered whilst hot through a 0.45 µm filter. The filtrate is cooled to 0-5° C. over 2-3 hours, and stirred for a further 2 hours to form oxymorphone hydrochloride as a precipitate. The precipitate is filtered off, washed with cold absolute alcohol and dried under vacuum to afford white crystals in 77% yield.

A sample of the product is tested by HPLC for the presence of alpha, beta unsaturated ketones, and is found to contain 2.8 ppm.

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Example 3.1B

Reduction of 14-hydroxymorphinone to Oxymorphone Hydrochloride

The procedure for forming the oxymorphone free base is followed as shown above, but instead of isolating the free base from a dichloromethane/methanol solution, 0.35 volume equivalents of 3N hydrochloric acid are added (vs the volume of the dichloromethane/methanol solution), the reaction mixture is stirred, allowed to stand, and the aqueous layer (contains the product) is separated from the organic layer. The aqueous layer is distilled under vacuum to remove ca. 50% of the volume, and then the remaining solution is cooled over 2 hour to 20-25° C., stirred for 1-2 hours, cooled to 0-5° C. and stirred 2-3 hours. The white solids that form during stirring are filtered off and washed with cold isopropanol. The yield is 64% and the product contains 0.34% of alpha, beta unsaturated ketones.

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Example 3.1C

Purification of Oxymorphone Hydrochloride

Using an analogous process to Example 1.1C, but starting from the product of Example 3.1B, purified oxymorphone hydrochloride is obtained in a yield of 92% and having an undetectable content of alpha, beta unsaturated ketones.

Example 3.2C

Preparation of Highly Pure Oxymorphone Hydrochloride

A reaction vessel is charged with 5.05 g of oxymorphone hydrochloride, prepared in Example 3.1B, together with 13.5 mL of absolute alcohol, 4.5 mL of water and 1.51 g of concentrated hydrochloric acid. The mixture is heated to 50-60° C. and a solution results. The pH is checked to ensure that it is <1.0. 0.21 g of 10% Pd on charcoal catalyst water-wet paste is added under a stream of nitrogen to the reaction mixture and the mixture is hydrogenated at 35±5 psi (2.41 bar) for 20 hours whilst maintaining a temperature of 65±3° C. The reaction mixture is filtered whilst hot through a 0.45 µm filter. The filtrate is cooled to 0-5° C. over 2-3 hours, and stirred for a further 2 hours to form a precipitate. The precipitate is collected by filtration, washed with cold absolute alcohol then dried. Yield is 92%.

A sample of the product is tested by HPLC and found to have an undetectable content of alpha, beta unsaturated ketones.

Without changing the basic process steps, but with small variations in the process steps for starting materials, such as isolation or not of such starting materials, and utilising the essential reduction requirements of the invention for the final step to the purified oxymorphone hydrochloride, other products have been obtained with levels of alpha, beta unsaturated ketones of 3.8 ppm, 1.7 ppm, 6.2 ppm, 6.9 ppm, 2.8 ppm, 3.1 ppm, 0.9 ppm, 6.0 ppm and another undetectable, or zero.

Example 3.2D

Hydration of Oxymorphone Hydrochloride

A drying dish is charged with oxymorphone hydrochloride, prepared as in Example 1.1C, 1.2C, 2.2C, 3.1C or 3.2C, which contains about 5-13 wt % of ethanol. The sample is placed in a vacuum oven along with a dish containing 100 mL of water. A vacuum is applied at 24-29 in Hg and the oven maintained at 20-40° C. for 24 hours. An ethanol-free or low ethanol (approx. 0.04 wt %) product is afforded containing about 10-13 wt % of water. The water absorbed by the sample may be removed in a vacuum oven at 50-55° C. The drying process is stopped when the product's KF is 6-8 wt %. The final hydrated oxymorphone hydrochloride affords a uniform polymorph with a consistent X-ray diffraction pattern.

What is claimed:

1. Oxymorphone hydrochloride having less than 10 ppm, as measured by HPLC, of 14-hydroxymorphinone.
2. Oxymorphone hydrochloride according to claim 1, wherein the content of 14-hydroxymorphinone is less than 5 ppm.
3. A pharmaceutical formulation comprising at least one pharmaceutically acceptable excipient and the oxymorphone hydrochloride according to claim 1.

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4. A method of treating pain comprising administering a pharmaceutical formulation according to claim 3 to a patient in need thereof.

5. A method of purifying a starting material of either oxymorphone or oxymorphone hydrochloride to yield the oxymorphone hydrochloride according to claim 1, comprising exposing the starting material oxymorphone or oxymorphone hydrochloride to hydrogen under reducing conditions in a strongly acid water and alcohol solvent reaction medium at a temperature in the range from 60 to 70° C. for a time sufficient to provide the less than 10 ppm of 14-hydroxymorphinone.

6. The method according to claim 5, wherein the exposing is carried out for a period of at least 20 hours.

7. The method according to claim 5, wherein the reaction medium has a pH of less than 1.

8. The method according to claim 5, wherein the acid is hydrochloric acid.

9. The method according to claim 5, wherein the temperature is approximately 65° C.

10. The method according to claim 5, wherein the starting material oxymorphone or oxymorphone hydrochloride has not been isolated from a reaction mixture in which it is formed.

11. The method according to claim 5, wherein the starting material oxymorphone or oxymorphone hydrochloride has been prepared by a process comprising reduction of 14-hydroxymorphinone.

12. The method according to claim 11, wherein the 14-hydroxymorphinone that is reduced is prepared by a process of hydroxylating oripavine.

13. The method according to claim 12, wherein the oripavine is derived from concentrated poppy straw.

14. The method according to claim 13, wherein the concentrated poppy straw is derived from a high-Thebaine-yielding strain of poppy.

15. The method according to claim 5, comprising the additional steps of subsequently forming crystalline oxymorphone hydrochloride and removing residual alcohol molecules from within the crystal structure of the crystalline oxymorphone hydrochloride by exposing the crystalline oxymorphone hydrochloride to water vapour, such that the residual alcohol molecules are displaced with water molecules.

16. The method according to claim 15, comprising the additional step of removing some of the water molecules from within the crystal structure of the oxymorphone hydrochloride by exposure to reduced pressure.

17. The method according to claim 15, comprising the additional step of removing some of the water molecules from within the crystal structure of the oxymorphone hydrochloride by heating the oxymorphone hydrochloride to a temperature in the range of from 50 to 55° C. under reduced pressure.

18. A method of making hydrated oxymorphone hydrochloride having less than 10 ppm, as measured by HPLC, of 14-hydroxymorphinone and a KF of 6-8 wt %, comprising exposing a starting material of oxymorphone or oxymorphone hydrochloride to gaseous hydrogen under reducing conditions in a strongly acid water and alcohol solvent reaction medium at a temperature in the range from 60 to 70° C., subsequently forming crystalline oxymorphone hydrochloride, and removing residual alcohol molecules from within the crystal structure of the crystalline oxymorphone hydrochloride by exposing the oxymorphone hydrochloride to water vapour, such that the residual alcohol molecules are displaced with water molecules.

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19. Hydrated oxymorphone hydrochloride having less than 10 ppm, as measured by HPLC, of 14-hydroxymorphinone and having peaks within the following 20 ranges when analyzed by Powder X-Ray Diffraction: 8.5-9.5, 11.0-12.0, 11.5-12.5, 12.4-13.4, 15.2-16.2, 17.6-18.6, 19.3-20.3, 19.9-20.9, 24.6-25.6, 24.9-25.9, 29.0-30.0 and 31.0-32.0.

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20. Oxymorphone hydrochloride prepared by the method of claim 5.

21. Hydrated oxymorphone hydrochloride prepared by the method of claim 18.

* * * * *

UNITED STATES PATENT AND TRADEMARK OFFICE
CERTIFICATE OF CORRECTION

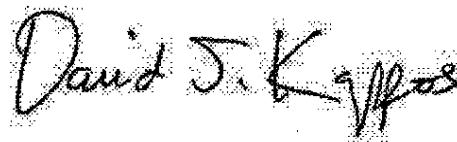
PATENT NO. : 7,851,482 B2
APPLICATION NO. : 11/866840
DATED : December 14, 2010
INVENTOR(S) : Jen-Sen Dung et al.

Page 1 of 1

It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

At column 23, line 3, delete "20 ranges" and insert therefor --2Θ ranges--.

Signed and Sealed this
Nineteenth Day of July, 2011



David J. Kappos
Director of the United States Patent and Trademark Office

Exhibit B



US008309122B2

(12) United States Patent
Kao et al.(10) Patent No.: US 8,309,122 B2
(45) Date of Patent: *Nov. 13, 2012

(54) OXYMORPHONE CONTROLLED RELEASE FORMULATIONS

(75) Inventors: **Huai-Hung Kao**, Syosset, NY (US); **Anand R. Baichwal**, Wappingers Falls, NY (US); **Troy McCall**, Smyrna, GA (US); **David Lee**, Chadds Ford, PA (US)(73) Assignee: **Endo Pharmaceuticals Inc.**, Chadds Ford, PA (US)

(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 1344 days.

This patent is subject to a terminal disclaimer.

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(63) Continuation of application No. 10/190,192, filed on Jul. 3, 2002.

(60) Provisional application No. 60/303,357, filed on Jul. 6, 2001, provisional application No. 60/329,432, filed on Oct. 15, 2001, provisional application No. 60/329,444, filed on Oct. 15, 2001, provisional application No. 60/329,445, filed on Oct. 15, 2001.

(51) Int. Cl.

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A61K 9/34 (2006.01)
A61K 9/36 (2006.01)

(52) U.S. Cl. 424/464; 424/468; 424/470; 424/479;
424/481; 424/482; 424/486(58) Field of Classification Search None
See application file for complete search history.

(56) References Cited

U.S. PATENT DOCUMENTS

2,806,033 A	9/1957	Lewenstein et al.
3,393,197 A	7/1968	Pachter et al.
3,845,770 A	11/1974	Theeuwes et al.
3,879,555 A	4/1975	Pachter et al.
3,966,940 A	6/1976	Pachter et al.
3,980,766 A	9/1976	Shaw et al.
4,070,494 A	1/1978	Hoffmeister et al.
4,366,159 A	12/1982	Magruder
4,457,933 A	7/1984	Gordon et al.
4,464,376 A	8/1984	Sunshine et al.
4,479,956 A	10/1984	Sunshine et al.

(Continued)

FOREIGN PATENT DOCUMENTS

CA 2314896 A1 7/1999

(Continued)

OTHER PUBLICATIONS

Ansel et al. Pharmaceutical dosage forms and drug delivery systems. 1999, 7th edition, p. 121-122.*

(Continued)

Primary Examiner — Lakshmi Channavajjala

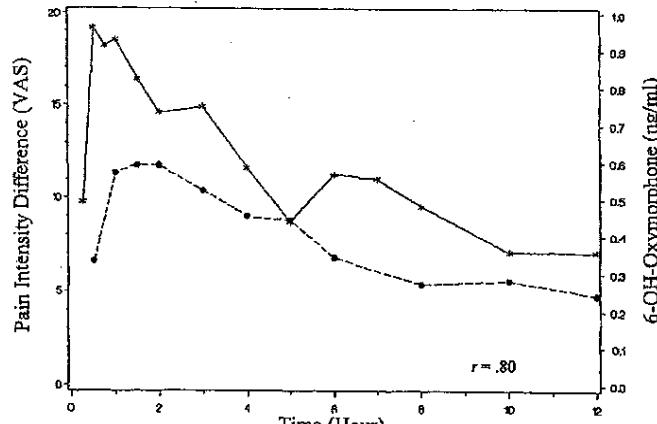
(74) Attorney, Agent, or Firm — Meyer Brown LLP

(57) ABSTRACT

The invention pertains to a method of relieving pain by administering a controlled release pharmaceutical tablet containing oxymorphone which produces a mean minimum blood plasma level 12 to 24 hours after dosing, as well as the tablet producing the sustained pain relief.

20 Claims, 10 Drawing Sheets

PK Profile for 6-OH-Oxymorphone with PID Scores



* Pain Intensity Difference • 6-OH-Oxymorphone Plasma Concentrations

US 8,309,122 B2

Page 2

U.S. PATENT DOCUMENTS					
4,486,436 A	12/1984	Sunshine et al.	6,387,394 B1	5/2002 Baichwal et al.	
4,558,051 A	12/1985	Sunshine et al.	6,391,336 B1	5/2002 Royer	
4,567,183 A	1/1986	Sunshine et al.	6,413,494 B1	7/2002 Lee et al.	
4,569,937 A *	2/1986	Baker et al.	514/282	6,432,438 B1 8/2002 Shukla	
4,582,835 A	4/1986	Lewis et al.	6,475,494 B2	11/2002 Kaiko et al.	
4,587,249 A	5/1986	Sunshine et al.	6,495,155 B1	12/2002 Tice et al.	
4,599,114 A	7/1986	Atkinson	6,506,730 B1	1/2003 Lee et al.	
4,656,177 A	4/1987	Sunshine et al.	6,514,531 B1	2/2003 Alaux et al.	
4,661,492 A	4/1987	Lewis et al.	6,555,127 B2	4/2003 Steiner	
4,711,782 A	12/1987	Okada et al.	6,627,635 B2	9/2003 Palermo et al.	
4,777,174 A	10/1988	Sunshine et al.	6,696,088 B2	2/2004 Oshlack et al.	
4,844,907 A	7/1989	Elger et al.	6,716,449 B2	4/2004 Oshlack et al.	
4,844,909 A	7/1989	Goldie et al.	6,806,294 B2	10/2004 Wimmer et al.	
4,861,598 A	8/1989	Oshlack	7,276,250 B2	10/2007 Baichwal et al.	
4,935,428 A	6/1990	Lewis et al.	2001/0008639 A1	7/2001 Oshlack et al.	
4,980,170 A	12/1990	Schneider et al.	2002/0010127 A1	1/2002 Oshlack et al.	
4,994,276 A	2/1991	Baichwal et al.	2002/0032581 A1	3/2002 Reitburg	
5,047,248 A *	9/1991	Calanchi et al.	424/485	2002/0044966 A1 4/2002 Bartholomaeus et al.	
5,128,143 A	7/1992	Baichwal et al.	2002/0058673 A1	5/2002 Kaiko et al.	
5,135,757 A	8/1992	Baichwal et al.	2002/0081333 A1	6/2002 Oshlack et al.	
5,164,193 A	11/1992	Okada et al.	2002/0090345 A1	7/2002 Baichwal et al.	
5,202,128 A	4/1993	Morella et al.	2002/0164373 A1	11/2002 Maloney	
5,236,714 A	8/1993	Lee et al.	2002/0187192 A1	12/2002 Joshi et al.	
5,266,331 A	11/1993	Oshlack et al.	2003/0004177 A1	1/2003 Kao et al.	
5,330,761 A	7/1994	Baichwal	2003/0031712 A1	2/2003 Kaiko et al.	
5,399,359 A	3/1995	Baichwal et al.	2003/0044458 A1	3/2003 Wright, IV et al.	
5,399,362 A	3/1995	Baichwal et al.	2003/0049272 A1	3/2003 Joshi et al.	
5,415,871 A	5/1995	Pankhania et al.	2003/0059397 A1	3/2003 Hughes	
5,431,922 A	7/1995	Nicklasson	2003/0064099 A1	4/2003 Oshlack et al.	
5,455,046 A	10/1995	Baichwal	2003/0064122 A1	4/2003 Goldberg et al.	
5,470,584 A	11/1995	Hendrickson et al.	2003/0065002 A1	4/2003 Caruso et al.	
5,478,577 A	12/1995	Sackler et al.	2003/0068276 A1	4/2003 Hughes et al.	
5,512,297 A	4/1996	Baichwal	2003/0068370 A1	4/2003 Sackler	
5,512,578 A	4/1996	Crain et al.	2003/0068371 A1	4/2003 Oshlack et al.	
5,554,387 A	9/1996	Baichwal	2003/0068375 A1	4/2003 Wright et al.	
5,567,754 A	10/1996	Stramet	2003/0068392 A1	4/2003 Sackler	
5,580,578 A	12/1996	Oshlack et al.	2003/0069263 A1	4/2003 Breder et al.	
5,612,053 A	3/1997	Baichwal et al.	2003/0073714 A1	4/2003 Breder et al.	
5,629,011 A	5/1997	Illum	2003/0091635 A1	5/2003 Baichwal et al.	
5,633,000 A	5/1997	Grossman et al.	2003/0124061 A1	7/2003 Roberts	
5,639,476 A	6/1997	Oshlack et al.	2003/0124185 A1	7/2003 Oshlack et al.	
5,662,933 A	9/1997	Baichwal et al.	2003/0125347 A1	7/2003 Anderson et al.	
5,672,360 A	9/1997	Sackler et al.	2003/0129230 A1	7/2003 Baichwal et al.	
5,738,865 A	4/1998	Baichwal et al.	2003/0129234 A1	7/2003 Baichwal et al.	
5,858,388 A	1/1999	Grossman et al.	2003/0143269 A1	7/2003 Oshlack et al.	
5,891,474 A	4/1999	Busetti et al.	2003/0147975 A1	8/2003 Joshi et al.	
5,914,131 A	6/1999	Merrill et al.	2003/0152638 A1	8/2003 Tice et al.	
5,948,438 A	9/1999	Stanforth et al.	2003/0157167 A1	8/2003 Kao et al.	
5,958,452 A	9/1999	Oshlack et al.	2003/0157168 A1	8/2003 Breder et al.	
5,958,456 A	9/1999	Baichwal et al.	2003/0158264 A1	8/2003 Radhakrishnan et al.	
5,958,458 A	9/1999	Norling et al.	2003/0163099 A1	8/2003 Wermeling et al.	
5,958,459 A	9/1999	Chasin et al.	2003/0170181 A1	9/2003 Midha	
5,965,161 A	10/1999	Oshlack et al.	2003/0190362 A1	10/2003 Sackler et al.	
5,965,163 A	10/1999	Miller et al.	2007/0098792 A1	5/2007 Kao et al.	
5,968,551 A	10/1999	Oshlack et al.	2007/0098793 A1	5/2007 Kao et al.	
RE36,547 E	2/2000	Crain et al.	2007/0098794 A1	5/2007 Kao et al.	
6,039,980 A	3/2000	Baichwal et al.	2007/0134328 A1	6/2007 Kao et al.	
6,093,420 A *	7/2000	Baichwal	424/468	2007/0140975 A1	6/2007 Baichwal et al.
6,103,258 A	8/2000	Simon	2008/0050431 A1	2/2008 Baichwal et al.	
6,103,261 A	8/2000	Chasin et al.	2008/0085303 A1	4/2008 Baichwal et al.	
6,129,933 A	10/2000	Oshlack et al.	2008/0085304 A1	4/2008 Baichwal et al.	
6,143,322 A	1/2000	Sackler et al.	2008/0085305 A1	4/2008 Baichwal et al.	
6,143,325 A	11/2000	Dennis et al.	2008/0119501 A1	5/2008 Hein et al.	
6,166,211 A	12/2000	Cain et al.	2008/0262013 A1	10/2008 Kao et al.	
6,228,398 B1	5/2001	Devane et al.	2008/0318993 A1	12/2008 Ahdich	
6,228,863 B1	5/2001	Palermo et al.	2008/0318994 A1	12/2008 Ahdich	
6,245,351 B1	6/2001	Nara et al.			
6,245,357 B1	6/2001	Edgren et al.			
6,248,789 B1	6/2001	Weg			
6,261,599 B1	7/2001	Oshlack et al.			
6,277,384 B1	8/2001	Kaiko et al.			
6,294,195 B1	9/2001	Oshlack et al.			
6,296,842 B1	10/2001	Jaworowicz et al.			
6,306,425 B1	10/2001	Tice et al.			
6,309,668 B1	10/2001	Bastin et al.			
6,316,031 B1	11/2001	Oshlack et al.			
6,340,475 B2	1/2002	Shell et al.			
6,375,957 B1	4/2002	Kaiko et al.			

FOREIGN PATENT DOCUMENTS

CA	2369302 A1	10/2000
DE	1 517 480 A1	7/1978
EP	0 253 104 A1	1/1988
EP	319243 A1	6/1989
EP	360562 B2	3/1990
EP	441833 B1	9/1993
EP	0636366 A2	2/1995
EP	751766 A1	1/1997
EP	0 793 959 A1	9/1997
EP	742711 B1	3/1999
EP	1293195 A1	3/2003

US 8,309,122 B2

Page 3

EP	1293209	A1	3/2003	Sargent et al., Hydroxylated Codeine Derivatives, <i>J. Org. Chem.</i> , vol. 23 at 1247 (Sep. 1958).
JP	2003113074	A	4/2003	Weiss, Derivatives of Morphine. IV. 14-Hydroxymorphine and 14-Hydroxydihydromorphine, <i>J. Med. Chem.</i> , vol. 8 at 123 (Jan. 1965).
NZ	0505192		7/1999	U.S. Appl. No. 12/426,112, Kao et al.
WO	80/00841	A1	5/1980	U.S. Appl. No. 11/766,748, Ahdieh.
WO	84/00488	A1	2/1984	U.S. Appl. No. 11/742,956, GerritsenvanderHoop.
WO	84/00490	A1	2/1984	U.S. Appl. No. 12/203,758, Ahdieh.
WO	85/02540	A1	6/1985	Abbott Laboratories, Package Insert for VICODIN®, Mar. 2007.
WO	85/02542	A1	6/1985	Adams & Ahdieh, "Single- and Multiple-Dose Pharmacokinetic and Dose-Proportionality Study of Oxymorphone Immediate Release Tablets." <i>Drugs R D</i> , vol. 6(2), pp. 91-99 (2005).
WO	91/07950	A1	6/1991	Adams & Ahdieh, "Single- and Multiple-Dose Pharmacokinetic and Dose-Proportionality Study of Oxymorphone Immediate Release Tablets." Amer. Pharmacists Assoc., Mar. 2004, Poster Presentation.
WO	WO-93/17673	A1	9/1993	Adams et al., "Oxymorphone Extended Release Does Not Affect CYP2C9 or CYP3A4 Metabolic Pathways." <i>J Clinical Pharmacology</i> , vol. 45, pp. 337-345 (2005).
WO	95/20947	A1	8/1995	Adams et al., "Oxymorphone Extended Release Does Not Affect CYP2C9 or CYP3A4 Metabolic Pathways." Amer. Acad. Pain Management Mar. 2004, Poster Presentation.
WO	95/22965	A2	8/1995	Adams et al., "A New Oral Opioid, Oxymorphone Extended Release, Does Not Affect Human Metabolic Enzymes CYP2C9 or CYP3A4." Amer. Pharmacists Assoc., Mar. 2004, Poster Presentation.
WO	96/00047	A1	1/1996	Adams & Ahdieh, "Pharmacokinetics and dose-proportionality of oxymorphone extended release and its metabolites: Results of a randomized crossover study." <i>Pharmacotherapy</i> , vol. 24(4), pp. 468-476 (2004).
WO	96/02251	A1	2/1996	Adams & Ahdieh, "Pharmacokinetic Analyses of New Formulations of Extended- and Immediate-Release Oxymorphone." Amer. Acad. Pain Management Mar. 2004, Poster Presentation.
WO	96/04007	A1	5/1996	Ahdieh et al., "Oxymorphone Extended Release Provides Safe and Effective Analgesia for Cancer Patients: A Randomized, Double-Blind, Crossover Study with Oxycodone Controlled Release." Amer. Pharmacists Assoc., Mar. 2004, Poster Presentation.
WO	96/20927	A1	7/1996	Ahdieh et al., "Efficacy of Oxymorphone extended release in post-surgical pain: A randomized clinical trial in knee arthroplasty." <i>J. Clinical Pharmacology</i> , vol. 44, pp. 767-776 (2004).
WO	97/07750	A1	3/1997	Ansel H.C. et al., <i>Pharmaceutical Dosage Forms and Drug Delivery Systems</i> , pp. 121-122, (7th Ed.) (1999).
WO	97/16172	A1	5/1997	Baichwal et al., "Gamma Scintigraphy Imaging and Pharmacokinetic Analysis of Extended-Release TIMERx Technology." Amer. Physiological Soc., May 2004, Poster Presentation.
WO	WO-98/00143	A1	1/1998	Beaver, et al., "Comparisons of the Analgesic Effects of Oral and Intramuscular Oxymorphone and of Intramuscular Oxymorphone and Morphine in Patients with Cancer." <i>J Clinical Pharmacology</i> , vol. 17(4), pp. 186-198 (1977).
WO	WO-99/01111	A1	1/1999	Cass et al., "The Control of Severe Pain with Oral Oxymorphone Hydrochloride." <i>Current Therapeutic Research</i> , vol. 5(11) pp. 579-586 (1963).
WO	99/32119	A1	7/1999	Cephalon, Package Insert for ACTIQ®, 2007.
WO	99/32120	A1	7/1999	Chiao et al., Sustained-Released Drug Delivery Systems, Chapter 94, Remington, 1995.
WO	00/01377	A2	1/2000	Cisternas et al., "Management of Chronic Pain With Long-Acting Opioids May Contribute to the Stabilization of Total Healthcare Costs: Results of a Retrospective Claims Database Pharmacoeconomic Study." Amer. Soc. Health-System Pharmacists, Dec. 2004, Poster Presentation.
WO	00/21520	A3	4/2000	Cone, "General procedure for the isolation and identification of 6- α -and 6- β -hydroxymetabolites of narcotic agonists and antagonists with a hydromorphone structure." <i>J. of Chromatogr.</i> , vol. 129, pp. 355-361 (1976).
WO	00/33835	A1	6/2000	Cone et al., "Oxymorphone metabolism and urinary excretion in human, rat, guinea pig, rabbit, and dog." <i>Drug Metabolism and Disposition</i> , vol. 11(5), pp. 446-450 (1983).
WO	00/38649	A1	7/2000	Dhopeshwarkar et al., "Evaluation of Xanthan Gum in the Preparation of Sustained Release Matrix Tablets." <i>Drug Development & Industrial Pharmacy</i> , vol. 19(9), pp. 999-1017 (1993).
WO	00/61147	A1	10/2000	Drakontides, "Drugs to Treat Pain." <i>Amer. J Nursing</i> , vol. 74(3), pp. 508-513 (Mar. 1974).
WO	01/00181	A2	1/2001	Earnes et al., "Clinical Trial of Oxymorphone in Labour." <i>Brit. Med. J.</i> , vol. 2, pp. 353-355 (1964).
WO	01/12230	A1	2/2001	
WO	WO-01/08661	A2	2/2001	
WO	01/15699	A1	3/2001	
WO	WO-01/32148	A1	5/2001	
WO	01/52813	A1	7/2001	
WO	01/58447	A1	8/2001	
WO	01/58451	A1	8/2001	
WO	02/05647	A1	1/2002	
WO	02/13886	A2	2/2002	
WO	02/087558	A1	11/2002	
WO	02/092059	A1	11/2002	
WO	02/092060	A1	11/2002	
WO	02/094172	A2	11/2002	
WO	02/094254	A2	11/2002	
WO	03/004029	A1	1/2003	
WO	03/004030	A1	1/2003	
WO	03/007802	A2	1/2003	
WO	03/013433	A2	2/2003	
WO	03/013476	A1	2/2003	
WO	03/013479	A1	2/2003	
WO	03/013525	A1	2/2003	
WO	03/015531	A2	2/2003	
WO	WO-03/013538	A1	2/2003	
WO	03/026743	A2	4/2003	
WO	03/039561	A1	5/2003	
WO	03/072106	A2	9/2003	
WO	2006/094083	A1	9/2006	
WO	2007/053698	A2	5/2007	
WO	2007/078895	A2	7/2007	

OTHER PUBLICATIONS

Extended Release Oral Dosage Forms: Development, Evaluation, and Application of In Vitro/In Vivo Correlations. Sep. 1997. total No. of pp. 27.*

Hinz et al., Bioavailability of diclofenac potassium at low doses, 59 British Journal of Clinical Pharmacology No. 1, pp. 80-84, 2005.

Cone, Edward J., "General procedure for the isolation and identification of 6- α -and 6- β -hydroxymetabolites of narcotic agonists and antagonists with a hydromorphone structure" *J. Chromatogr.* vol. 129, pp. 355-361 (1976).

Cone, Edward J., "Oxymorphone metabolism and urinary excretion in human, rat, guinea pig, rabbit, and dog" *Drug Metabolism and Disposition* vol. 11(5), pp. 446-450 (1983).

Cass, Use of Oral Analgesic for Severe Pain, Western Medicine, p. 107-108, 120 (Mar. 1961).

Numorphan Oral Advertisement, British Journal of Anaesthesia, vol. 34, No. 8 (Aug. 1962).

News of Products and Services, Products for Dispensing: Numorphan oral, The Pharmaceutical Journal (Jul. 7, 1962).

Cisternas et al., "Management of Chronic Pain With Long-Acting Opioids May Contribute to the Stabilization of Total Healthcare Costs: Results of a Retrospective Claims Database Pharmacoeconomic Study." Amer. Soc. Health-System Pharmacists, Dec. 2004, Poster Presentation.

Cone, "General procedure for the isolation and identification of 6- α -and 6- β -hydroxymetabolites of narcotic agonists and antagonists with a hydromorphone structure." *J. of Chromatogr.*, vol. 129, pp. 355-361 (1976).

Cone et al., "Oxymorphone metabolism and urinary excretion in human, rat, guinea pig, rabbit, and dog." *Drug Metabolism and Disposition*, vol. 11(5), pp. 446-450 (1983).

Dhopeshwarkar et al., "Evaluation of Xanthan Gum in the Preparation of Sustained Release Matrix Tablets." *Drug Development & Industrial Pharmacy*, vol. 19(9), pp. 999-1017 (1993).

Drakontides, "Drugs to Treat Pain." *Amer. J Nursing*, vol. 74(3), pp. 508-513 (Mar. 1974).

Earnes et al., "Clinical Trial of Oxymorphone in Labour." *Brit. Med. J.*, vol. 2, pp. 353-355 (1964).

US 8,309,122 B2

Page 4

- Eddy & Lee, "The analgesic equivalence to morphine and relative side action liability of oxymorphone (4-Hydroxydihydromorphinone)." *J Pharmacology and Experimental Therapeutics*, vol. 125(2), pp. 116-121 (1959).
- Endo Pharmaceuticals, Package Insert for NUMORPHAN®, Apr. 2004.
- Endo Pharmaceuticals, Package Insert for OPANA® (Jul. 2006).
- Endo Pharmaceuticals, Package Insert for PERCOSET®, Nov. 2006.
- Endo Pharmaceuticals, Package Insert for ZYDONE®, Jun. 2003.
- Gabrail et al., "Oxymorphone Extended Release Provides Safe and Effective Analgesia During Opioid Rotation: Results of Randomized, Double-Blind, Crossover, Comparative Study with Oxycodone Controlled Release." *Amer. Acad. Pain Management*, Mar. 2004, Poster Presentation.
- Gabrail et al., "Establishing the dosage equivalency of oxymorphone extended release and oxycodone controlled release in patients with cancer pain: A randomized controlled study." *Current Medical Research Opin.*, vol. 20(6), pp. 911-918 (2004).
- Galer et al., "Safety of New Oral Formulations of the Opioid Oxymorphone." *Int'l Assoc. for the Study of Pain*, May 2004, Poster Presentation.
- Gallagher et al., "Assessment of dosing frequency of sustained-release opioid preparations in patients with chronic nonmalignant pain." *Pain Medicine*, vol. 8(1), pp. 71-74 (2004).
- Gibofsky & Barkin, "Chronic Pain of Osteoarthritis: Considerations for Selecting an Extended Release Opioid Analgesic." *Amer. J Therapeutics*, vol. 15, pp. 241-255 (2008).
- Gimbel & Adams, "Oxymorphone Immediate Release for Postsurgical Pain: Translating Pharmacokinetic Drug Properties Into Clinical Efficacy." *Amer. Cancer Soc. (Abstract)*.
- Gimbel & Ahdieh, "The Efficacy and Safety of Oral Immediate-Release Oxymorphone for Postsurgical Pain." *Anesth. Analg.*, vol. 99, pp. 1472-1477 (2004).
- Gimbel & Walker, "A Randomized Double-Blind Trial of Low-Dose Oxymorphone Immediate Release (5 mg) for Mild to Moderate Pain in Ambulatory Patients." *Amer. Physiological Soc.*, May 2004, Poster Presentation.
- Gimbel et al., "Analgesic Efficacy of Oxymorphone Immediate Release in Postsurgical Orthopedic Pain: Results of a Randomized, Double-Blind, Placebo-Controlled, Dose-Ranging Comparison with Oxycodone." *Amer. Pharmacists Assoc.*, Mar. 2004, Poster Presentation.
- Gimbel et al., "Low-Dose Oxymorphone Immediate Release (5mg) for Mild to Moderate Pain Following Arthroscopic Knee Surgery in Ambulatory Patients: A Randomized Double-Blind Trial." *Amer. Acad. Nurse Practitioners*, Jun.-Jul. 2004, Poster Presentation.
- Gimbel et al., "Efficacy and safety of oxymorphone immediate release for the treatment of mild to moderate pain after ambulatory orthopedic surgery: results of a randomized, double-blind, placebo-controlled trial." *Archives of Physical Medicine Rehabilitation*, vol. 86(12), pp. 2284-2289 (2005).
- Gould et al., "Retrospective and Prospective Analyses of Oxymorphone Extended Release for Neuropathic Pain." *Neuropathic Pain*, Nov. 2004 (Abstract).
- Gould et al., "Effective Titration With Oxymorphone Extended Release in Opioid-Naïve Patients with Low Back Pain." *Amer. Acad. Pain Management*, Feb. 2005 (Abstract).
- Gould et al., "Effective long-term management of opioid-naïve patients with oxymorphone extended release." *Amer. Physiologic Soc.*, Mar. 2005 (Abstract).
- Gould et al., "Oxymorphone extended release for effective long-term treatment of elderly opioid-naïve patients with moderate to severe nonmalignant pain." *Amer. Physiologic Soc.*, Mar. 2005, (Abstract).
- Gould & Ahdieh, "Case Studies of Opioid Treatments for Neuropathic Pain." *Neuropathic Pain*, Nov. 2004 (Abstract).
- Hale & Drass, "Safety, Tolerability, and Effectiveness of Oxymorphone Extended Release in Opioid-Naïve Patients: An Interim Analysis." *Amer. Orthopedic Assoc.*, Nov. 2004, Poster Presentation.
- Hale et al., "Long-Term Safety and Efficacy of Oxymorphone Extended Release in Opioid-Naïve Patients with Chronic Pain." *Amer. Acad. Pain Management*, Mar. 2004, Poster Presentation.
- Hale et al., "Efficacy and safety of oxymorphone extended release in chronic low back pain: results of a randomized, double-blind, placebo- and active-controlled phase III study." *J Pain*, vol. 6(1), pp. 21-28 (2005).
- Hale et al., "Open-Label Long-Term Assessment of Tolerability, Safety, and Effectiveness of Oxymorphone Extended Release for Low Back Pain." *Amer. Acad. Pain Management*, Mar. 2004, Poster Presentation.
- Hale et al., "Tolerability and Effectiveness of Oxymorphone Extended Release in Opioid-Naïve Patients with Chronic Pain." *Amer. Pharmacists Assoc.*, Mar. 2004, Poster Presentation.
- Hale et al., "Low-Dose Titration of Oxymorphone Extended Release in Opioid-Naïve Patients With Chronic Pain: Short- and Long-Term Results." *Amer. Acad. of Physical Medicine and Rehabilitation*, Oct. 2004, Poster Presentation.
- Hinz et al., "Bioavailability of diclofenac potassium at low doses." *British Journal of Clinical Pharmacology*, vol. 59, No. 1, pp. 80-84 (2005).
- International Search Report issued in PCT/US02/21403, mailed Oct. 31, 2002.
- International Search Report issued in PCT/US02/21396, mailed Nov. 6, 2002.
- International Search Report issued in PCT/US02/21398, mailed Nov. 6, 2002.
- International Search Report issued in PCT/US02/21400, mailed Oct. 31, 2002.
- International Search Report issued in PCT/US02/21354, mailed Nov. 6, 2002.
- Kafka et al., "Effective Titration With Oxymorphone Extended Release for Opioid-Naïve Osteoarthritis Patients with Moderate to Severe Pain." *Amer. Acad. Pain Management*, Feb. 2005 (Abstract).
- King Pharmaceuticals, Package Insert for AVINZA®, Oct. 2006.
- Kivitz et al., "A 2-week, multicenter, randomized, double-blind, placebo-controlled, dose-ranging, phase III trial comparing the efficacy of oxymorphone extended release and placebo in adults with pain associated with osteoarthritis of the hip or knee." *Clinical Therapeutics*, vol. 28(3), pp. 352-364 (2006).
- Loan et al., "Studies of drugs given before anaesthesia, XVII: The natural and semi-synthetic opiates." *Brit. J. Anaesth.*, vol. 41, pp. 57-63 (1969).
- Matsumoto et al., "Oxymorphone extended-release tablets relieve moderate to severe pain and improve physical function in osteoarthritis: Results of randomized, double-blind, placebo- and active-controlled phase III trial." *Pain Medicine*, vol. 6(5), pp. 357-366 (2005).
- McIlwain, "Safety and Tolerability of Oxymorphone Extended Release During Long-Term Treatment of Moderate to Severe Pain From Osteoarthritis." *ASCP*, Nov. 2004, Poster Presentation.
- McIlwain, "Safety and Effectiveness of Oxymorphone Extended Release During Long-Term Treatment of Moderate to Severe Pain from Osteoarthritis: One-Year Results." *Amer. Osteopathic Assoc.*, Nov. 2004, Poster Presentation.
- McIlwain et al., "Oxymorphone Extended Release Maintains Effectiveness and Is Well Tolerated During Long-Term Treatment of Moderate to Severe Osteoarthritis Pain." *Amer. Acad. Pain Management*, Mar. 2004, Poster Presentation.
- McIlwain & Ahdieh, "Long-Term Effectiveness and Safety of a New Oral Opioid, Oxymorphone Extended Release, for Moderate to Severe Osteoarthritis Pain." *Amer. Pharmacists Assoc.*, Mar. 2004, Poster Presentation.
- McIlwain & Ahdieh, "Safety, Tolerability, and Effectiveness of Oxymorphone Extended release for moderate to severe osteoarthritis pain: a one-year study." *Amer. J Therapeutics*, vol. 11(5), pp. 1-7 (2004).
- Ossipov & Porreca, "Challenges in the Development of Novel Treatment Strategies for Neuropathic Pain." *J. Amer. Society for Experimental NeuroTherapeutics*, vol. 2, pp. 650-661 (2005).
- Pformulate: <http://www.pformulate.com/disintegrals.html> (published on May 5, 2000).
- Pieniaszek et al., "Oxymorphone Does Not Affect CYP450 Enzymes 2C9 or 3A4: Positive Implications for Pain Management." *Amer. Physiological Soc.*, May 2004, Poster Presentation.

US 8,309,122 B2

Page 5

- Pieniaszek et al., "Oxymorphone Exhibits a Low Potential for Drug Interactions Through CYP450 Isozyme 3A4: In Vitro and Clinical Studies." Amer. Acad. Physical Medicine and Rehabilitation, Oct. 2004 (Abstract).
- Plummer et al., "Influence of polarity on dose-response relationships of intrathecal opioids in rats." Pain, vol. 40, pp. 339-347 (1989).
- Prager & Rauck, "Oxymorphone Extended Release for Moderate to Severe Neuropathic Pain: Open-Label, Long-Term Study of Safety and Effectiveness." Amer. Physiological Soc., May 2004, Poster Presentation.
- Purdue Pharma L.P., Package Insert for MS CONTIN®, 2007.
- Rauck, "Oxymorphone Extended Release for Moderate to Severe Neuropathic Pain." ASCP, Nov. 2004, Poster Presentation.
- Rhiner et al., "Long-Term Safety, Effectiveness, and Dose Stabilization of Oxymorphone Extended Release in Cancer Pain." Amer. Acad. Nurse Practitioners, Jun. 2004, Poster Presentation.
- Rowland & Tozer, *Clinical Pharmacokinetics: Concepts and Applications*, pp. 152-160, 392-393 (2d ed. 1989).
- Shuey et al., "Reproductive and Developmental Toxicity Studies with Oxymorphone: A Potent Opioid Analgesic." Teratology Soc., Jun. 2004, Poster Presentation.
- Slatkin et al., "Oxymorphone Maintains Effectiveness and Is Well Tolerated During Long-Term Treatment of Moderate to Severe Cancer Pain." Amer. Acad. Pain Management, Mar. 2004, Poster Presentation.
- Slatkin et al., "Long-Term Effectiveness and Safety of a New Oral Opioid, Oxymorphone Extended Release, for Moderate to Severe Cancer Pain." Amer. Pharmacists Assoc., Mar. 2004, Poster Presentation.
- Slatkin et al., "Case Studies of Cancer Patients with Neuropathic Pain Treated with Oxymorphone." Int'l Assoc. for the Study of Pain, May 2004, Poster Presentation.
- Slatkin et al., "Long-Term Treatment of Moderate to Severe Cancer Pain: A 2-Year Study." Amer. Physiological Soc., May 2004, Poster Presentation.
- Slatkin et al., "Effectiveness, safety, and tolerability of oxymorphone extended release for moderate to severe cancer pain." Amer. Soc. Clinical Oncology, Jun. 2004 (Abstract).
- Sloan et al., "Effectiveness and safety of oral extended-release oxymorphone for the treatment of cancer pain: a pilot study." Supportive Care Cancer, vol. 13(1), pp. 57-65 (2005).
- Swerdlow & Brown, "Numorphan: A new supplement to anaesthesia." Brit. J. Anaesth., vol. 33, pp. 126-129 (1961).
- Tark et al., "Safety, Tolerability, and Effectiveness of Oxymorphone Extended Release During Long-Term Treatment of Cancer Pain: Results of a 12-month Open-Label Study." Multi-National Assoc. Supportive Cancer Care, Jun. 2004, Poster Presentation.
- Vashi et al., "Oral and I.V. oxymorphone clinical pharmacokinetics compared to those of oral oxycodone pharmacokinetics." ASHP Mid-year Clinical Meeting, vol. 39, pp. P435E (2004) (abstract of meeting presentation).
- Walker et al., "A Randomized Double-Blind Trial of Low-Dose Oxymorphone Immediate Release (5 mg) for Mild to Moderate Pain in Ambulatory Patients." Amer. Pharmacists Assoc., Mar. 2004, Poster Presentation.
- White et al., "Application of Propensity Scores to Claims Data: Correcting Biases in Drug-Prescribing Patterns of Long-Acting Opioids." Amer. Soc. Health-System Pharmacists, Dec. 2004, Poster Presentation.
- White et al., "Improved quality of life during long-term treatment of moderate to severe pain with oxymorphone ER." Amer. Physiologic Soc., Mar. 2005 (Abstract).
- White, "Comment: therapy switching in patients receiving long-acting opioids." Annals of Pharmacotherapy, vol. 38(10), pp. 1752-1752 (2004).
- Zeller et al., "Acute Pain Treatment." JAMA vol. 299(1) (2008).
- McConvile et al., "Use of a Novel Modified TSI for the Evaluation of Controlled-Release Aerosol Formulations. I", Drug Dev Ind Pharmacy, vol. 26, No. 11, pp. 1191-1198 (2000).
- Complaint, *Endo Pharmaceuticals, Inc. and Penwest Pharmaceuticals Inc. v. Impax Laboratories, Inc.*, Docket No. 1:07cv731, entered Nov. 16, 2007.
- Answer to Complaint and Counterclaim filed by Impax Laboratories, Inc., Docket No. 1:07cv731, Entered Dec. 20, 2007.
- Answer to Counterclaim by Endo Pharmaceuticals Inc. and Penwest Pharmaceuticals Co., Docket No. 1:07cv731, entered Jan. 14, 2008.
- Complaint, *Endo Pharmaceuticals, Inc. and Penwest Pharmaceuticals, Inc. v. Impax Laboratories, Inc.*, Docket No. 1:08-CV-00057-UNA filed Jan. 25, 2008.
- Complaint, *Endo Pharmaceuticals Inc. and Penwest Pharmaceuticals Co. v. Actavis South Atlantic LLC*, Docket No. 2:08-cv-01563-KSH-PS, United States District Court, District of New Jersey, filed Mar. 28, 2008.
- International Search Report for PCT/US2007/005560, Sep. 17, 2007.
- United States Patent and Trademark Office, Before the Board of Appeals and Interference, "Decision on Appeal, Appeal No. 2005-0416, U.S. Appl. No. 09/970,020." Apr. 28, 2005.
- Staniforth and Baichwal, "Synergistically Interacting Heterodisperse Polysaccharides—Function in Achieving Controllable Drug Delivery." American Chemical Society, pp. 327-350 (1993).
- Mandema et al., "Characterization and validation of a pharmacokinetic model for controlled-release oxycodone." British J Clinical Pharmacology, vol. 42(6), pp. 747-756 (1996).
- Benzinger et al., "A Pharmacokinetic/Pharmacodynamic Study of Controlled-Release Oxycodeone." J Pain and Symptom Management, vol. 13(2), pp. 75-82 (1997).
- Sathyam et al., "Pharmacokinetic profile of a 24-hour controlled-release OROS® formulation of hydromorphone in the presence and absence of food." BMC Clinical Pharmacology, vol. 7(2) (2007).
- Johnson et al., "Effect of Concomitant Ingestion of Alcohol on the In Vivo Pharmacokinetics of KADIAN (Morphine Sulfate Extended-Release) Capsules." J of Pain, vol. 9(4), pp. 330-336 (2008).
- Cappola, "A Better Dissolution Method for Ranitidine Tablets USP." Pharmaceutical Development and Technology, vol. 6(1), pp. 11-17 (2001).
- De Haan & Lerk, "Studies on different dissolution models." Pharmaceutisch Weekblad Scientific Edition, vol. 4, pp. 191-196 (1982).
- Karasulu & Ertan, "Different geometric shaped hydrogel theophylline tablets: statistical approach for estimating drug release." II Farmaco, vol. 57, pp. 939-945 (2002).
- United States Food and Drug Administration Alert Alcohol and Palladone Interaction available at www.fda.gov/CDER/Drug/info/page/palladone/default.htm, updated Jul. 2005.
- "Don't mix alcohol & pain killers" American Running & Fitness Association, available at findarticles/p/articles/mi_m0NHF/is_6_17/ai_8664966/print?tag=artBody.col1 (1999).
- McNicol et al., "Management of opioids side effects in cancer related and chronic non cancer pain: a systematic review." J of Pain 4(5), pp. 231-256 (2003).
- Weiss, "Derivatives of Morphine. I. 14-Hydroxydihydromorphine." J American Chemical Society, 77(22), p. 5891-92 (1955).

* cited by examiner

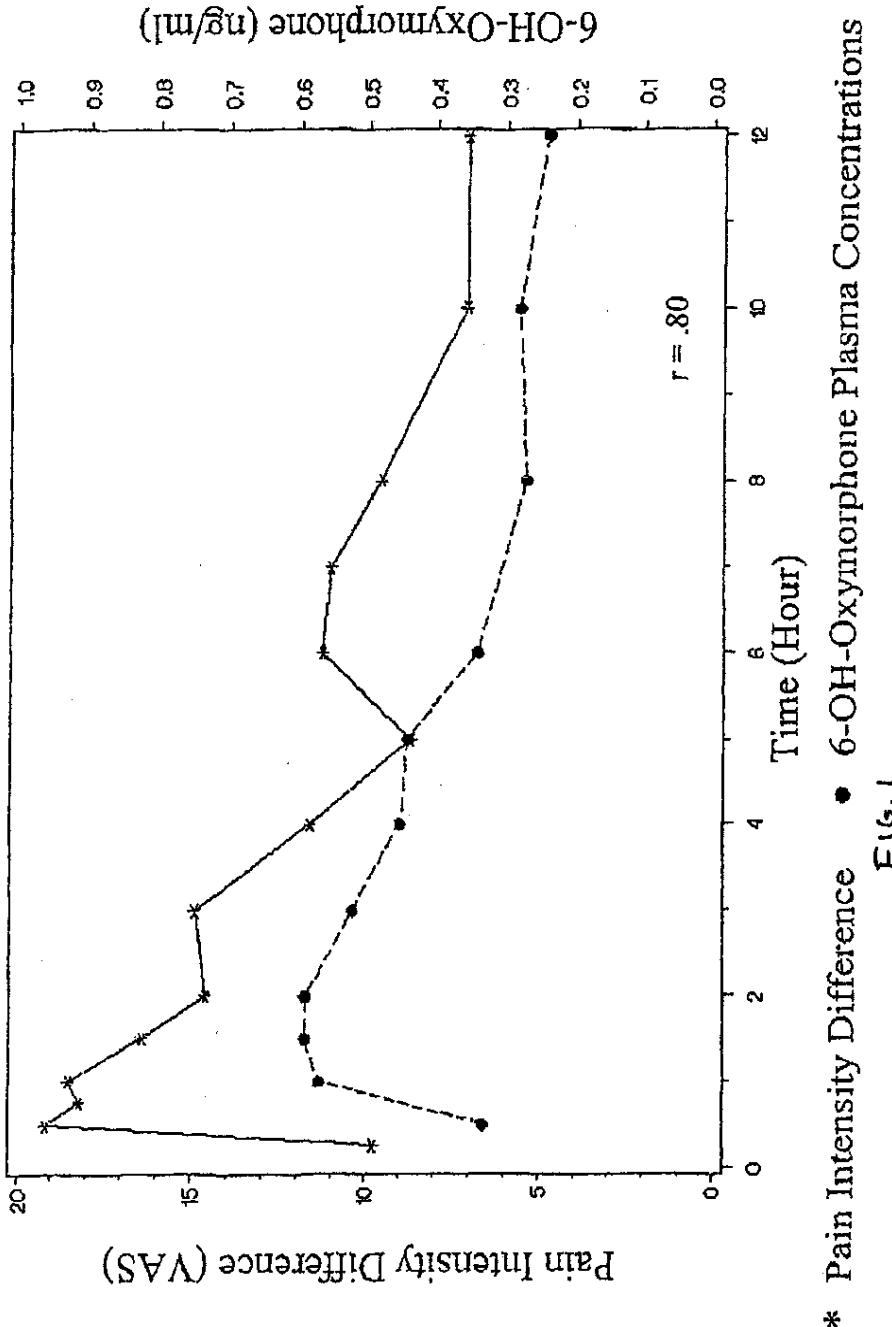
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PK Profile for 6-OH-Oxymorphone with PID Scores

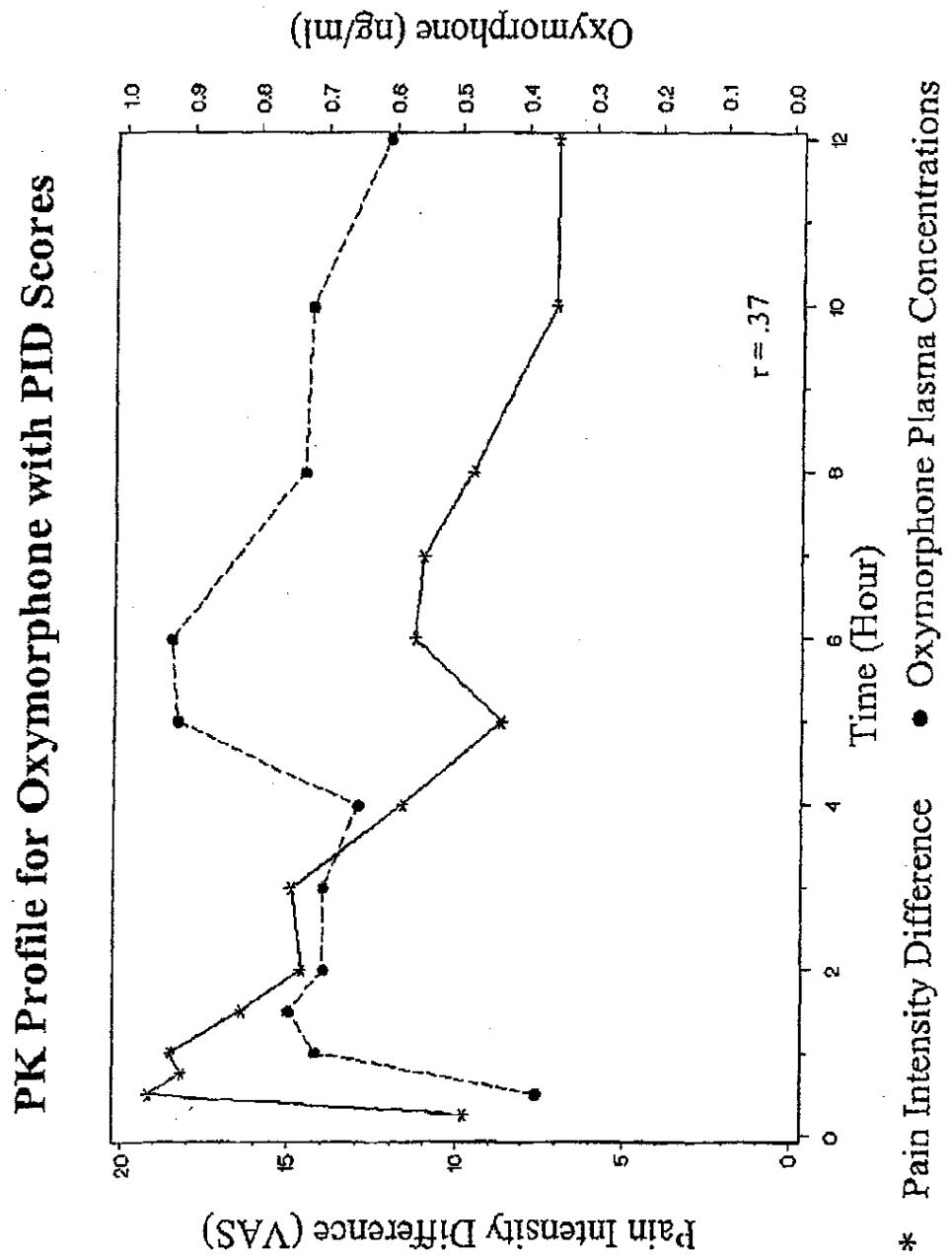


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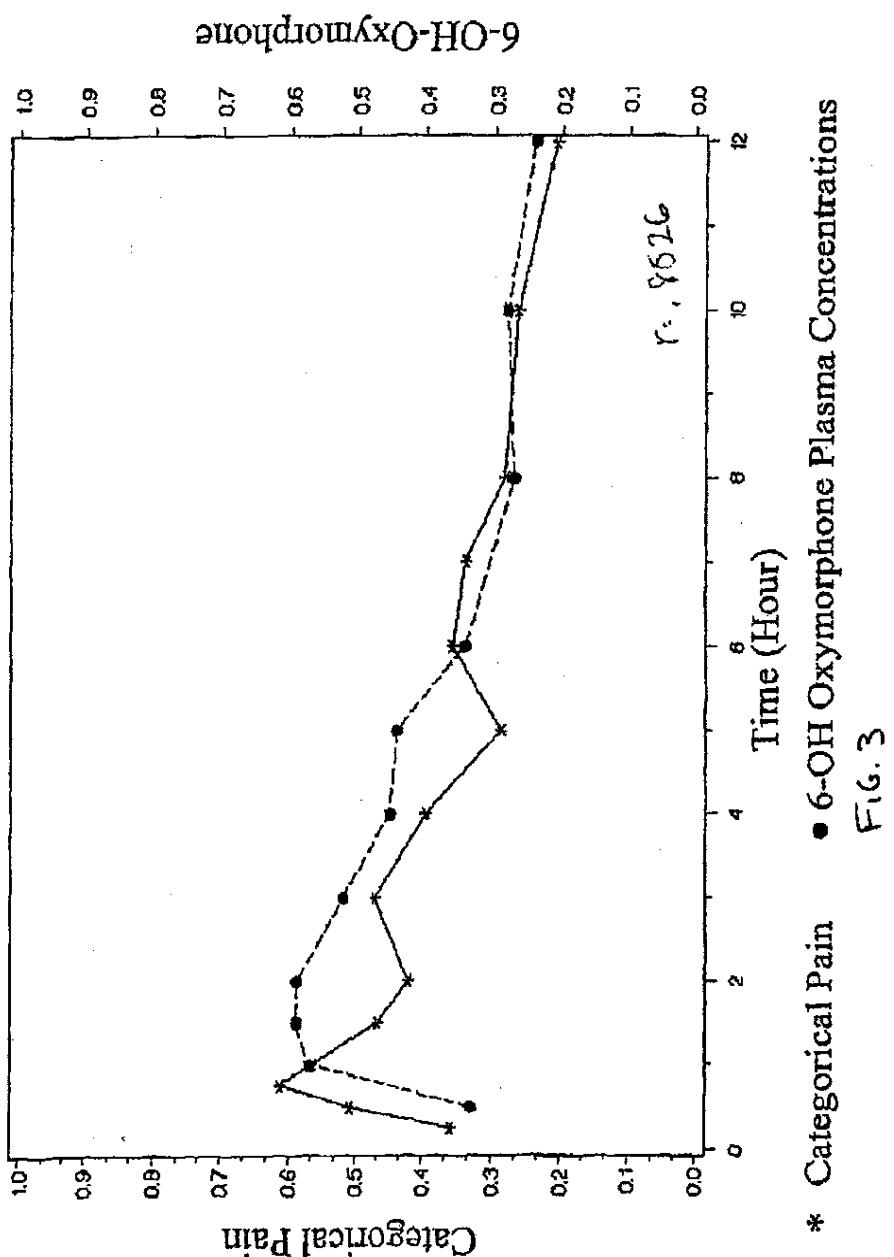
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PK Profile for 6-OH-Oxymorphone with Categorical Pain Scores

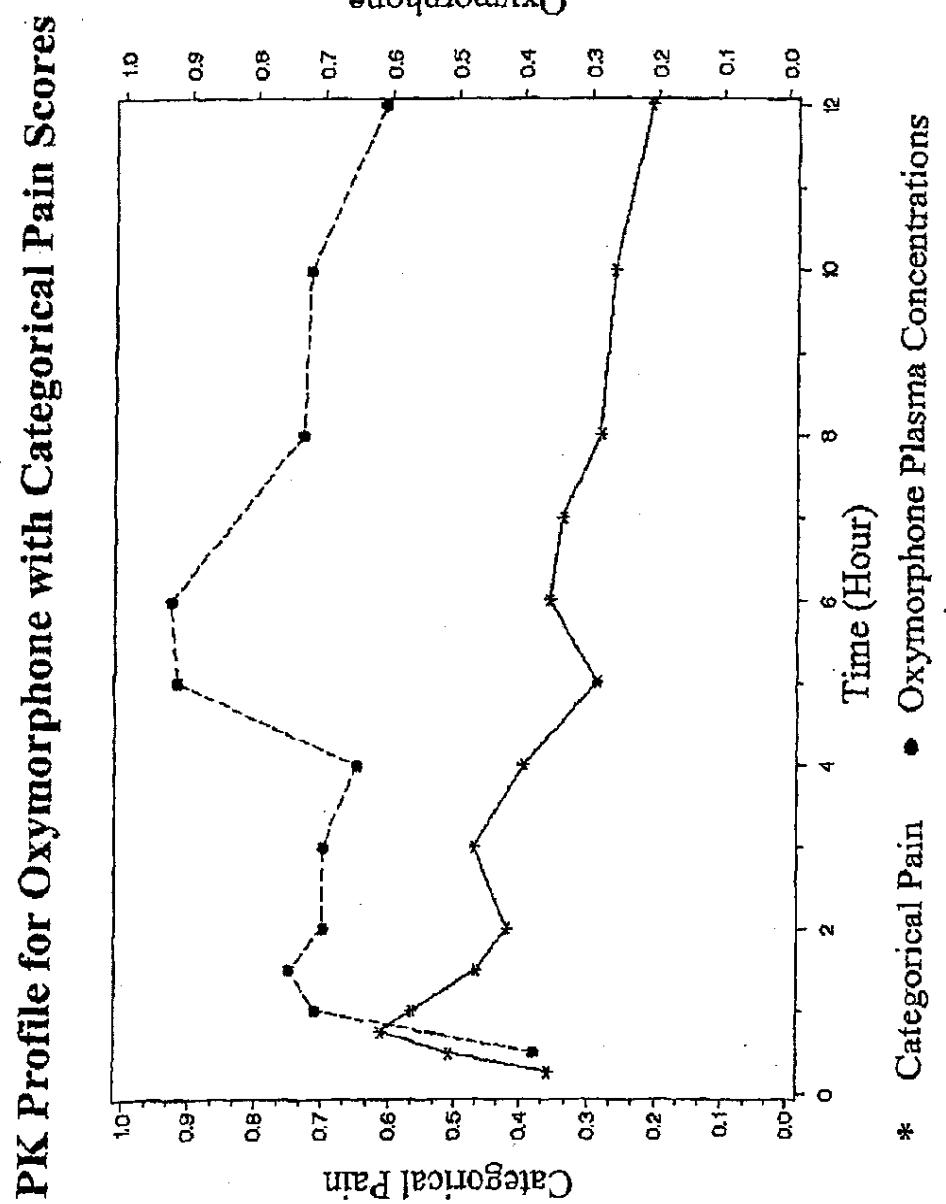


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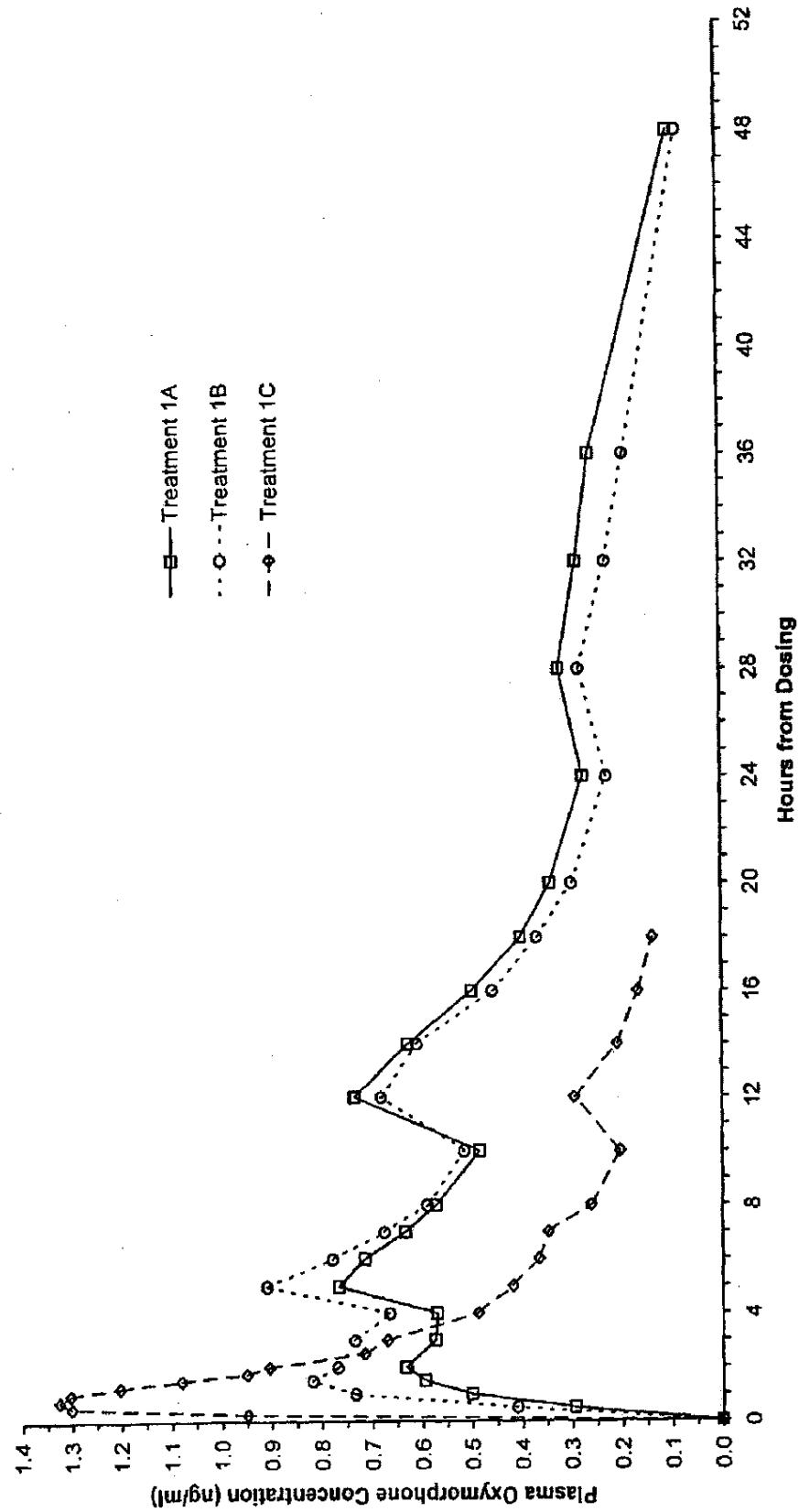


Figure 5

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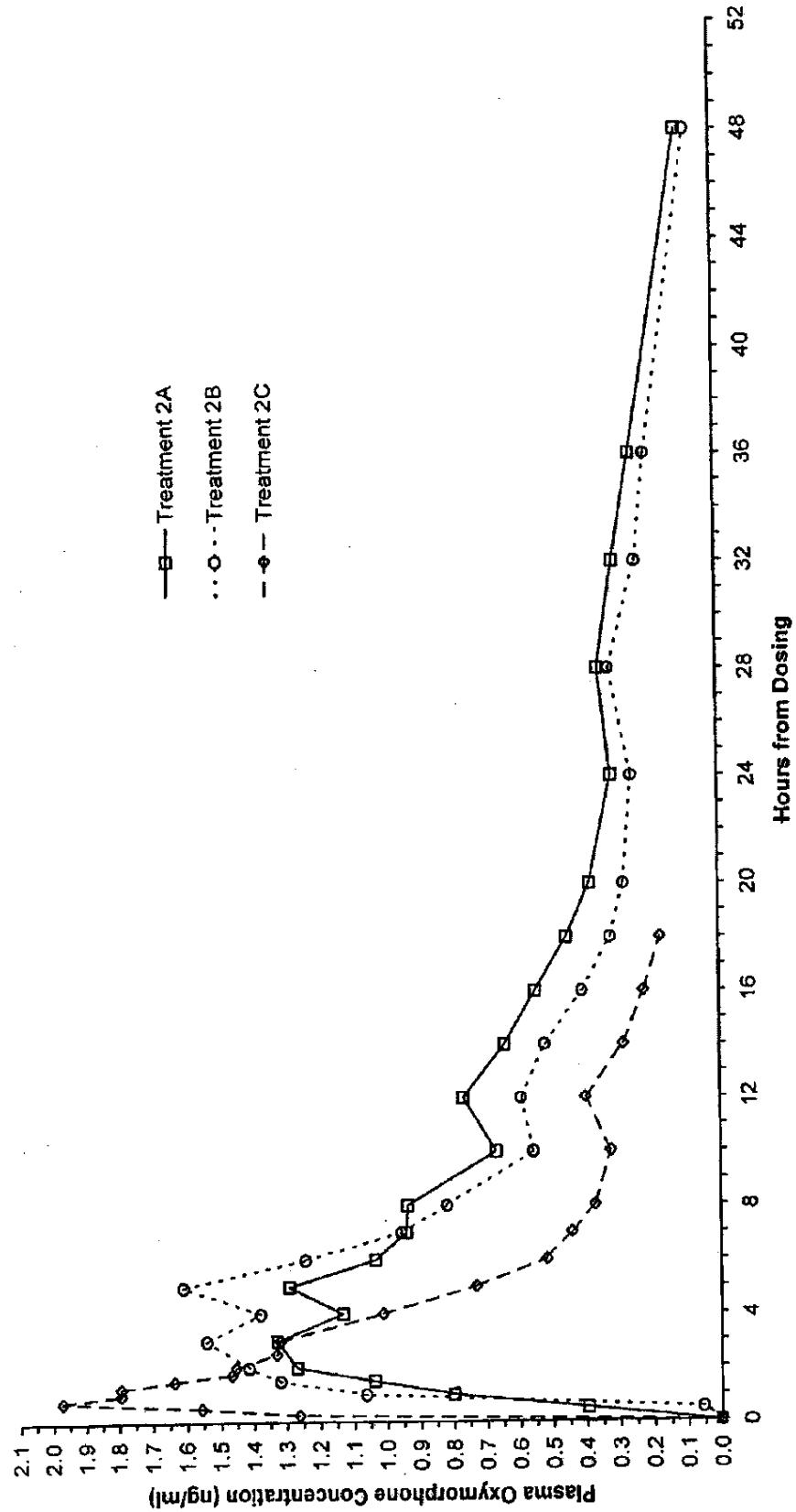


Figure 6

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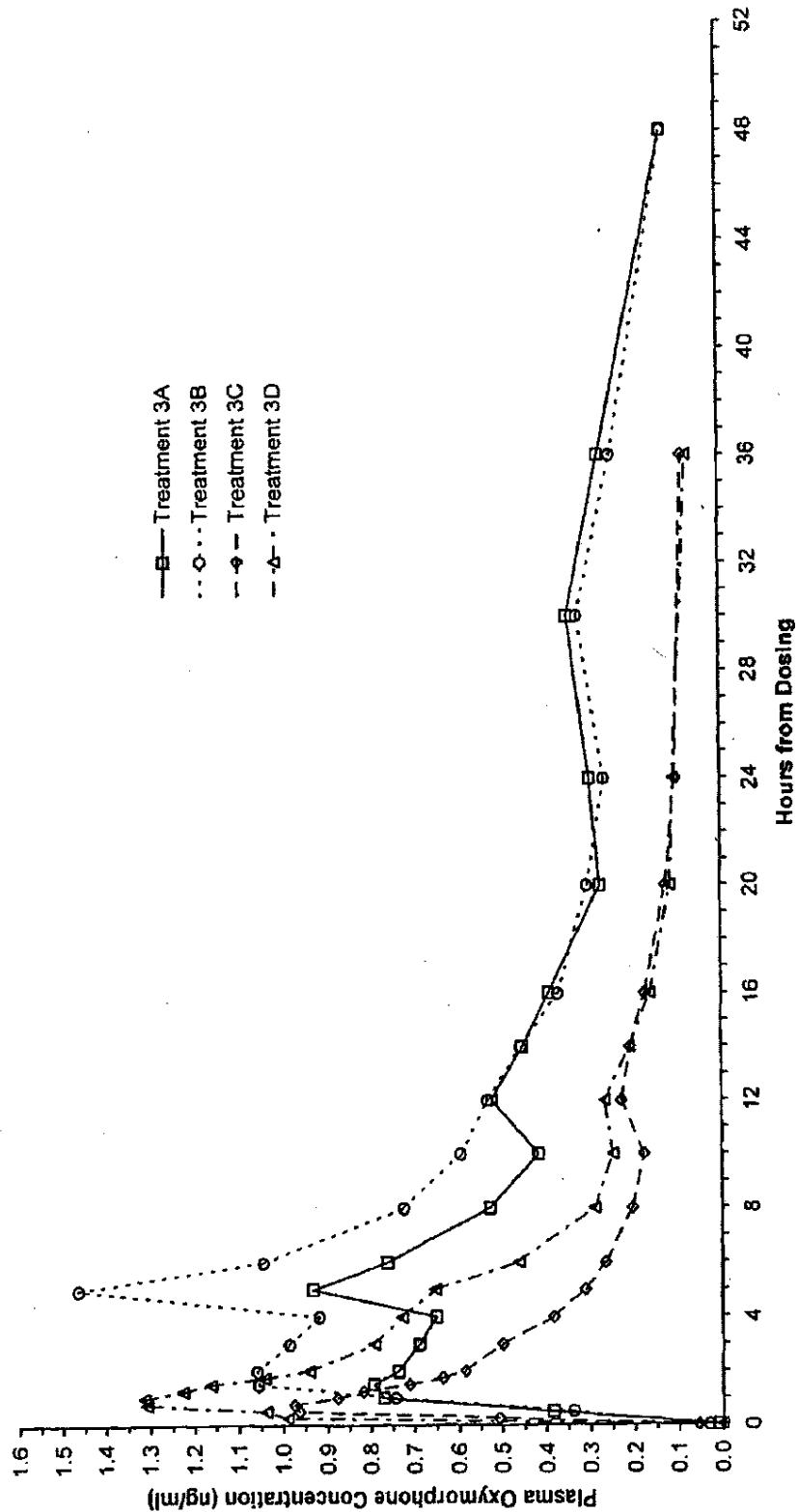


Figure 7

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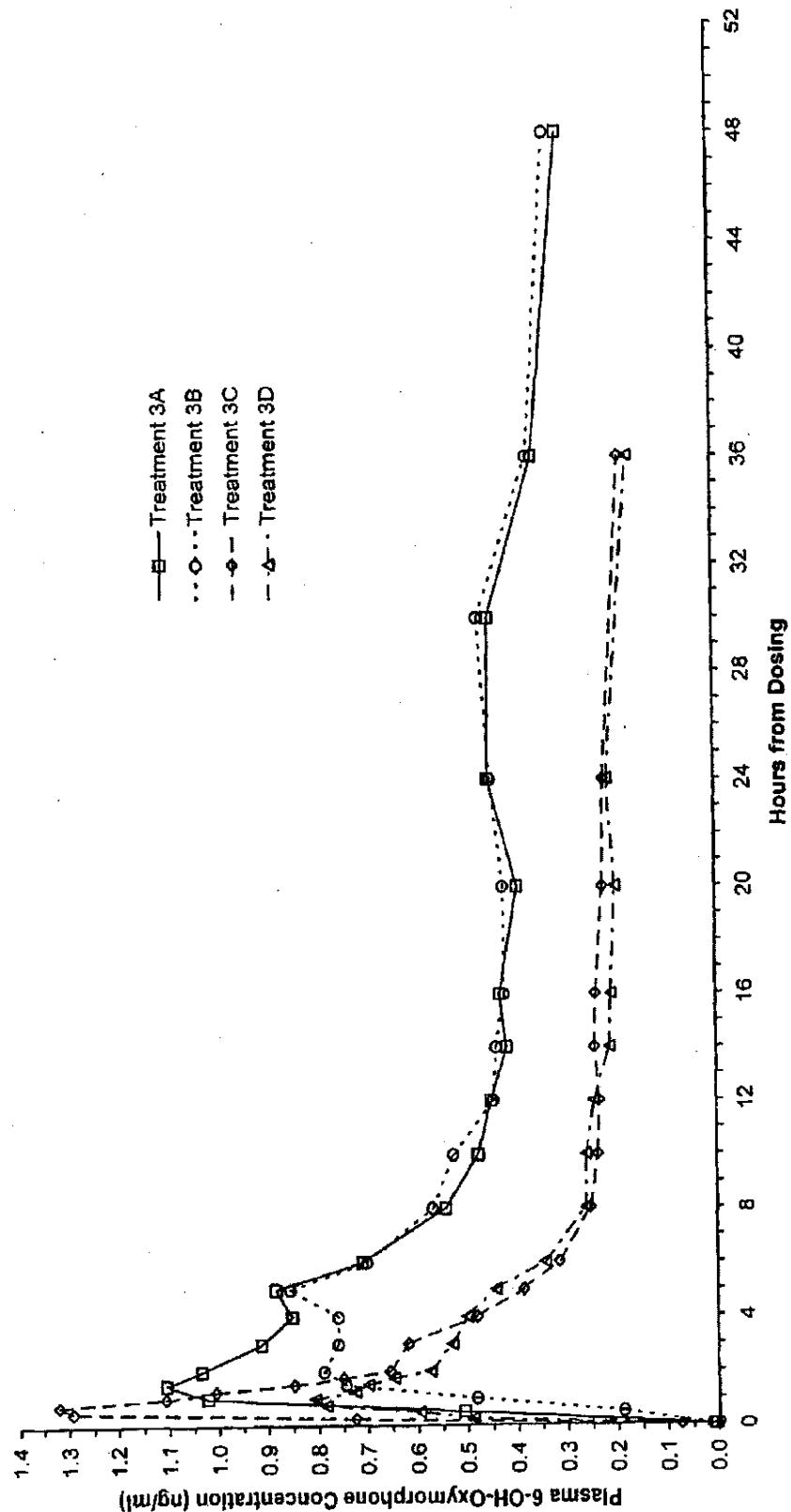


Figure 8

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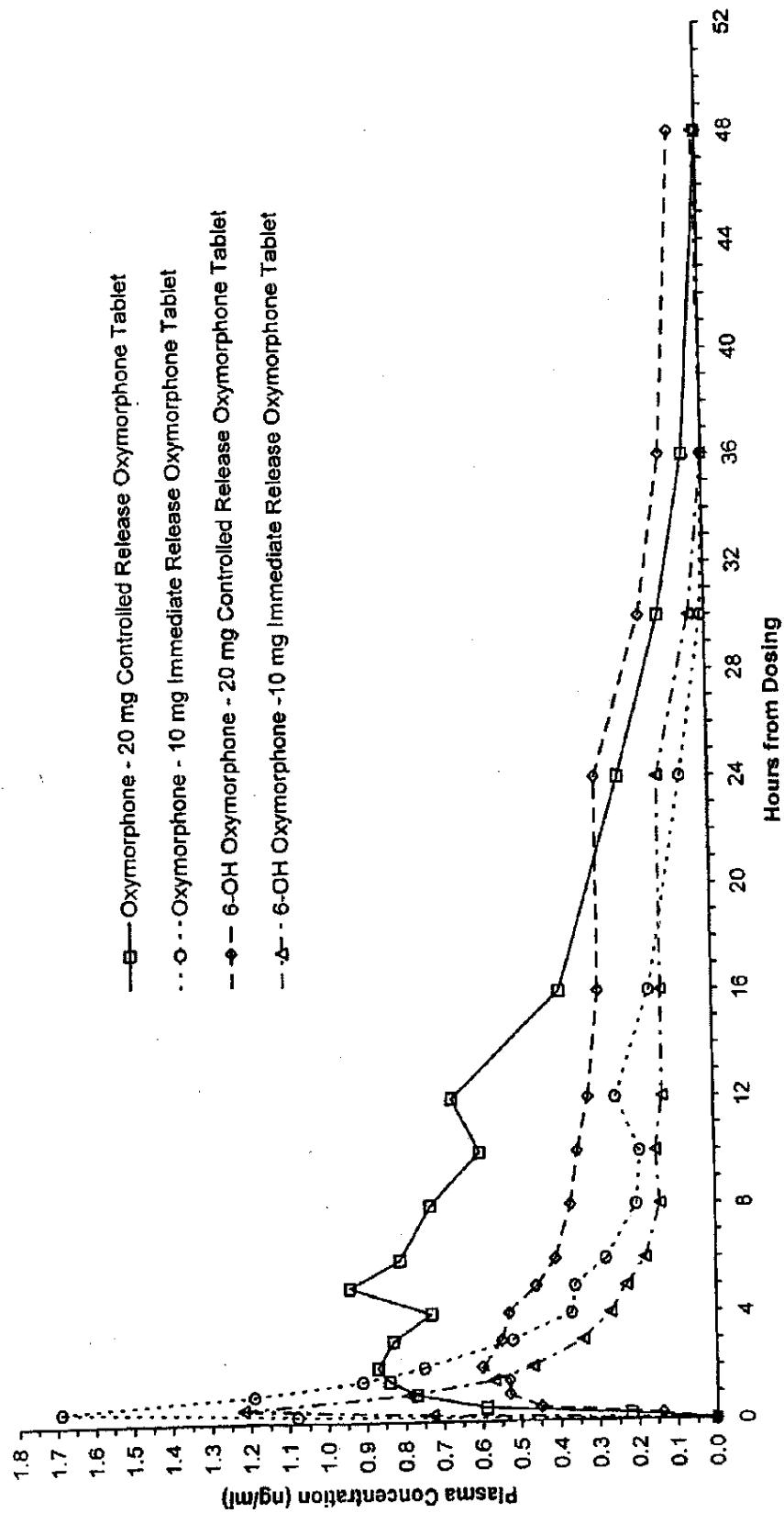


Figure 9

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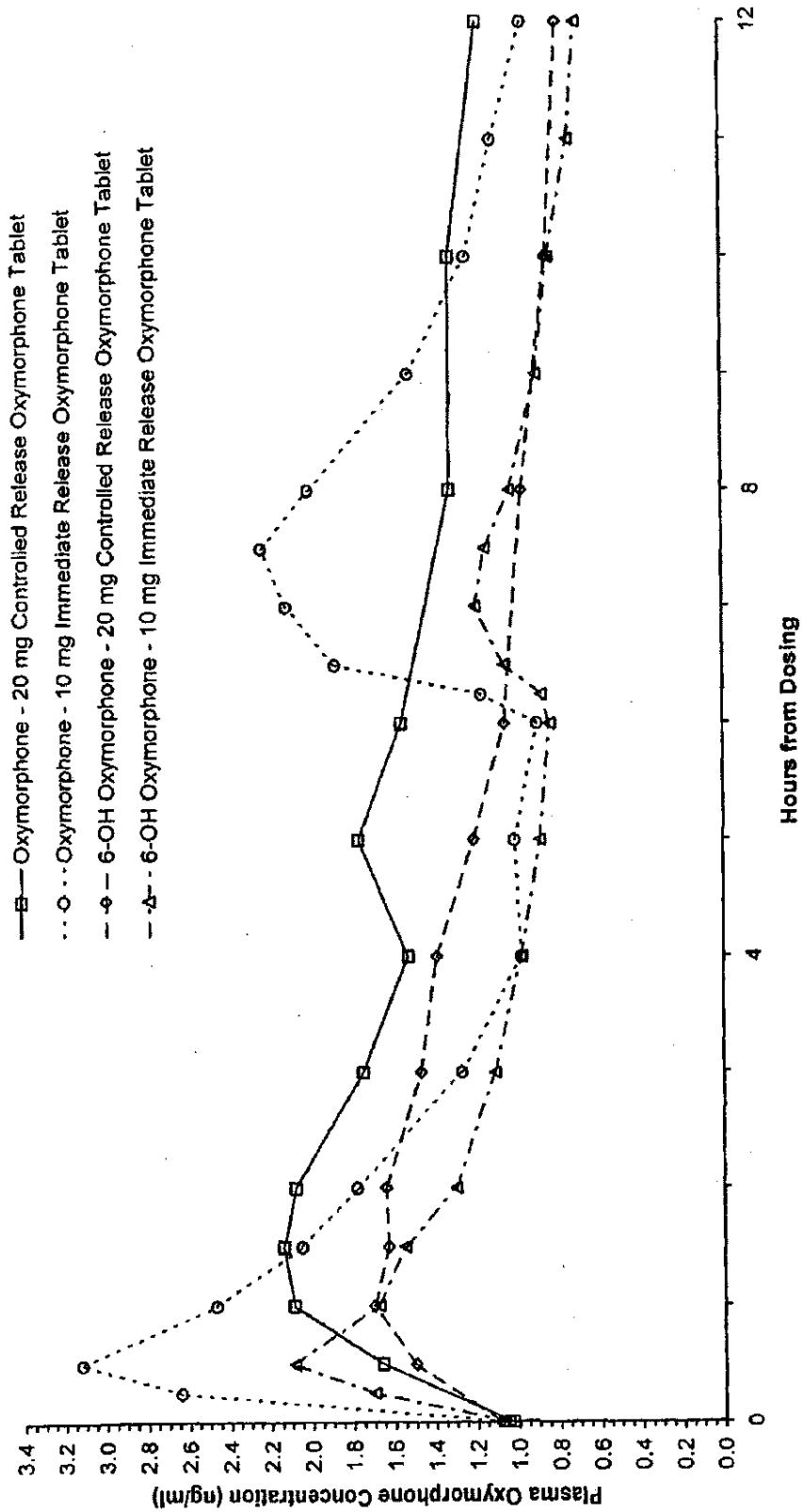


Figure 10

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OXYMORPHONE CONTROLLED RELEASE FORMULATIONS

RELATED APPLICATIONS

This application is a continuation of U.S. patent application Ser. No. 10/190,192 filed Jul. 3, 2002 and claims priority to U.S. Provisional Patent Application Ser. Nos. 60/329,445 filed Oct. 15, 2001, 60/329,432 filed Oct. 15, 2001, 60/303, 357 filed Jul. 6, 2001, and 60/329,444 filed Oct. 15, 2001, which are incorporated herein by reference to the extent permitted by law.

BACKGROUND OF THE INVENTION

Pain is the most frequently reported symptom and it is a common clinical problem which confronts the clinician. Many millions of people in the USA suffer from severe pain that, according to numerous recent reports, is chronically undertreated or inappropriately managed. The clinical usefulness of the analgesic properties of opioids has been recognized for centuries, and morphine and its derivatives have been widely employed for analgesia for decades in a variety of clinical pain states.

Oxymorphone HCl (14-hydroxydihydromorphinone hydrochloride) is a semi-synthetic phenanthrene-derivative opioid agonist, widely used in the treatment of acute and chronic pain, with analgesic efficacy comparable to other opioid analgesics. Oxymorphone is currently marketed as an injection (1 mg/ml in 1 ml ampules; 1.5 mg/ml in 1 ml ampules; 1.5 mg/ml in 10 ml multiple dose vials) for intramuscular, subcutaneous, and intravenous administration, and as 5 mg rectal suppositories. At one time, 2 mg, 5 mg and 10 mg oral immediate release (IR) tablet formulations of oxymorphone HCl were marketed. Oxymorphone HCl is metabolized principally in the liver and undergoes conjugation with glucuronic acid and reduction to 6-alpha- and beta-hydroxy epimers.

An important goal of analgesic therapy is to achieve continuous relief of chronic pain. Regular administration of an analgesic is generally required to ensure that the next dose is given before the effects of the previous dose have worn off. Compliance with opioids increases as the required dosing frequency decreases. Non-compliance results in suboptimal pain control and poor quality of life outcomes. (Ferrell B et al. Effects of controlled-release morphine on quality of life for cancer pain. *Oncol. Nur. Forum* 1989; 4:521-26). Scheduled, rather than "as needed" administration of opioids is currently recommended in guidelines for their use in chronic non-malignant pain. Unfortunately, evidence from prior clinical trials and clinical experience suggests that the short duration of action of immediate release oxymorphone would necessitate administration every 4-6 hours in order to maintain optimal levels of analgesia in chronic pain. A controlled release formulation which would allow less frequent dosing of oxymorphone would be useful in pain management.

For instance, a controlled release formulation of morphine has been demonstrated to provide patients fewer interruptions in sleep, reduced dependence on caregivers, improved compliance, enhanced quality of life outcomes, and increased control over the management of pain. In addition, the controlled release formulation of morphine was reported to provide more constant plasma concentration and clinical effects, less frequent peak to trough fluctuations, reduced dosing frequency, and possibly fewer side effects. (Thirlwell M P et al., Pharmacokinetics and clinical efficacy of oral morphine solution and controlled-release morphine tablets in cancer

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patients. *Cancer* 1989; 63:2275-83; Goughnour B R et al., Analgesic response to single and multiple doses of controlled-release morphine tablets and morphine oral solution in cancer patients. *Cancer* 1989; 63:2294-97; Ferrell B. et al., Effects of controlled-release morphine on quality of life for cancer pain. *Oncol. Nur. Forum* 1989; 4:521-26.

There are two factors associated with the metabolism of some drugs that may present problems for their use in controlled release systems. One is the ability of the drug to induce or inhibit enzyme synthesis, which may result in a fluctuating drug blood plasma level with chronic dosing. The other is a fluctuating drug blood level due to intestinal (or other tissue) metabolism or through a hepatic first-pass effect.

Oxymorphone is metabolized principally in the liver, resulting in an oral bioavailability of about 10%. Evidence from clinical experience suggests that the short duration of action of immediate release oxymorphone necessitates a four hour dosing schedule to maintain optimal levels of analgesia.

It would be useful to clinicians and patients alike to have controlled release dosage forms of oxymorphone to use to treat pain and a method of treating pain using the dosage forms.

SUMMARY OF THE INVENTION

The present invention provides methods for relieving pain by administering a controlled release pharmaceutical tablet containing oxymorphone which produces at least a predetermined minimum blood plasma level for at least 12 hours after dosing, as well as tablets that produce the sustained pain relief over this time period.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 is a pharmacokinetic profile for 6-hydroxy oxymorphone with PID scores.

FIG. 2 is a pharmacokinetic profile for oxymorphone with PID scores.

FIG. 3 is a pharmacokinetic profile for 6-hydroxy oxymorphone with categorical pain scores.

FIG. 4 is a pharmacokinetic profile for oxymorphone with categorical pain scores.

FIG. 5 is a graph of the mean blood plasma concentration of oxymorphone versus time for clinical study 1.

FIG. 6 is a graph of the mean blood plasma concentration of oxymorphone versus time for clinical study 2.

FIG. 7 is a graph of the mean blood plasma concentration of oxymorphone versus time for clinical study 3.

FIG. 8 is a graph of the mean blood plasma concentration of 6-hydroxy oxymorphone versus time for clinical study 3.

FIG. 9 is a graph of the mean blood plasma concentration of oxymorphone for immediate and controlled release tablets from a single dose study.

FIG. 10 is a graph of the mean blood plasma concentration of oxymorphone for immediate and controlled release tablets from a steady state study.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides methods for alleviating pain for 12 to 24 hours using a single dose of a pharmaceutical composition by producing a blood plasma level of oxymorphone and/or 6-OH oxymorphone of at least a minimum value for at least 12 hours or more. As used herein, the terms "6-OH oxymorphone" and "6-hydroxy oxymorphone" are interchangeable and refer to the analog of oxymorphone hav-

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ing an alcohol (hydroxy) moiety that replaces the carboxy moiety found on oxymorphone at the 6-position.

To overcome the difficulties associated with a 4-6 hourly dosing frequency of oxymorphone, this invention provides an oxymorphone controlled release oral solid dosage form, comprising a therapeutically effective amount of oxymorphone or a pharmaceutically acceptable salt of oxymorphone. It has been found that the decreased rate of release of oxymorphone from the oral controlled release formulation of this invention does not substantially decrease the bioavailability of the drug as compared to the same dose of a solution of oxymorphone administered orally. The bioavailability is sufficiently high and the release rate is such that a sufficient plasma level of oxymorphone and/or 6-OH oxymorphone is maintained to allow the controlled release dosage to be used to treat patients suffering moderate to severe pain with once or twice daily dosing. The dosing form of the present invention can also be used with thrice daily dosing.

It is critical when considering the present invention that the difference between a controlled release tablet and an immediate release formulation be fully understood. In classical terms, an immediate release formulation releases at least 80% of its active pharmaceutical ingredient within 30 minutes. With reference to the present invention, the definition of an immediate release formulation will be broadened further to include a formulation which releases more than about 80% of its active pharmaceutical ingredient within 60 minutes in a standard USP Paddle Method dissolution test at 50 rpm in 500 ml media having a pH of between 1.2 and 6.8 at 37° C. "Controlled release" formulations, as referred to herein, will then encompass any formulations which release no more than about 80% of their active pharmaceutical ingredients within 60 minutes under the same conditions.

The controlled release dosage form of this invention exhibits a dissolution rate *in vitro*, when measured by USP Paddle Method at 50 rpm in 500 ml media having a pH between 1.2 and 6.8 at 37° C., of about 15% to about 50% by weight oxymorphone released after 1 hour, about 45% to about 80% by weight oxymorphone released after 4 hours, and at least about 80% by weight oxymorphone released after 10 hours.

When administered orally to humans, an effective controlled release dosage form of oxymorphone should exhibit the following *in vivo* characteristics: (a) peak plasma level of oxymorphone occurs within about 1 to about 8 hours after administration; (b) peak plasma level of 6-OH oxymorphone occurs within about 1 to about 8 hours after administration; (c) duration of analgesic effect is through about 8 to about 24 hours after administration; (d) relative oxymorphone bioavailability is in the range of about 0.5 to about 1.5 compared to an orally-administered aqueous solution of oxymorphone; and (e) the ratio of the area under the curve of blood plasma level vs. time for 6-OH oxymorphone compared to oxymorphone is in the range of about 0.5 to about 1.5. Of course, there is variation of these parameters among subjects, depending on the size and weight of the individual subject, the subject's age, individual metabolism differences, and other factors. Indeed, the parameters may vary in an individual from day to day. Accordingly, the parameters set forth above are intended to be mean values from a sufficiently large study so as to minimize the effect of individual variation in arriving at the values. A convenient method for arriving at such values is by conducting a study in accordance with standard FDA procedures such as those employed in producing results for use in a new drug application (or abbreviated new drug application) before the FDA. Any reference to mean values herein, in conjunction with desired results, refer to results from such a study, or some comparable study. Reference to mean values

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reported herein for studies actually conducted are arrived at using standard statistical methods as would be employed by one skilled in the art of pharmaceutical formulation and testing for regulatory approval.

In one specific embodiment of the controlled release matrix form of the invention, the oxymorphone or salt of oxymorphone is dispersed in a controlled release delivery system that comprises a hydrophilic material which, upon exposure to gastrointestinal fluid, forms a gel matrix that releases oxymorphone at a controlled rate. The rate of release of oxymorphone from the matrix depends on the drug's partition coefficient between components of the matrix and the aqueous phase within the gastrointestinal tract. In a preferred form of this embodiment, the hydrophilic material of the controlled release delivery system comprises a mixture of a heteropolysaccharide gum and an agent capable of cross-linking the heteropolysaccharide in presence of gastrointestinal fluid. The controlled release delivery system may also comprise a water-soluble pharmaceutical diluent mixed with the hydrophilic material. Preferably, the cross-linking agent is a homopolysaccharide gum and the inert pharmaceutical diluent is a monosaccharide, a disaccharide, or a polyhydric alcohol, or a mixture thereof.

In a specific preferred embodiment, the appropriate blood plasma levels of oxymorphone and 6-hydroxy oxymorphone are achieved using oxymorphone in the form of oxymorphone hydrochloride, wherein the weight ratio of heteropolysaccharide to homopolysaccharide is in the range of about 1:3 to about 3:1, the weight ratio of heteropolysaccharide to diluent is in the range of about 1:8 to about 8:1, and the weight ratio of heteropolysaccharide to oxymorphone hydrochloride is in the range of about 10:1 to about 1:10. A preferred heteropolysaccharide is xanthan gum and a preferred homopolysaccharide is locust bean gum. The dosage form also comprises a cationic cross-linking agent and a hydrophobic polymer. In the preferred embodiment, the dosage form is a tablet containing about 5 mg to about 80 mg of oxymorphone hydrochloride. In a most preferred embodiment, the tablet contains about 20 mg oxymorphone hydrochloride.

The invention includes a method which comprises achieving appropriate blood plasma levels of drug while providing extended pain relief by administering one to three times per day to a patient suffering moderate to severe, acute or chronic pain, an oxymorphone controlled release oral solid dosage form of the invention in an amount sufficient to alleviate the pain for a period of about 8 hours to about 24 hours. This type and intensity of pain is often associated with cancer, autoimmune diseases, infections, surgical and accidental traumas and osteoarthritis.

The invention also includes a method of making an oxymorphone controlled release oral solid dosage form of the invention which comprises mixing particles of oxymorphone or a pharmaceutically acceptable salt of oxymorphone with granules comprising the controlled release delivery system, preferably followed by directly compressing the mixture to form tablets.

Pharmaceutically acceptable salts of oxymorphone which can be used in this invention include salts with the inorganic and organic acids which are commonly used to produce non-toxic salts of medicinal agents. Illustrative examples would be those salts formed by mixing oxymorphone with hydrochloric, sulfuric, nitric, phosphoric, phosphorous, hydrobromic, maleic, malic, ascorbic, citric or tartaric, pamoic, lauric, stearic, palmitic, oleic, myristic, lauryl sulfuric, naphthylene-sulfonic, linoleic or linolenic acid, and the like. The hydrochloride salt is preferred.

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It has now been found that 6-OH oxymorphone, which is one of the metabolites of oxymorphone, may play a role in alleviating pain. When oxymorphone is ingested, part of the dosage gets into the bloodstream to provide pain relief, while another part is metabolized to 6-OH oxymorphone. This metabolite then enters the bloodstream to provide further pain relief. Thus it is believed that both the oxymorphone and 6-hydroxyoxymorphone levels are important to pain relief.

The effectiveness of oxymorphone and 6-hydroxyoxymorphone at relieving pain and the pharmacokinetics of a single dose of oxymorphone were studied. The blood plasma levels of both oxymorphone and 6-hydroxyoxymorphone were measured in patients after a single dose of oxymorphone was administered. Similarly, the pain levels in patients were measured after a single administration of oxymorphone to determine the effective duration of pain relief from a single dose. FIGS. 1-2 show the results of these tests, comparing pain levels to oxymorphone and 6-hydroxy oxymorphone levels.

For these tests, pain was measured using a Visual Analog Scale (VAS) or a Categorical Scale. The VAS scales consisted of a horizontal line, 100 mm in length. The left-hand end of the scale (0 mm) was marked with the descriptor "No Pain" and the right-hand end of the scale (100 mm) was marked with the descriptor "Extreme Pain". Patients indicated their level of pain by making a vertical mark on the line. The VAS score was equal to the distance (in mm) from the left-hand end of the scale to the patient's mark. For the categorical scale, patients completed the following statement, "My pain at this time is" using the scale None=0, Mild=1, Moderate=2, or Severe=3.

As can be seen from these figures, there is a correlation between pain relief and both oxymorphone and 6-hydroxyoxymorphone levels. As the blood plasma levels of oxymorphone and 6-hydroxyoxymorphone increase, pain decreases (and pain intensity difference and pain relief increases). Thus, to the patient, it is the level of oxymorphone and 6-hydroxyoxymorphone in the blood plasma which is most important. Further it is these levels which dictate the efficacy of the dosage form. A dosage form which maintains a sufficiently high level of oxymorphone or 6-hydroxyoxymorphone for a longer period need not be administered frequently. Such a result is accomplished by embodiments of the present invention.

The oxymorphone controlled release oral solid dosage form of this invention can be made using any of several different techniques for producing controlled release oral solid dosage forms of opioid analgesics.

In one embodiment, a core comprising oxymorphone or oxymorphone salt is coated with a controlled release film which comprises a water insoluble material and which upon exposure to gastrointestinal fluid releases oxymorphone from the core at a controlled rate. In a second embodiment, the oxymorphone or oxymorphone salt is dispersed in a controlled release delivery system that comprises a hydrophilic material which upon exposure to gastrointestinal fluid forms a gel matrix that releases oxymorphone at a controlled rate. A third embodiment is a combination of the first two: a controlled release matrix coated with a controlled release film. In a fourth embodiment the oxymorphone is incorporated into an osmotic pump. In any of these embodiments, the dosage form can be a tablet, a plurality of granules in a capsule, or other suitable form, and can contain lubricants, colorants, diluents, and other conventional ingredients.

Osmotic Pump

An osmotic pump comprises a shell defining an interior compartment and having an outlet passing through the shell. The interior compartment contains the active pharmaceutical

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ingredient. Generally the active pharmaceutical ingredient is mixed with excipients or other compositions such as a polyalkylene. The shell is generally made, at least in part, from a material (such as cellulose acetate) permeable to the liquid of the environment where the pump will be used, usually stomach acid. Once ingested, the pump operates when liquid diffuses through the shell of the pump. The liquid dissolves the composition to produce a saturated situation. As more liquid diffuses into the pump, the saturated solution containing the pharmaceutical is expelled from the pump through the outlet. This produces a nearly constant release of active ingredient, in the present case, oxymorphone.

Controlled Release Coating

In this embodiment, a core comprising oxymorphone or oxymorphone salt is coated with a controlled release film which comprises a water insoluble material. The film can be applied by spraying an aqueous dispersion of the water insoluble material onto the core. Suitable water insoluble materials include alkyl celluloses, acrylic polymers, waxes (alone or in admixture with fatty alcohols), shellac and zein. The aqueous dispersions of alkyl celluloses and acrylic polymers preferably contain a plasticizer such as triethyl citrate, dibutyl phthalate, propylene glycol, and polyethylene glycol. The film coat can contain a water-soluble material such as polyvinylpyrrolidone (PVP) or hydroxypropylmethylcellulose (HPMC).

The core can be a granule made, for example, by wet granulation of mixed powders of oxymorphone or oxymorphone salt and a binding agent such as HPMC, or by coating an inert bead with oxymorphone or oxymorphone salt and a binding agent such as HPMC, or by spheronising mixed powders of oxymorphone or oxymorphone salt and a spheronising agent such as microcrystalline cellulose. The core can be a tablet made by compressing such granules or by compressing a powder comprising oxymorphone or oxymorphone salt.

The in vitro and in vivo release characteristics of this controlled release dosage form can be modified by using mixtures of different water insoluble and water soluble materials, using different plasticizers, varying the thickness of the controlled release film, including release-modifying agents in the coating, or by providing passageways through the coating.

Controlled Release Matrix

It is important in the present invention that appropriate blood plasma levels of oxymorphone and 6-hydroxy oxymorphone be achieved and maintained for sufficient time to provide pain relief to a patient for a period of 12 to 24 hours. The preferred composition for achieving and maintaining the proper blood plasma levels is a controlled-release matrix. In this embodiment, the oxymorphone or oxymorphone salt is dispersed in a controlled release delivery system that comprises a hydrophilic material (gelling agent) which upon exposure to gastrointestinal fluid forms a gel matrix that releases oxymorphone at a controlled rate. Such hydrophilic materials include gums, cellulose ethers, acrylic resins, and protein-derived materials. Suitable cellulose ethers include hydroxyalkyl celluloses and carboxyalkyl celluloses, especially hydroxyethyl cellulose (HEC), hydroxypropyl cellulose (HPC), HPMC, and carboxy methylcellulose (CMC). Suitable acrylic resins include polymers and copolymers of acrylic acid, methacrylic acid, methyl acrylate and methyl methacrylate. Suitable gums include heteropolysaccharide and homopolysaccharide gums, e.g., xanthan, tragacanth, acacia, karaya, alginates, agar, guar, hydroxypropyl guar, carageenan, and locust bean gums.

Preferably, the controlled release tablet of the present invention is formed from (I) a hydrophilic material comprising (a) a heteropolysaccharide; or (b) a heteropolysaccharide

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and a cross-linking agent capable of cross-linking said heteropolysaccharide; or (c) a mixture of (a), (b) and a polysaccharide gum; and (II) an inert pharmaceutical filler comprising up to about 80% by weight of the tablet; and (III) oxymorphone.

The term "heteropolysaccharide" as used herein is defined as a water-soluble polysaccharide containing two or more kinds of sugar units, the heteropolysaccharide having a branched or helical configuration, and having excellent water-wicking properties and immense thickening properties.

A preferred heteropolysaccharide is xanthan gum, which is a high molecular weight ($>10^6$) heteropolysaccharide. Other preferred heteropolysaccharides include derivatives of xanthan gum, such as deacylated xanthan gum, the carboxymethyl ether, and the propylene glycol ester.

The cross linking agents used in the controlled release embodiment of the present invention which are capable of cross-linking with the heteropolysaccharide include homopolysaccharide gums such as the galactomannans, i.e., polysaccharides which are composed solely of mannose and galactose. Galactomannans which have higher proportions of unsubstituted mannose regions have been found to achieve more interaction with the heteropolysaccharide. Locust bean gum, which has a higher ratio of mannose to the galactose, is especially preferred as compared to other galactomannans such as guar and hydroxypropyl guar.

Preferably, the ratio of heteropolysaccharide to homopolysaccharide is in the range of about 1:9 to about 9:1, preferably about 1:3 to about 3:1. Most preferably, the ratio of xanthan gum to polysaccharide material (i.e., locust bean gum, etc.) is preferably about 1:1.

In addition to the hydrophilic material, the controlled release delivery system can also contain an inert pharmaceutical diluent such as a monosaccharide, a disaccharide, a polyhydric alcohol and mixtures thereof. The ratio of diluent to hydrophilic matrix-forming material is generally in the range of about 1:3 to about 3:1.

The controlled release properties of the controlled release embodiment of the present invention may be optimized when the ratio of heteropolysaccharide gum to homopolysaccharide material is about 1:1, although heteropolysaccharide gum in an amount of from about 20 to about 80% or more by weight of the heterodisperse polysaccharide material provides an acceptable slow release product. The combination of any homopolysaccharide gums known to produce a synergistic effect when exposed to aqueous solutions may be used in accordance with the present invention. It is also possible that the type of synergism which is present with regard to the gum combination of the present invention could also occur between two homogeneous or two heteropolysaccharides. Other acceptable gelling agents which may be used in the present invention include those gelling agents well-known in the art. Examples include vegetable gums such as alginates, carrageenan, pectin, guar gum, xanthan gum, modified starch, hydroxypropylmethylcellulose, methylcellulose, and other cellulosic materials such as sodium carboxymethylcellulose and hydroxypropyl cellulose. This list is not meant to be exclusive.

The combination of xanthan gum with locust bean gum with or without the other homopolysaccharide gums is an especially preferred gelling agent. The chemistry of certain of the ingredients comprising the excipients of the present invention such as xanthan gum is such that the excipients are considered to be self-buffering agents which are substantially

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insensitive to the solubility of the medicament and likewise insensitive to the pH changes along the length of the gastrointestinal tract.

The inert filler of the sustained release excipient preferably comprises a pharmaceutically acceptable saccharide, including a monosaccharide, a disaccharide, or a polyhydric alcohol, and/or mixtures of any of the foregoing. Examples of suitable inert pharmaceutical fillers include sucrose, dextrose, lactose, microcrystalline cellulose, fructose, xylitol, sorbitol, mixtures thereof and the like. However, it is preferred that a soluble pharmaceutical filler such as lactose, dextrose, sucrose, or mixtures thereof be used.

The cationic cross-linking agent which is optionally used in conjunction with the controlled release embodiment of the present invention may be monovalent or multivalent metal cations. The preferred salts are the inorganic salts, including various alkali metal and/or alkaline earth metal sulfates, chlorides, borates, bromides, citrates, acetates, lactates, etc. Specific examples of suitable cationic cross-linking agents include calcium sulfate, sodium chloride, potassium sulfate, sodium carbonate, lithium chloride, tripotassium phosphate, sodium borate, potassium bromide, potassium fluoride, sodium bicarbonate, calcium chloride, magnesium chloride, sodium citrate, sodium acetate, calcium lactate, magnesium sulfate and sodium fluoride. Multivalent metal cations may also be utilized. However, the preferred cationic cross-linking agents are bivalent. Particularly preferred salts are calcium sulfate and sodium chloride. The cationic cross-linking agents of the present invention are added in an amount effective to obtain a desirable increased gel strength due to the cross-linking of the gelling agent (e.g., the heteropolysaccharide and homopolysaccharide gums). In preferred embodiments, the cationic cross-linking agent is included in the sustained release excipient of the present invention in an amount from about 1 to about 20% by weight of the sustained release excipient, and in an amount about 0.5% to about 16% by weight of the final dosage form.

In the controlled release embodiments of the present invention, the sustained release excipient comprises from about 10 to about 99% by weight of a gelling agent comprising a heteropolysaccharide gum and a homopolysaccharide gum, from about 1 to about 20% by weight of a cationic crosslinking agent, and from about 0 to about 89% by weight of an inert pharmaceutical diluent. In other embodiments, the sustained release excipient comprises from about 10 to about 75% gelling agent, from about 2 to about 15% cationic crosslinking agent, and from about 30 to about 75% inert diluent. In yet other embodiments, the sustained release excipient comprises from about 30 to about 75% gelling agent, from about 5 to about 10% cationic cross-linking agent, and from about 15 to about 65% inert diluent.

The sustained release excipient used in this embodiment of the present invention (with or without the optional cationic cross-linking agent) may be further modified by incorporation of a hydrophobic material which slows the hydration of the gums without disrupting the hydrophilic matrix. This is accomplished in preferred embodiments of the present invention by granulating the sustained release excipient with the solution or dispersion of a hydrophobic material prior to the incorporation of the medicament. The hydrophobic polymer may be selected from an alkylcellulose such as ethylcellulose, other hydrophobic cellulosic materials, polymers or copolymers derived from acrylic or methacrylic acid esters, copolymers of acrylic and methacrylic acid esters, zein, waxes, shellac, hydrogenated vegetable oils, and any other pharmaceutically acceptable hydrophobic material known to those skilled in the art. The amount of hydrophobic material incor-

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porated into the sustained release excipient is that which is effective to slow the hydration of the gums without disrupting the hydrophilic matrix formed upon exposure to an environmental fluid. In certain preferred embodiments of the present invention, the hydrophobic material is included in the sustained release excipient in an amount from about 1 to about 20% by weight. The solvent for the hydrophobic material may be an aqueous or organic solvent, or mixtures thereof.

Examples of commercially available alkylcelluloses are Aquacoat coating (aqueous dispersion of ethylcellulose available from FMC of Philadelphia, Pa.) and Surelease coating (aqueous dispersion of ethylcellulose available from Colordon of West Point, Pa.). Examples of commercially available acrylic polymers suitable for use as the hydrophobic material include Eudragit RS and RL polymers (copolymers of acrylic and methacrylic acid esters having a low content (e.g., 1:20 or 1:40) of quaternary ammonium compounds available from Rohm America of Piscataway, N.J.).

The controlled release matrix useful in the present invention may also contain a cationic cross-linking agent such as calcium sulfate in an amount sufficient to cross-link the gelling agent and increase the gel strength, and an inert hydrophobic material such as ethyl cellulose in an amount sufficient to slow the hydration of the hydrophilic material without disrupting it. Preferably, the controlled release delivery system is prepared as a pre-manufactured granulation.

EXAMPLES

Example 1

Two controlled release delivery systems are prepared by dry blending xanthan gum, locust bean gum, calcium sulfate dehydrate, and dextrose in a high speed mixed/granulator for 3 minutes. A slurry is prepared by mixing ethyl cellulose with alcohol. While running choppers/impellers, the slurry is added to the dry blended mixture, and granulated for another 3 minutes. The granulation is then dried to a LOD (loss on drying) of less than about 10% by weight. The granulation is then milled using 20 mesh screen. The relative quantities of the ingredients are listed in the table below.

TABLE 1

Excipient	Controlled Release Delivery System	
	Formulation 1 (%)	Formulation 2 (%)
Locust Bean Gum, FCC	25.0	30.0
Xanthan Gum, NF	25.0	30.0
Dextrose, USP	35.0	40.0
Calcium Sulfate Dihydrate, NF	10.0	0.0
Ethylcellulose, NF	5.0	0.0
Alcohol, SD3A (Anhydrous)	(10) ¹	(20.0) ¹
Total	100.0	100.0

A series of tablets containing different amounts of oxymorphone hydrochloride were prepared using the controlled release delivery Formulation 1 shown in Table 1. The quantities of ingredients per tablet are as listed in the following table.

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TABLE 2

Component	Sample Tablets of Differing Strengths				
	Amounts in Tablet (mg)				
Oxymorphone HCl, USP (mg)	5	10	20	40	80
Controlled release delivery system	160	160	160	160	160
Silicified microcrystalline cellulose, N.F.	20	20	20	20	20
Sodium stearyl fumarate, NF	2	2	2	2	2
Total weight	187	192	202	222	262
Opadry (colored)	7.48	7.68	8.08	8.88	10.48
Opadry (clear)	0.94	0.96	1.01	1.11	1.31

Examples 2 and 3

Two batches of 20 mg tablets were prepared as described above, using the controlled release delivery system of Formulation 1. One batch was formulated to provide relatively fast controlled release, the other batch was formulated to provide relatively slow controlled release. Compositions of the tablets are shown in the following table.

TABLE 3

Ingredients	Slow and Fast Release Compositions		
	Example 2 Slow (mg)	Example 3 Fast (mg)	Example 4 Fast (mg)
Oxymorphone HCl, USP	20	20	20
Controlled Release Delivery System	360	160	160
Silicified Microcrystalline Cellulose, NF	20	20	20
Sodium stearyl fumarate, NF	4	2	2
Total weight	404	202	202
Coating (color or clear)	12	12	9

The tablets of Examples 2, 3, and 4 were tested for in vitro release rate according to USP Procedure Drug Release U.S. Pat. No. 23. Release rate is a critical variable in attempting to control the blood plasma levels of oxymorphone and 6-hydroxyoxymorphone in a patient. Results are shown in the following Table 4.

TABLE 4

Time (hr)	Release Rates of Slow and Fast Release Tablets		
	Example 2 (Slow Release)	Example 3 (Fast Release)	Example 4 (Fast Release)
0.5	18.8	21.3	20.1
1	27.8	32.3	31.7
2	40.5	47.4	46.9
3	50.2	58.5	57.9
4	58.1	66.9	66.3
5	64.7	73.5	74.0
6	70.2	78.6	83.1
8	79.0	86.0	92.0
10	85.3	90.6	95.8
12	89.8	93.4	97.3

Clinical Studies

Three clinical studies were conducted to assess the bioavailability (rate and extent of absorption) of oxymorphone. Study 1 addressed the relative rates of absorption of con-

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trolled release (CR) oxymorphone tablets (of Examples 2 and 3) and oral oxymorphone solution in fasted patients. Study 2 addressed the relative rates of absorption of CR oxymorphone tablets (of Examples 2 and 3) and oral oxymorphone solution in fed patients. Study 3 addressed the relative rates of absorption of CR oxymorphone tablets (of Example 4) and oral oxymorphone solution in fed and fasted patients.

The blood plasma levels set forth herein as appropriate to achieve the objects of the present invention are mean blood plasma levels. As an example, if the blood plasma level of oxymorphone in a patient 12 hours after administration of a tablet is said to be at least 0.5 ng/ml, any particular individual may have lower blood plasma levels after 12 hours. However, the mean minimum concentration should meet the limitation set forth. To determine mean parameters, a study should be performed with a minimum of 8 adult subjects, in a manner acceptable for filing an application for drug approval with the US Food and Drug Administration. In cases where large fluctuations are found among patients, further testing may be necessary to accurately determine mean values.

For all studies, the following procedures were followed, unless otherwise specified for a particular study.

The subjects were not to consume any alcohol-, caffeine-, or xanthine-containing foods or beverages for 24 hours prior to receiving study medication for each study period. Subjects were to be nicotine and tobacco free for at least 6 months prior to enrolling in the study. In addition, over-the-counter medications were prohibited 7 days prior to dosing and during the study. Prescription medications were not allowed 14 days prior to dosing and during the study.

Pharmacokinetic and Statistical Methods

The following pharmacokinetic parameters were computed from the plasma oxymorphone concentration-time data:

$AUC_{(0-\infty)}$	Area under the drug concentration-time curve from time zero to the time of the last quantifiable concentration (C_t), calculated using linear trapezoidal summation.
$AUC_{(0-\infty)}$	Area under the drug concentration-time curve from time zero to infinity: $AUC_{(0-\infty)} = AUC_{(0-\infty)} + C_t/K_{el}$, where K_{el} is the terminal elimination rate constant.
$AUC_{(0-24)}$	Partial area under the drug concentration-time curve from time zero to 24 hours.
C_{max}	Maximum observed drug concentration.
T_{max}	Time of the observed maximum drug concentration.
K_{el}	Elimination rate constant based on the linear regression of the terminal linear portion of the LN(concentration) time curve.

Terminal elimination rate constants for use in the above calculations were in turn computed using linear regression of a minimum of three time points, at least two of which were consecutive. K_{el} values for which correlation coefficients were less than or equal to 0.8 were not reported in the pharmacokinetic parameter tables or included in the statistical analysis. Thus $AUC_{(0-\infty)}$ was also not reported in these cases.

A parametric (normal-theory) general linear model was applied to each of the above parameters (excluding T_{max}), and the LN-transformed parameters C_{max} , $AUC_{(0-24)}$, $AUC_{(0-\infty)}$, and $AUC_{(0-\infty)}$. Initially, the analysis of variance (ANOVA) model included the following factors: treatment, sequence, subject within sequence, period, and carryover effect. If carryover effect was not significant, it was dropped from the model. The sequence effect was tested using the subject within sequence mean square, and all other main effects were tested using the residual error (error mean square).

Plasma oxymorphone concentrations were listed by subject at each collection time and summarized using descriptive

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statistics. Pharmacokinetic parameters were also listed by subject and summarized using descriptive statistics.

Study 1—Two Controlled Release Formulations; Fasted Patients

5 Healthy volunteers received a single oral dose of 20 mg CR oxymorphone taken with 240 ml water after a 10-hour fast. Subjects received the tablets of Example 2 (Treatment 1A) or Example 3 (Treatment 1B). Further subjects were given a single oral dose of 10 mg/10 ml oxymorphone solution in 180 ml apple juice followed with 60 ml water (Treatment 1C). The orally dosed solution was used to simulate an immediate release (IR) dose.

This study had a single-center, open-label, randomized, 15 three-way crossover design using fifteen subjects. Subjects were in a fasted state following a 10-hour overnight fast. There was a 14-day washout interval between the three dose administrations. The subjects were confined to the clinic during each study period. Subjects receiving Treatment 1C were confined for 18 hours and subjects receiving Treatments 1A or 1B were confined for 48 hours after dosing. Ten-milliliter blood samples were collected during each study period at the 20 0 hour (predose), and at 0.5, 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 10, 12, 14, 16, 18, 20, 24, 28, 32, 36, and 48 hours postdose for subjects receiving Treatment 1A or 1B and 0, 0.25, 0.5, 0.75, 25 1, 1.25, 1.5, 1.75, 2, 2.5, 3, 4, 5, 6, 7, 8, 10, 12, 14, 16, and 18 hours post-dose. The mean plasma concentration of oxymorphone versus time for each treatment across all subjects is shown in table 5.

TABLE 5

Mean Plasma Concentration vs. Time (ng/ml)			
Time (hr)	Treatment 1A	Treatment 1B	Treatment 1C
35	0	0.000	0.000
	0.25		0.9489
	0.5	0.2941	0.4104
	0.75		1.3016
	1	0.5016	0.7334
	1.25		1.3264
	1.5	0.5951	1.3046
	1.75		1.2041
	2	0.6328	0.8192
	2.5		1.0813
40	3	0.5743	0.9502
	4	0.5709	0.9055
	5	0.7656	0.7161
	6	0.7149	0.6689
	7	0.6334	0.4879
	8	0.5716	0.4184
	10	0.4834	0.3658
	12	0.7333	0.3464
	14	0.6271	0.2610
	16	0.4986	0.2970
45	18	0.4008	0.2083
	20	0.3405	0.1661
	24	0.2736	0.1368
	28	0.3209	0.2270
	32	0.2846	0.2805
	36	0.2583	0.2272
	48	0.0975	0.1903
			0.0792

The results are shown graphically in FIG. 5. In both Table 5 and FIG. 5, the results are normalized to a 20 mg dosage. 60 The immediate release liquid of Treatment 1C shows a classical curve, with a high and relatively narrow peak, followed by an exponential drop in plasma concentration. However, the controlled release oxymorphone tablets exhibit triple peaks in blood plasma concentration. The first peak occurs (on average) at around 3 hours. The second peak of the mean blood plasma concentration is higher than the first, occurring around 6-7 hours, on average).

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Occasionally, in an individual, the first peak is higher than the second, although generally this is not the case. This makes it difficult to determine the time to maximum blood plasma concentration (T_{max}) because if the first peak is higher than the second, maximum blood plasma concentration (C_{max}) occurs much earlier (at around 3 hours) than in the usual case where the second peak is highest. Therefore, when we refer to the time to peak plasma concentration (T_{max}) unless otherwise specified, we refer to the time to the second peak. Further, when reference is made to the second peak, we refer to the time or blood plasma concentration at the point where the blood plasma concentration begins to drop the second time. Generally, where the first peak is higher than the second, the difference in the maximum blood plasma concentration at the two peaks is small. Therefore, this difference (if any) was ignored and the reported C_{max} was the true maximum blood plasma concentration and not the concentration at the second peak.

TABLE 6

Pharmacokinetic Parameters of Plasma Oxymorphone for Study 1						
Treatment 1A		Treatment 1B		Treatment 1C		
Mean	SD	Mean	SD	Mean	SD	
0.8956	0.2983	1.0362	0.3080	2.9622	1.0999	
7.03	4.10	4.89	3.44	0.928	0.398	
17.87	6.140	17.16	6.395	14.24	5.003	
19.87	6.382	18.96	6.908	16.99	5.830	
10.9	2.68	11.4	2.88	6.96	4.61	

Units:
 C_{max} in ng/ml,
 T_{max} in hours,
AUC in ng * hr/ml,
 $T_{1/2rel}$ in hours.

Relative bioavailability determinations are set forth in Tables 7 and 8. For these calculations, AUC was normalized for all treatments to a 20 mg dose.

TABLE 7

Relative Bioavailability (F_{rel}) Determination Based on $AUC_{(0-t)}$		
F_{rel} (1A vs. 1C)	F_{rel} (1B vs. 1C)	F_{rel} (1A vs. 1B)
1.193 ± 0.203	1.121 ± 0.211	1.108 ± 0.152

TABLE 8

Relative Bioavailability Determination Based on $AUC_{(0-t)}$		
F_{rel} (1A vs. 1C)	F_{rel} (1B vs. 1C)	F_{rel} (1A vs. 1B)
0.733 ± 0.098	0.783 ± 0.117	0.944 ± 0.110

Study 2—Two CR Formulations; Fed Patients

Healthy volunteers received a single oral dose of 20 mg CR oxymorphone taken with 240 ml water in a fed state. Subjects received the tablets of Example 2 (Treatment 2A) or Example 3 (Treatment 2B). Further subjects were given a single oral dose of 10 mg/10 ml oxymorphone solution in 180 ml apple juice followed with 60 ml water (Treatment 2C). The orally dosed solution was used to simulate an immediate release (IR) dose.

This study had a single-center, open-label, randomized, three-way crossover design using fifteen subjects. The subjects were in a fed state, after a 10-hour overnight fast fol-

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lowed by a standardized FDA high-fat breakfast. There was a 14-day washout interval between the three dose administrations. The subjects were confined to the clinic during each study period. Subjects receiving Treatment 2C were confined for 18 hours and subjects receiving Treatments 2A or 2B were confined for 48 hours after dosing. Ten-milliliter blood samples were collected during each study period at the 0 hour (predose), and at 0.5, 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 10, 12, 14, 16, 18, 20, 24, 28, 32, 36, and 48 hours postdose for subjects receiving Treatment 2A or 2B and 0, 0.25, 0.5, 0.75, 1, 1.25, 1.5, 1.75, 2, 2.5, 3, 4, 5, 6, 7, 8, 10, 12, 14, 16, and 18 hours postdose. The mean plasma concentration of oxymorphone versus time for each treatment across all subjects is shown in table 9.

TABLE 9

Mean Plasma Concentration vs. Time (ng/ml)			
Time (hr)	Treatment 2A	Treatment 2B	Treatment 2C
0	0.000	0.000	0.0000
0.25			1.263
0.5	0.396	.0553	1.556
0.75			1.972
25	1	0.800	1.063
	1.25		1.795
	1.5	1.038	1.319
	1.75		1.637
	2	1.269	1.414
	2.5		1.454
30	3	1.328	1.540
	4	1.132	1.378
	5	1.291	1.609
	6	1.033	1.242
	7	0.941	0.955
	8	0.936	0.817
35	10	0.669	0.555
	12	0.766	0.592
	14	0.641	0.519
	16	0.547	0.407
	18	0.453	0.320
	20	0.382	0.280
40	24	0.315	0.254
	28	0.352	0.319
	32	0.304	0.237
	36	0.252	0.207
	48	0.104	0.077

The results are shown graphically in FIG. 6. Again, the results have been normalized to a 20 mg dosage. As with Study 1, the immediate release liquid of Treatment 2C shows a classical curve, with a high and relatively narrow peak, followed by an exponential drop in plasma concentration, while the controlled release oxymorphone tablets exhibit triple peaks in blood plasma concentration. Thus, again when we refer to the time to peak plasma concentration (T_{max}) unless otherwise specified, we refer to the time to the second peak.

TABLE 10

Pharmacokinetic Parameters of Plasma Oxymorphone for Study 2						
Treatment 2A		Treatment 2B		Treatment 2C		
	Mean	SD	Mean	SD	Mean	SD
C_{max}	1.644	0.365	1.944	0.465	4.134	0.897
T_{max}	3.07	1.58	2.93	1.64	0.947	0.313
AUC _(0-t)	22.89	5.486	21.34	5.528	21.93	5.044

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TABLE 10-continued

Pharmacokinetic Parameters of Plasma Oxymorphone for Study 2						
	Treatment 2A		Treatment 2B		Treatment 2C	
	Mean	SD	Mean	SD	Mean	SD
AUC _(0-t_{last})	25.28	5.736	23.62	5.202	24.73	6.616
T _{1/2rel}	12.8	3.87	11.0	3.51	5.01	2.02

Units:
 C_{max} in ng/ml,
 T_{max} in hours,
AUC in ng * hr/ml,
 $T_{1/2rel}$ in hours.

In Table 10, the T_{max} has a large standard deviation due to the two comparable peaks in blood plasma concentration. Relative bioavailability determinations are set forth in Tables 11 and 12.

TABLE 11

Relative Bioavailability Determination Based on AUC _(0-t_{last})		
F _{rel} (2A vs. 2C)	F _{rel} (2B vs. 2C)	F _{rel} (2A vs. 2B)
1.052 ± 0.187	0.949 ± 0.154	1.148 ± 0.250

TABLE 12

Relative bioavailability Determination Based on AUC _(0-t_{last})		
F _{rel} (2A vs. 2C)	F _{rel} (2B vs. 2C)	F _{rel} (2A vs. 2B)
0.690 ± 0.105	0.694 ± 0.124	1.012 ± 0.175

As may be seen from tables 5 and 10 and FIGS. 1 and 2, the C_{max} for the CR tablets (treatments 1A, 1B, 2A and 2B) is considerably lower, and the T_{max} much higher than for the immediate release oxymorphone. The blood plasma level of oxymorphone remains high well past the 8 (or even the 12) hour dosing interval desired for an effective controlled release tablet.

Study 3—One Controlled Release Formulation; Fed and Fasted Patients

This study had a single-center, open-label, analytically blinded, randomized, four-way crossover design. Subjects randomized to Treatment 3A and Treatment 3C, as described below, were in a fasted state following a 10-hour overnight fast. Subjects randomized to Treatment 3B and Treatment 3D, as described below, were in the fed state, having had a high fat meal, completed ten minutes prior to dosing. There was a 14-day washout interval between the four dose administrations. The subjects were confined to the clinic during each study period. Subjects assigned to receive Treatment 3A and Treatment 3B were discharged from the clinic on Day 3 following the 48-hour procedures, and subjects assigned to receive Treatment 3C and Treatment 3D were discharged from the clinic on Day 2 following the 36-hour procedures. On Day 1 of each study period the subjects received one of four treatments:

Treatments 3A and 3B: Oxymorphone controlled release 60 mg tablets from Example 3. Subjects randomized to Treatment 3A received a single oral dose of one 20 mg oxymorphone controlled release tablet taken with 240 ml of water after a 10-hour fasting period. Subjects randomized to Treatment 3B received a single oral dose of one 20 mg oxymorphone controlled release tablet taken with 240 ml of water 10 minutes after a standardized high fat meal.

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Treatments 3C and 3D: oxymorphone HCl solution, USP, 1.5 mg/ml 10 ml vials. Subjects randomized to Treatment 3C received a single oral dose of 10 mg (6.7 ml) oxymorphone solution taken with 240 ml of water after a 10-hour fasting period. Subjects randomized to Treatment 3D received a single oral dose of 10 mg (6.7 ml) oxymorphone solution taken with 240 ml of water 10 minutes after a standardized high-fat meal.

10 A total of 28 male subjects were enrolled in the study, and 24 subjects completed the study. The mean age of the subjects was 27 years (range of 19 through 38 years), the mean height of the subjects was 69.6 inches (range of 64.0 through 75.0 inches), and the mean weight of the subjects was 169.0 pounds (range 117.0 through 202.0 pounds).

15 A total of 28 subjects received at least one treatment. Only subjects who completed all 4 treatments were included in the summary statistics and statistical analysis.

20 Blood samples (7 ml) were collected during each study period at the 0 hour (predose), and at 0.5, 1, 1.5, 2, 3, 4, 5, 6, 8, 10, 12, 14, 16, 20, 24, 30, 36, and 48 hours post-dose (19 samples) for subjects randomized to Treatment 3A and Treatment 3B. Blood samples (7 ml) were collected during each 25 study period at the 0 hour (predose), and at 0.25, 0.5, 0.75, 1, 1.25, 1.5, 1.75, 2, 3, 4, 5, 6, 8, 10, 12, 14, 16, 20, and 36 hours post-dose (21 samples) for subjects randomized to Treatment 3C and Treatment 3D.

30 The mean oxymorphone plasma concentration versus time curves for Treatments 3A, 3B, 3C, and 3D are presented in FIG. 7. The results have been normalized to a 20 mg dosage. The data is contained in Table 13. The arithmetic means of the plasma oxymorphone pharmacokinetic parameters and the statistics for all Treatments are summarized in Table 14.

TABLE 13

Time (hr)	Mean Plasma Concentration vs. Time (ng/ml)			
	Treatment 3A	Treatment 3B	Treatment 3C	Treatment 3D
0	0.0084	0.0309	0.0558	0.0000
0.25			0.5074	0.9905
0.5	0.3853	0.3380	0.9634	1.0392
0.75			0.9753	1.3089
1	0.7710	0.7428	0.8777	1.3150
1.25			0.8171	1.2274
1.5	0.7931	1.0558	0.7109	1.1638
1.75			0.6357	1.0428
2	0.7370	1.0591	0.5851	0.9424
3	0.6879	0.9858	0.4991	0.7924
4	0.6491	0.9171	0.3830	0.7277
5	0.9312	1.4633	0.3111	0.6512
6	0.7613	1.0441	0.2650	0.4625
8	0.5259	0.7228	0.2038	0.2895
10	0.4161	0.5934	0.1768	0.2470
12	0.5212	0.5320	0.2275	0.2660
14	0.4527	0.4562	0.2081	0.2093
16	0.3924	0.3712	0.1747	0.1623
20	0.2736	0.3021	0.1246	0.1144
24	0.2966	0.2636	0.1022	0.1065
30	0.3460	0.3231		
36	0.2728	0.2456	0.0841	0.0743
48	0.1263	0.1241		

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TABLE 14

Pharmacokinetic Parameters of Plasma Oxymorphone for Study 3								
	Treatment 3B		Treatment 3A		Treatment 3C		Treatment 3D	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
C_{max}	1.7895	0.6531	1.1410	0.4537	2.2635	1.0008	3.2733	1.3169
T_{max}	5.56	9.39	5.57	7.14	0.978	1.14	1.11	0.768
$AUC_{(0-24)}$	14.27	4.976	11.64	3.869	12.39	4.116	17.30	5.259
$AUC_{(0-t)}$	19.89	6.408	17.71	8.471	14.53	4.909	19.20	6.030
$AUC_{(0-Inf)}$	21.29	6.559	19.29	5.028	18.70	6.618	25.86	10.03
$T_{1/2rel}$	12.0	3.64	12.3	3.99	16.2	11.4	20.6	19.3

The relative bioavailability calculations are summarized in tables 15 and 16.

TABLE 15

Relative Bioavailability Determination Based on $AUC_{(0-Inf)}$			
F_{rel} (3A vs. 3C)	F_{rel} (3B vs. 3D)	F_{rel} (3D vs. 3C)	F_{rel} (3B vs. 3A)
1.040 ± 0.1874	0.8863 ± 0.2569	1.368 ± 0.4328	1.169 ± 0.2041

TABLE 16

Relative Bioavailability Determination Based on $AUC_{(0-24)}$			
F_{rel} (3A vs. 3C)	F_{rel} (3B vs. 3D)	F_{rel} (3D vs. 3C)	F_{rel} (3B vs. 3A)
0.9598 ± 0.2151	0.8344 ± 0.100	1.470 ± 0.3922	1.299 ± 0.4638

The objectives of this study were to assess the relative bioavailability of oxymorphone from oxymorphone controlled release (20 mg) compared to oxymorphone oral solution (10 mg) under both fasted and fed conditions, and to determine the effect of food on the bioavailability of oxymorphone from the controlled release formulation, oxymorphone CR, and from the oral solution.

The presence of a high fat meal had a substantial effect on the oxymorphone C_{max} but less of an effect on oxymorphone AUC from oxymorphone controlled release tablets. Least Squares (LS) mean C_{max} was 58% higher and LS mean $AUC_{(0-t)}$ and $AUC_{(0-Inf)}$ were 18% higher for the fed condition (Treatment B) compared to the fasted condition (Treatment A) based on LN-transformed data. This was consistent with the relative bioavailability determination from $AUC_{(0-Inf)}$ since mean F_{rel} was 1.17. Mean T_{max} values were similar (approximately 5.6 hours), and no significant difference in T_{max} was shown using nonparametric analysis. Half value durations were significantly different between the two treatments.

The effect of food on oxymorphone bioavailability from the oral solution was more pronounced, particularly in terms of AUC. LS mean C_{max} was 50% higher and LS mean $AUC_{(0-t)}$ and $AUC_{(0-Inf)}$ were 32-34% higher for the fed condition (Treatment D) compared to the fasted condition (Treatment C) based on LN-transformed data. This was consistent with the relative bioavailability determination from $AUC_{(0-Inf)}$ since mean F_{rel} was 1.37. Mean T_{max} (approximately 1 hour) was similar for the two treatments and no significant difference was shown.

Under fasted conditions, oxymorphone controlled release 20 mg tablets exhibited similar extent of oxymorphone availability compared to 10 mg oxymorphone oral solution normalized to a 20 mg dose (Treatment A versus Treatment C).

15 From LN-transformed data, LS mean $AUC_{(0-t)}$ was 17% higher for oxymorphone CR, whereas LS mean $AUC_{(0-Inf)}$ values were nearly equal (mean ratio=99%). Mean F_{rel} values calculated from $AUC_{(0-Inf)}$ and $AUC_{(0-24)}$, (1.0 and 0.96, respectively) also showed similar extent of oxymorphone availability between the two treatments.

20 As expected, there were differences in parameters reflecting rate of absorption. LS mean C_{max} was 49% lower for oxymorphone controlled release tablets compared to the dose-normalized oral solution, based on LN-transformed data. Half-value duration was significantly longer for the controlled release formulation (means, 12 hours versus 2.5 hours).

25 Under fed conditions, oxymorphone availability from oxymorphone controlled release 20 mg was similar compared to 10 mg oxymorphone oral solution normalized to a 20 mg dose (Treatment B versus Treatment D). From LN-transformed data, LS mean $AUC_{(0-Inf)}$ was 12% lower for oxymorphone CR. Mean F_{rel} values calculated from $AUC_{(0-Inf)}$ and $AUC_{(0-24)}$, (0.89 and 0.83 respectively) also showed similar extent of oxymorphone availability from the tablet. As expected, there were differences in parameters reflecting rate of absorption. LS mean C_{max} was 46% lower for oxymorphone controlled release tablets compared to the dose-normalized oral solution, based on LN-transformed data. Mean T_{max} was 5.7 hours for the tablet compared to 1.1 hours for the oral solution. Half-value duration was significantly longer for the controlled release formulation (means, 7.8 hours versus 3.1 hours).

30 The presence of a high fat meal did not appear to substantially affect the availability of 6-hydroxyoxymorphone following administration of oxymorphone controlled release tablets. LS mean ratios were 97% for $AUC_{(0-t)}$ and 91% for C_{max} (Treatment B versus A), based on LN-transformed data. This was consistent with the relative bioavailability determination from $AUC_{(0-24)}$, since mean F_{rel} was 0.97. Mean T_{max} was later for the fed treatment compared to the fasted treatment (5.2 and 3.6 hours, respectively), and difference was significant.

35 Under the fasted conditions, oxymorphone controlled release 20 mg tablets exhibited similar availability of 6-hydroxyoxymorphone compared to 10 mg oxymorphone oral solution normalized to a 20 mg dose (Treatment A versus Treatment C). From LN-transformed data, LS mean ratio for $AUC_{(0-t)}$ was 104.5%. Mean F_{rel} (0.83) calculated from $AUC_{(0-24)}$ also showed similar extent of oxymorphone availability between the two treatments. Mean T_{max} was 3.6 hours for the tablet compared to 0.88 for the oral solution. Half-value duration was significantly longer for the controlled release formulation (means, 11 hours versus 2.2 hours).

40 Under fed conditions, availability of 6-hydroxyoxymorphone from oxymorphone controlled release 20 mg was similar compared to 10 mg oxymorphone oral solution normal-

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ized to a 20 mg dose (Treatment B versus Treatment D). From LN-transformed data, LS mean $AUC_{(0-t)}$ was 14% higher for oxymorphone CR. Mean F_{rel} (0.87) calculated from $AUC_{(0-24)}$ also indicated similar extent of availability between the treatments. Mean T_{max} was 5.2 hours for the tablet compared to 1.3 hour for the oral solution. Half-value duration was significantly longer for the controlled release formulation (means, 14 hours versus 3.9 hours).

The extent of oxymorphone availability from oxymorphone controlled release 20 mg tablets was similar under fed and fasted conditions since there was less than a 20% difference in LS mean $AUC_{(0-t)}$ and $AUC_{(0-\infty)}$ values for each treatment, based on LN-transformed data. T_{max} was unaffected by food; however, LS mean C_{max} was increased 58% in the presence of the high fat meal. Both rate and extent of oxymorphone absorption from the oxymorphone oral solution were affected by food since LS mean C_{max} and AUC values were increased approximately 50 and 30%, respectively. T_{max} was unaffected by food. Under both fed and fasted conditions, oxymorphone controlled release tablets exhibited similar extent of oxymorphone availability compared to oxymorphone oral solution since there was less than a 20% difference in LS mean $AUC_{(0-t)}$ and $AUC_{(0-\infty)}$ values for each treatment.

Bioavailability of 6-hydroxyoxymorphone following oxymorphone controlled release 20 mg tablets was also similar under fed and fasted conditions since there was less than a 20% difference in LS mean C_{max} and AUC values for each treatment. T_{max} was later for the fed condition. The presence of food did not affect the extent

20

TABLE 17-continued

Time (hr)	Mean Plasma Concentration vs. Time (ng/ml) 6-Hydroxyoxymorphone			
	Treatment 3A Treatment 3B Treatment 3C Treatment 3D			
	3A	3B	3C	3D
1	1.0233	0.4830	1.1072	0.8080
1.25			1.0069	0.7266
1.5	1.1062	0.7456	0.8494	0.7001
1.75			0.7511	0.6472
2	1.0351	0.7898	0.6554	0.5758
3	0.9143	0.7619	0.6196	0.5319
4	0.8522	0.7607	0.4822	0.5013
5	0.8848	0.8548	0.3875	0.4448
6	0.7101	0.7006	0.3160	0.3451
8	0.5421	0.5681	0.2525	0.2616
10	0.4770	0.5262	0.2361	0.2600
12	0.4509	0.4454	0.2329	0.2431
14	0.4190	0.4399	0.2411	0.2113
16	0.4321	0.4230	0.2385	0.2086
20	0.3956	0.4240	0.2234	0.1984
24	0.4526	0.4482	0.2210	0.2135
30	0.4499	0.4708		
36	0.3587	0.3697	0.1834	0.1672
48	0.3023	0.3279		

TABLE 18

Pharmacokinetic Parameters of Plasma 6-Hydroxyoxymorphone for Study 3									
	Treatment 3A		Treatment 3B		Treatment 3C		Treatment 3D		SD
	Mean	SD	Mean	SD	Mean	SD	Mean	SD	
C_{max}	1.2687	0.5792	1.1559	0.4848	1.5139	0.7616	0.9748	0.5160	
T_{max}	3.61	7.17	5.20	9.52	0.880	0.738	1.30	1.04	
$AUC_{(0-t)}$	22.47	10.16	22.01	10.77	10.52	4.117	9.550	4.281	
$AUC_{(0-\infty)}$	38.39	23.02	42.37	31.57	20.50	7.988	23.84	11.37	
$T_{1/2el}$	39.1	36.9	39.8	32.6	29.3	12.0	44.0	35.00	

of availability from oxymorphone oral solution since LS mean AUC values were less than 20% different. However, C_{max} was decreased 35% in the presence of food. T_{max} was unaffected by food. Under both fed and fasted conditions, oxymorphone controlled release tablets exhibited similar extent availability compared to oxymorphone oral solution since there was less than a 20% difference in LS mean AUC values for each treatment.

The mean 6-OH oxymorphone plasma concentration versus time curves for Treatments 3A, 3B, 3C, and 3D are presented in FIG. 8. The data is contained in Table 17.

45 Study 4—Controlled Release 20 mg vs. Immediate Release 10 mg

A study was conducted to compare the bioavailability and pharmacokinetics of controlled release and immediate release oxymorphone tablets under single-dose and multiple-dose (steady state) conditions. For the controlled release study, healthy volunteers received a single dose of a 20 mg controlled release oxymorphone tablet on the morning of Day 1. Beginning on the morning of Day 3, the volunteers were administered a 20 mg controlled release oxymorphone tablet every 12 hours through the morning dose of Day 9. For the immediate release study, healthy volunteers received a single 10 mg dose of an immediate release oxymorphone tablet on the morning of Day 1. On the morning of Day 3, additional 10 mg immediate release tablets were administered every six hours through the first two doses on Day 9.

60 FIG. 9 shows the average plasma concentrations of oxymorphone and 6-hydroxyoxymorphone for all subjects after a single dose either controlled release (CR) 20 mg or immediate release (IR) 10 mg oxymorphone. The data in the figure (as with the other relative experimental data herein) is normalized to a 20 mg dose. The immediate release tablet shows a classical curve, with a high, relatively narrow peak followed

TABLE 17

Time (hr)	Mean Plasma Concentration vs. Time (ng/ml) 6-Hydroxyoxymorphone			
	Treatment 3A	Treatment 3B	Treatment 3C	Treatment 3D
0	0.0069	0.0125	0.0741	0.0000
0.25			0.7258	0.4918
0.5	0.5080	0.1879	1.2933	0.5972
0.75			1.3217	0.7877

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by an exponential drop in plasma concentration. The controlled release oxymorphone tablets show a lower peak with extended moderate levels of oxymorphone and 6-hydroxy oxymorphone. Table 19 shows the levels of oxymorphone and 6-hydroxy oxymorphone from FIG. 9 in tabular form.

TABLE 19

Mean Plasma Concentration (ng/ml)							
Oxymorphone				6-Hydroxyoxymorphone			
Hour	Controlled Release 20 mg	Immediate Release 10 mg	Controlled Release 20 mg	Immediate Release 10 mg	Controlled Release 20 mg	Immediate Release 10 mg	Controlled Release 20 mg
0.00	0.00	0.00	0.00	0.00			
0.25	0.22	1.08	0.14	0.73			
0.50	0.59	1.69	0.45	1.22			
1.00	0.77	1.19	0.53	0.79			
1.50	0.84	0.91	0.53	0.57			
2.00	0.87	0.75	0.60	0.47			
3.00	0.83	0.52	0.55	0.34			
4.00	0.73	0.37	0.53	0.27			
5.00	0.94	0.36	0.46	0.23			
6.00	0.81	0.28	0.41	0.18			
8.00	0.73	0.20	0.37	0.14			
10.0	0.60	0.19	0.35	0.15			
12.0	0.67	0.25	0.32	0.13			
16.0	0.39	0.16	0.29	0.13			
24.0	0.23	0.07	0.29	0.13			
30.0	0.12	0.01	0.17	0.04			
36.0	0.05	0.00	0.11	0.00			
48.0	0.00	0.00	0.07	0.01			

FIG. 10 shows the average plasma concentrations of oxymorphone and 6-hydroxyoxymorphone for all subjects in the steady state test, for doses of controlled release 20 mg tablets and immediate release 10 mg tablets of oxymorphone. The figure shows the plasma concentrations after the final controlled release tablet is given on Day 9, and the final immediate release tablet is given 12 hours thereafter. The steady state administration of the controlled release tablets clearly shows a steady moderate level of oxymorphone ranging from just over 1 ng/ml to almost 1.75 ng/ml over the course of a twelve hour period, where the immediate release tablet shows wide variations in blood plasma concentration. Table 20 shows the levels of oxymorphone and 6-hydroxyoxymorphone from FIG. 10 in tabular form.

TABLE 20

Summary of Mean Plasma Concentration (ng/ml)							
Oxymorphone				6-Hydroxyoxymorphone			
Day	Hour	Controlled Release 20 mg	Immediate Release 10 mg	Controlled Release 20 mg	Immediate Release 10 mg	Controlled Release 20 mg	Immediate Release 10 mg
4	0.00	1.10	0.75	0.89	0.72		
5	0.00	1.12	0.84	1.15	0.88		
6	0.00	1.20	0.92	1.15	0.87		
7	0.00	1.19	0.91	1.27	1.00		
8	0.00	1.19	0.86	1.29	0.98		
9	0.00	1.03	1.07	1.09	1.05		
	0.25		2.64		1.70		
	0.50		3.12		2.09		
	1.00		2.47		1.70		
	1.50		2.05		1.63		
	2.00		1.78		1.64		
	3.00		1.27		1.47		
	4.00		0.98		1.39		
	5.00		1.01		1.21		
	6.00		0.90		1.06		
	6.25		1.17		0.88		

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TABLE 20-continued

5	Summary of Mean Plasma Concentration (ng/ml)							
	Oxymorphone				6-Hydroxyoxymorphone			
	Day	Hour	Controlled Release 20 mg	Immediate Release 10 mg	Controlled Release 20 mg	Immediate Release 10 mg	Controlled Release 20 mg	Immediate Release 10 mg
10		6.50			1.88			1.06
		7.00			2.12			1.20
		7.50			2.24			1.15
		8.00	1.32		2.01		0.97	1.03
		9.00			1.52			0.90
		10.00	1.32		1.24		0.85	0.84
		11.00			1.11			0.74
		12.00	1.18		0.96		0.79	0.70
15								

20	Mean Single-Dose Pharmacokinetic Results							
	Controlled Release 20 mg				Immediate Release 10 mg			
	oxy-morphone	6-OH-oxymorphone	oxymor-phone	6-OH-oxymorphone				
25	AUC _(o-t)	14.74	11.54	7.10	5.66			
	AUC _(o-inf)	15.33	16.40	7.73	8.45			
	C _{max} (ng/ml)	1.12	0.68	1.98	1.40			
	T _{max} (hr)	5.00	2.00	0.50	0.50			
	T _{1/2} (hr)	9.25	26.09	10.29	29.48			
30								

Parent 6-OH oxymorphone AUC_(o-t) values were lower than the parent compound after administration of either dosage form, but the AUC_(o-inf) values are slightly higher due to the longer half-life for the metabolite. This relationship was similar for both the immediate-release (IR) and controlled release (CR) dosage forms. As represented by the average plasma concentration graph, the CR dosage form has a significantly longer time to peak oxymorphone concentration and a lower peak oxymorphone concentration. The 6-OH oxymorphone peak occurred sooner than the parent peak following the CR dosage form, and simultaneously with the parent peak following the IR dosage form.

It is important to note that while the present invention is described and exemplified, using 20 mg tablets, the invention may also be used with other strengths of tablets. In each strength, it is important to note how a 20 mg tablet of the same composition (except for the change in strength) would act. The blood plasma levels and pain intensity information are provided for 20 mg tablets, however the present invention is also intended to encompass 5 to 80 mg controlled release tablets. For this reason, the blood plasma level of oxymorphone or 6-hydroxyoxymorphone in nanograms per milliliter of blood, per mg oxymorphone (ng/mg·ml) administered is measured. Thus at 0.02 ng/mg·ml, a 5 mg tablet should produce a minimum blood plasma concentration of 0.1 ng/ml. A stronger tablet will produce a higher blood plasma concentration of active molecule, generally proportionally. Upon administration of a higher dose tablet, for example 80 mg, the blood plasma level of oxymorphone and 6-OH oxymorphone may more than quadruple compared to a 20 mg dose, although conventional treatment of low bioavailability substances would lead away from this conclusion. If this is the case, it may be because the body can only process a limited amount oxymorphone at one time. Once the bolus is processed, the blood level of oxymorphone returns to a proportional level.

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It is the knowledge that controlled release oxymorphone tablets are possible to produce and effective to use, which is most important, made possible with the high bioavailability of oxymorphone in a controlled release tablet. This also holds true for continuous periodic administration of controlled release formulations. The intent of a controlled release opioid formulation is the long-term management of pain. Therefore, the performance of a composition when administered periodically (one to three times per day) over several days is important. In such a regime, the patient reaches a "steady state" where continued administration will produce the same results, when measured by duration of pain relief and blood plasma levels of pharmaceutical. Such a test is referred to as a "steady state" test and may require periodic administration over an extended time period ranging from several days to a week or more. Of course, since a patient reaches steady state in such a test, continuing the test for a longer time period should not affect the results. Further, when testing blood plasma levels in such a test, if the time period for testing exceeds the interval between doses, it is important the regimen be stopped after the test is begun so that observations of change in blood level and pain relief may be made without a further dose affecting these parameters.

Study 5—Controlled Release 40 mg vs. Immediate Release 4.times.10 mg under Fed and Fasting Conditions

The objectives of this study were to assess the relative bioavailability of oxymorphone from oxymorphone controlled release (40 mg) compared to oxymorphone immediate release (4.times.10 mg) under both fasted and fed conditions, and to determine the effect of food on the bioavailability of oxymorphone from the controlled release formulation, oxymorphone CR, and from the immediate release formulation, oxymorphone IR.

This study had a single-center, open-label, analytically blinded, randomized, four-way crossover design. Subjects randomized to Treatment 5A and Treatment 5C, as described below, were in a fasted state following a 10-hour overnight fast. Subjects randomized to Treatment 5B and Treatment 5D, as described below, were in the fed state, having had a high fat meal, completed ten minutes prior to dosing. There was a 14-day washout interval between the four dose administrations. The subjects were confined to the clinic during each study period. Subject assigned to receive Treatment 5A and Treatment 5B were discharged from the clinic on Day 3 following the 48-hour procedures, and subjects assigned to receive Treatment 5C and Treatment 5D were discharged from the clinic on Day 2 following the 36-hour procedures. On Day 1 of each study period the subjects received one of four treatments:

Treatments 5A and 5B: Oxymorphone controlled release 40 mg tablets from Table 2. Subjects randomized to Treatment 5A received a single oral dose of one 40 mg oxymorphone controlled release tablet taken with 240 ml of water after a 10-hour fasting period. Subjects randomized to Treatment 5B received a single oral dose of one 40 mg oxymorphone controlled release tablet taken with 240 ml of water 10 minutes after a standardized high fat meal.

Treatments 5C and 5D: Immediate release tablet (IR) 4.times.10 mg Oxymorphone. Subjects randomized to Treatment 5C received a single oral dose of 4.times.10 mg oxymorphone IR tablet taken with 240 ml of water after a 10-hour fasting period. Subjects randomized to Treatment 5D received a single oral dose of 4.times.10 mg oxymorphone IR tablet taken with 240 ml of water 10 minutes after a standardized high-fat meal.

A total of 28 male subjects were enrolled in the study, and 25 subjects completed the study. A total of 28 subjects

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received at least one treatment. Only subjects who completed all 4 treatments were included in the summary statistics and statistical analysis.

Blood samples (7 ml) were collected during each study period at the 0 hour (predose), and at 0.25, 0.5, 0.75, 1.0, 1.5, 2, 3, 4, 5, 6, 8, 10, 12, 24, 36, 48, 60, and 72 hours post-dose (19 samples) for subjects randomized to all Treatments.

The mean oxymorphone plasma concentration versus time is presented in Table 22. The arithmetic means of the plasma oxymorphone pharmacokinetic parameters and the statistics for all Treatments are summarized in Table 23.

TABLE 22

Time (hr)	Mean Plasma Concentration vs. Time (ng/ml)			
	Treatment 5A	Treatment 5B	Treatment 5C	Treatment 5D
0	0.00	0.00	0.00	0.00
0.25	0.47	0.22	3.34	1.79
0.50	1.68	0.97	7.28	6.59
0.75	1.92	1.90	6.60	9.49
1	2.09	2.61	6.03	9.91
1.5	2.18	3.48	4.67	8.76
2	2.18	3.65	3.68	7.29
3	2.00	2.86	2.34	4.93
4	1.78	2.45	1.65	3.11
5	1.86	2.37	1.48	2.19
6	1.67	2.02	1.28	1.71
8	1.25	1.46	0.92	1.28
10	1.11	1.17	0.78	1.09
12	1.34	1.21	1.04	1.24
24	0.55	0.47	0.40	0.44
36	0.21	0.20	0.16	0.18
48	0.06	0.05	0.04	0.05
60	0.03	0.01	0.01	0.01
72	0.00	0.00	0.00	0.00

TABLE 23

	Pharmacokinetic Parameters of Plasma Oxymorphone for Study 5							
	Treatment 5A		Treatment 5B		Treatment 5C		Treatment 5D	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
C _{max}	2.79	0.84	4.25	1.21	9.07	4.09	12.09	5.42
T _{max}	2.26	2.52	1.96	1.06	0.69	0.43	1.19	0.62
AUC _(0-∞)	35.70	10.58	38.20	11.04	36.00	12.52	51.35	20.20
AUC ₍₀₋₂₄₎	40.62	11.38	41.17	10.46	39.04	12.44	54.10	20.26
T _{1/2el}	12.17	7.57	10.46	5.45	11.65	6.18	9.58	3.63

The relative bioavailability calculations are summarized in Tables 24 and 25.

TABLE 24

Relative Bioavailability Determination Based on AUC _(0-∞)	
F _{rel} (5D vs. 5C)	F _{rel} (5B vs. 5A)
1.3775	1.0220

TABLE 25

Relative bioavailability Determination Based on AUC ₍₀₋₂₄₎	
F _{rel} (5D vs. 5C)	F _{rel} (5B vs. 5A)
1.4681	1.0989

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The mean 6-OH oxymorphone plasma concentration versus time is presented in Table 26.

TABLE 26

Time (hr)	Mean Plasma Concentration vs. Time (ng/ml) 6-Hydroxyoxymorphone			
	Treatment 5A	Treatment 5B	Treatment 5C	Treatment 5D
0	0.00	0.00	0.00	0.00
0.25	0.27	0.05	2.36	0.50
0.50	1.32	0.31	5.35	1.98
0.75	1.37	0.59	4.53	2.97
1	1.44	0.82	3.81	2.87
1.5	1.46	1.09	2.93	2.58
2	1.46	1.28	2.37	2.29
3	1.39	1.14	1.69	1.72
4	1.25	1.14	1.33	1.26
5	1.02	1.00	1.14	1.01
6	0.93	0.86	0.94	0.86
8	0.69	0.72	0.73	0.77
10	0.68	0.67	0.66	0.75
12	0.74	0.66	0.70	0.77
24	0.55	0.52	0.54	0.61
36	0.23	0.30	0.28	0.27
48	0.18	0.20	0.20	0.19
60	0.09	0.10	0.09	0.09
72	0.06	0.06	0.04	0.05

TABLE 27

Pharmacokinetic Parameters of Plasma 6-Hydroxyoxymorphone for Study 5								
Treatment 5A		Treatment 5B		Treatment 5C		Treatment 5D		
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
C _{max}	1.88	0.69	1.59	0.63	6.41	3.61	3.79	1.49
T _{max}	1.48	1.18	2.73	1.27	0.73	0.47	1.18	0.74
AUC _(0->t)	28.22	10.81	26.95	11.39	33.75	10.29	32.63	13.32
AUC _(0->inf)	33.15	11.25	32.98	10.68	37.63	17.01	36.54	13.79
T _{1/2el}	17.08	7.45	21.92	8.41	16.01	6.68	16.21	7.42

The above description incorporates preferred embodiments and examples as a means of describing and enabling the invention to be practiced by one of skill in the art. It is imagined that changes can be made without departing from the spirit and scope of the invention described herein and defined in the appended claims.

We claim:

1. An analgesically effective controlled release pharmaceutical composition with a twelve hour dosing interval in the form of a tablet, comprising oxymorphone or a pharmaceutically acceptable salt thereof as the sole active ingredient in the tablet, and a controlled release delivery system comprising at least one pharmaceutical excipient, wherein upon placement of the composition in an in vitro dissolution test comprising USP Paddle Method at 50 rpm in 500 ml media having a pH of 1.2 to 6.8 at 37° C., about 15% to about 50%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 1 hour in the test.

2. The pharmaceutical composition of claim 1 wherein about 45% to about 80%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 4 hours in the test.

3. The pharmaceutical composition of claim 1 wherein at least about 80%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 10 hours in the test.

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4. The pharmaceutical composition of claim 1 wherein the controlled release delivery system comprises a hydrophilic material that forms a gel upon exposure to gastrointestinal fluid.

5. The pharmaceutical composition of claim 1 wherein the controlled release delivery system comprises a heteropolysaccharide and an agent capable of cross-linking the heteropolysaccharide in presence of gastrointestinal fluid.

6. The pharmaceutical composition of claim 5 wherein the heteropolysaccharide and the agent capable of cross-linking the heteropolysaccharide are present in a weight ratio of about 1:3 to about 3:1.

7. The pharmaceutical composition of claim 5 wherein the heteropolysaccharide comprises xanthan gum or deacylated xanthan gum.

8. The pharmaceutical composition of claim 5 wherein the agent capable of cross-linking the heteropolysaccharide comprises a homopolysaccharide gum.

9. The pharmaceutical composition of claim 8 wherein the homopolysaccharide gum comprises locust bean gum.

10. The pharmaceutical composition of claim 1 wherein the controlled release delivery system further comprises a hydrophobic polymer.

11. The pharmaceutical composition of claim 10 wherein the hydrophobic polymer comprises an alkylcellulose.

12. The pharmaceutical composition of claim 8 further comprising a cationic cross-linking agent.

13. The pharmaceutical composition of claim 12 wherein the cationic cross-linking agent is selected from calcium sulfate, sodium chloride, potassium sulfate, sodium carbonate, lithium chloride, tripotassium phosphate, sodium borate, potassium bromide, potassium fluoride, sodium bicarbonate, calcium chloride, magnesium chloride, sodium citrate, sodium acetate, calcium lactate, magnesium sulfate, sodium fluoride, and combinations thereof.

14. The pharmaceutical composition of claim 13 wherein the cationic cross-linking agent is present in an amount of about 0.5% to about 16%, by weight of the composition.

15. The pharmaceutical composition of claim 5 wherein the weight ratio of heteropolysaccharide to oxymorphone or pharmaceutically acceptable salt thereof is about 10:1 to about 1:10.

16. The pharmaceutical composition of claim 1 wherein oxymorphone or pharmaceutically acceptable salt thereof is present in an amount of about 5 mg to about 80 mg.

17. The pharmaceutical composition of claim 5 wherein the controlled release delivery system comprises about 10% to about 99% of a gelling agent comprising a heteropolysaccharide gum and a homopolysaccharide gum, about 1% to about 20% of a cationic crosslinking agent, and about 0% to about 89% of other ingredients which qualify as an inert pharmaceutical diluent, by total weight of the controlled release delivery system.

18. A method of treating pain in a subject in need thereof, the method comprising administering to the subject the pharmaceutical composition of claim 1 comprising about 5 mg to about 80 mg of oxymorphone or pharmaceutically acceptable salt thereof.

19. An analgesically effective controlled release pharmaceutical composition with a twelve hour dosing interval in the form of a tablet, comprising oxymorphone or pharmaceutically acceptable salt thereof as the sole active ingredient in the tablet and a controlled release delivery system comprising a hydrophilic material that forms a gel upon exposure to gastrointestinal fluid, wherein upon placement of the composition in an in vitro dissolution test comprising USP Paddle Method at 50 rpm in 500 ml media having a pH of 1.2 to 6.8

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at 37° C., about 15% to about 50%, by weight, of the oxymorphone or salt thereof is released from the composition at about 1 hour in the test, about 45% to about 80%, by weight, of the oxymorphone or salt thereof is released from the composition at about 4 hours in the test, and at least about 80%, by weight, of the oxymorphone or salt thereof is released from the composition at about 10 hours in the test.

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20. The method of claim 18 wherein upon oral administration of the composition the oxymorphone $AUC_{(0-t)}$ is no more than 20% higher when the composition is administered to the subject under fed as compared to fasted conditions.

* * * * *

Exhibit C



US008329216B2

(12) United States Patent
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(45) Date of Patent: *Dec. 11, 2012

(54) OXYMORPHONE CONTROLLED RELEASE FORMULATIONS

(75) Inventors: **Hau-Hung Kao**, Syosset, NY (US); **Anand R. Baichwal**, Wappingers Falls, NY (US); **Troy McCall**, Smyrna, GA (US); **David Lee**, Chadds, PA (US)(73) Assignee: **Endo Pharmaceuticals Inc.**, Chadds Ford, PA (US)

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This patent is subject to a terminal disclaimer.

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(58) Field of Classification Search None
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(56) References Cited

U.S. PATENT DOCUMENTS

2,806,033 A	9/1957	Lewenstein et al.
3,393,197 A	7/1968	Pachter et al.
3,845,770 A	11/1974	Theeuwes et al.
3,879,555 A	4/1975	Pachter et al.
3,966,940 A	6/1976	Pachter et al.
3,980,766 A	9/1976	Shaw et al.
4,070,494 A	1/1978	Hoffmeister et al.
4,366,159 A	12/1982	Magruder
4,457,933 A	7/1984	Gordon et al.
4,464,376 A	8/1984	Sunshine et al.
4,479,956 A	10/1984	Sunshine et al.
4,486,436 A	12/1984	Sunshine et al.
4,558,051 A	12/1985	Sunshine et al.
4,567,183 A	1/1986	Sunshine et al.
4,569,937 A	2/1986	Baker et al.

(Continued)

FOREIGN PATENT DOCUMENTS

CA 2314896 A1 7/1999

(Continued)

OTHER PUBLICATIONS

U.S. Appl. No. 12/426,122, Kao et al.

(Continued)

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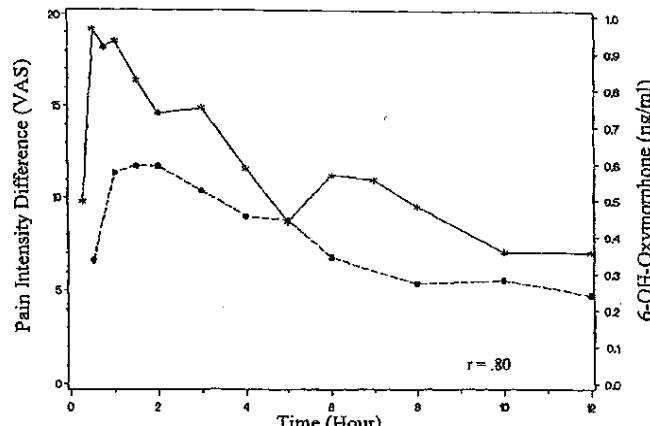
(74) Attorney, Agent, or Firm — Mayer Brown LLP

(57) ABSTRACT

The invention pertains to a method of relieving pain by administering a controlled release pharmaceutical tablet containing oxymorphone which produces a mean minimum blood plasma level 12 to 24 hours after dosing, as well as the tablet producing the sustained pain relief.

82 Claims, 10 Drawing Sheets

PK Profile for 6-OH-Oxymorphone with PID Scores



* Pain Intensity Difference • 6-OH-Oxymorphone Plasma Concentrations

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U.S. PATENT DOCUMENTS

4,582,835 A	4/1986	Lewis et al.	6,475,494 B2	11/2002	Kaiko et al.
4,587,249 A	5/1986	Sunshine et al.	6,495,155 B1	12/2002	Tice et al.
4,599,114 A	7/1986	Atkinson	6,506,730 B1	1/2003	Lee et al.
4,656,177 A	4/1987	Sunshine et al.	6,514,531 B1	2/2003	Alaux et al.
4,661,492 A	4/1987	Lewis et al.	6,555,127 B2	4/2003	Steiner
4,711,782 A	12/1987	Okada et al.	6,627,635 B2	9/2003	Palermo et al.
4,777,174 A	10/1988	Sunshine et al.	6,696,088 B2	2/2004	Oshlack et al.
4,844,907 A	7/1989	Elger et al.	6,716,449 B2	4/2004	Oshlack et al.
4,844,909 A	7/1989	Goldie et al.	6,806,294 B2	10/2004	Wimmer et al.
4,861,598 A	8/1989	Oshlack	7,276,250 B2	10/2007	Baichwal et al.
4,935,428 A	6/1990	Lewis et al.	2001/0008639 A1	7/2001	Oshlack et al.
4,980,170 A	12/1990	Schneider et al.	2002/0010127 A1	1/2002	Oshlack et al.
4,994,276 A	2/1991	Baichwal et al.	2002/0032581 A1	3/2002	Reitberg
5,047,248 A	9/1991	Calanchi et al.	2002/0044966 A1	4/2002	Bartholomaeus et al.
5,128,143 A	7/1992	Baichwal et al.	2002/0058673 A1	5/2002	Kaiko et al.
5,135,757 A	8/1992	Baichwal et al.	2002/0081333 A1	6/2002	Oshlack et al.
5,164,193 A	11/1992	Okada et al.	2002/0090345 A1	7/2002	Baichwal et al.
5,202,128 A	4/1993	Morella et al.	2002/0164373 A1	11/2002	Maione
5,236,714 A	8/1993	Lee et al.	2002/0187192 A1	12/2002	Joshi et al.
5,266,331 A	11/1993	Oshlack et al.	2003/0004177 A1	1/2003	Kao et al.
5,330,761 A	7/1994	Baichwal	2003/0031712 A1	2/2003	Kaiko et al.
5,399,359 A	3/1995	Baichwal et al.	2003/0044458 A1	3/2003	Wright, IV et al.
5,399,362 A	3/1995	Baichwal et al.	2003/0049272 A1	3/2003	Joshi et al.
5,415,871 A	5/1995	Pankania et al.	2003/0059397 A1	3/2003	Hughes
5,431,922 A	7/1995	Nicklasson	2003/0064099 A1	4/2003	Oshlack et al.
5,455,046 A	10/1995	Baichwal	2003/0064122 A1	4/2003	Goldberg et al.
5,470,584 A	11/1995	Hendrickson et al.	2003/0065002 A1	4/2003	Caruso et al.
5,478,577 A	12/1995	Sackler et al.	2003/0068276 A1	4/2003	Hughes et al.
5,512,297 A	4/1996	Baichwal	2003/0068370 A1	4/2003	Sackler
5,512,578 A	4/1996	Crain et al.	2003/0068371 A1	4/2003	Oshlack et al.
5,554,387 A	9/1996	Baichwal	2003/0068375 A1	4/2003	Wright et al.
5,567,754 A	10/1996	Stramel	2003/0068392 A1	4/2003	Sackler
5,580,578 A	12/1996	Oshlack et al.	2003/0069263 A1	4/2003	Breder et al.
5,612,053 A	3/1997	Baichwal et al.	2003/0073714 A1	4/2003	Breder et al.
5,629,011 A	5/1997	Illum	2003/0091635 A1	5/2003	Baichwal et al.
5,633,000 A	5/1997	Grossman et al.	2003/0124061 A1	7/2003	Roberts
5,639,476 A	6/1997	Oshlack et al.	2003/0124185 A1	7/2003	Oshlack et al.
5,662,933 A	9/1997	Baichwal et al.	2003/0125347 A1	7/2003	Anderson et al.
5,672,360 A	9/1997	Sackler et al.	2003/0129234 A1	7/2003	Baichwal et al.
5,738,865 A	4/1998	Baichwal et al.	2003/0143269 A1	7/2003	Oshlack et al.
5,858,388 A	1/1999	Grossman et al.	2003/0147975 A1	8/2003	Joshi et al.
5,891,474 A	4/1999	Busetti et al.	2003/0152638 A1	8/2003	Tice et al.
5,914,131 A	6/1999	Merrill et al.	2003/0157167 A1	8/2003	Kao et al.
5,948,438 A	9/1999	Stanforth et al.	2003/0157168 A1	8/2003	Breder et al.
5,958,452 A	9/1999	Oshlack et al.	2003/0158264 A1	8/2003	Radhakrishnan et al.
5,958,456 A	9/1999	Baichwal et al.	2003/0163099 A1	8/2003	Wermeling et al.
5,958,458 A	9/1999	Noring et al.	2003/0170181 A1	9/2003	Midha
5,958,459 A	9/1999	Chasin et al.	2003/0190362 A1	10/2003	Sackler et al.
5,965,161 A	10/1999	Oshlack et al.	2007/0098792 A1	5/2007	Kao et al.
5,965,163 A	10/1999	Miller et al.	2007/0098793 A1	5/2007	Kao et al.
5,968,551 A	10/1999	Oshlack et al.	2007/0098794 A1	5/2007	Kao et al.
RE36,547 E	2/2000	Crain et al.	2007/0134328 A1	6/2007	Kao et al.
6,039,980 A	3/2000	Baichwal et al.	2007/0140975 A1	6/2007	Baichwal et al.
6,093,420 A	7/2000	Baichwal	2008/0050431 A1	2/2008	Baichwal et al.
6,103,258 A	8/2000	Simon	2008/0085303 A1	4/2008	Baichwal et al.
6,103,261 A	8/2000	Chasin et al.	2008/0085304 A1	4/2008	Baichwal et al.
6,129,933 A	10/2000	Oshlack et al.	2008/0085305 A1	4/2008	Baichwal et al.
6,143,322 A	11/2000	Sackler et al.	2008/0119501 A1	5/2008	Hein et al.
6,143,325 A	11/2000	Dennis et al.	2008/0262013 A1	10/2008	Kao et al.
6,166,211 A	12/2000	Cain et al.	2008/0318993 A1	12/2008	Ahdieh
6,228,398 B1	5/2001	Devane et al.	2008/0318994 A1	12/2008	Ahdieh
6,228,863 B1	5/2001	Palermo et al.			
6,245,351 B1	6/2001	Nara et al.			
6,245,357 B1	6/2001	Edgren et al.			
6,248,789 B1	6/2001	Weg			
6,261,599 B1	7/2001	Oshlack et al.			
6,277,384 B1	8/2001	Kaiko et al.			
6,294,195 B1	9/2001	Oshlack et al.			
6,296,842 B1	10/2001	Jaworowicz et al.			
6,306,425 B1	10/2001	Tice et al.			
6,309,668 B1	10/2001	Bastin et al.			
6,316,031 B1	11/2001	Oshlack et al.			
6,340,475 B2	1/2002	Shell et al.			
6,375,957 B1	4/2002	Kaiko et al.			
6,387,394 B1	5/2002	Baichwal et al.			
6,391,336 B1	5/2002	Royer			
6,413,494 B1	7/2002	Lee et al.			
6,432,438 B1	8/2002	Shukla			

FOREIGN PATENT DOCUMENTS

CA	2369302 A1	10/2000
DE	1 517 480 A1	7/1978
EP	0 253 104 A1	1/1988
EP	319243 A1	6/1989
EP	0360562 B2	3/1990
EP	441833 B1	9/1993
EP	0636366 A2	2/1995
EP	751766 A1	1/1997
EP	0 793 959 A1	9/1997
EP	742711 B1	3/1999
EP	1293195 A1	3/2003
EP	1293209-AI A1	3/2003
EP	1293209-AI	3/2003
JP	2003113074 A	4/2003
NZ	0505192	7/1999
WO	80/00841 A1	5/1980

US 8,329,216 B2

Page 3

WO	84/00488 A1	2/1984	randomized crossover study." <i>Pharmacotherapy</i> , vol. 24(4), pp. 468-476 (2004).
WO	84/00490 A1	2/1984	Adams & Ahdieh, "Pharmacokinetic Analyses of New Formulations of Extended- and Immediate-Release Oxymorphone." <i>Amer. Acad. Pain Management</i> Mar. 2004, Poster Presentation.
WO	85/02540 A1	6/1985	Ahdieh et al., "Oxymorphone Extended Release Provides Safe and Effective Analgesia For Cancer Patients: A Randomized, Double-Blind, Crossover Study with Oxycodone Controlled Release." <i>Amer. Pharmacists Assoc.</i> , Mar. 2004, Poster Presentation.
WO	85/02542 A1	6/1985	Ahdieh et al., "Efficacy of Oxymorphone extended release in post-surgical pain: A randomized clinical trial in knee arthroplasty." <i>J. Clinical Pharmacology</i> , vol. 44, pp. 767-776 (2004).
WO	91/07950 A1	6/1991	Ansel H.C. et al., <i>Pharmaceutical Dosage Forms and Drug Delivery Systems</i> , pp. 121-122, (7th Ed.) (1999).
WO	93/17673 A1	9/1993	Baichwal et al., "Gamma Scintigraphy Imaging and Pharmacokinetic Analysis of Extended-Release TIMERx Technology." <i>Amer. Physiological Soc.</i> , May 2004, Poster Presentation.
WO	95/20947 A1	8/1995	Beaver, et al., "Comparisons of the Analgesic Effects of Oral and Intramuscular Oxymorphone and of Intramuscular Oxymorphone and Morphine in Patients with Cancer." <i>J Clinical Pharmacology</i> , vol. 17(4), pp. 186-198 (1977).
WO	95/22965 A2	8/1995	Cass, <i>Use of Oral Analgesic for Severe Pain</i> . Western Medicine, pp. 107-108, 120 (Mar. 1961).
WO	96/00047 A1	1/1996	Cass et al., "The Control of Severe Pain with Oral Oxymorphone Hydrochloride." <i>Current Therapeutic Research</i> , vol. 5(11) pp. 579-586 (1963).
WO	96/02251 A1	2/1996	Cephalon, Package Insert for ACTIQ®, 2007.
WO	96/04007 A1	5/1996	Chiao et al., Sustained-Released Drug Delivery Systems, Chapter 94, Remington 1995.
WO	96/20927 A1	7/1996	Cisternas et al., "Management of Chronic Pain With Long-Acting Opioids May Contribute to the Stabilization of Total Healthcare Costs: Results of a Retrospective Claims Database Pharmacoeconomic Study." <i>Amer. Soc. Health-System Pharmacists</i> , Dec. 2004, Poster Presentation.
WO	97/07750 A1	3/1997	Cone, "General procedure for the isolation and identification of 6- α - and 6- β -hydroxymetabolites of narcotic agonists and antagonists with a hydromorphone structure." <i>J. of Chromatogr</i> , vol. 129, pp. 355-361 (1976).
WO	97/16172 A1	5/1997	Cone et al., "Oxymorphone metabolism and urinary excretion in human, rat, guinea pig, rabbit, and dog." <i>Drug Metabolism and Disposition</i> , vol. 11(5), pp. 446-450 (1983).
WO	98/00143 A1	1/1998	Dhopeshwarkar et al., "Evaluation of Xanthan Gum in the Preparation of Sustained Release Matrix Tablets." <i>Drug Development & Industrial Pharmacy</i> , vol. 19(9), pp. 999-1017 (1993).
WO	99/01111 A1	1/1999	Drakontides, "Drugs to Treat Pain." <i>Amer. J Nursing</i> , vol. 74(3), pp. 508-513 (Mar. 1974).
WO	99/32119 A1	7/1999	Eames et al., "Clinical Trial of Oxymorphone in Labour." <i>Brit. Med. J.</i> , vol. 2, pp. 353-355 (1964).
WO	99/32120 A1	7/1999	Eddy & Lee, "The analgesic equivalence to morphine and relative side action liability of oxymorphone (4-Hydroxydihydromorphinone)." <i>J Pharmacology and Experimental Therapeutics</i> , vol. 125(2), pp. 116-121 (1959).
WO	00/01377 A2	1/2000	Endo Pharmaceuticals, Package Insert for NUMORPHAN®, Apr. 2004.
WO	00/21520 A3	4/2000	Endo Pharmaceuticals, Package Insert for OPANA® (Jul. 2006).
WO	00/33835 A1	6/2000	Endo Pharmaceuticals, Package Insert for PERCOSET®, Nov. 2006.
WO	00/38649 A1	7/2000	Endo Pharmaceuticals, Package Insert for ZYDONE®, Jun. 2003.
WO	00/61147 A1	10/2000	Gabrail et al., "Oxymorphone Extended Release Provides Safe and Effective Analgesia During Opioid Rotation: Results of Randomized, Double-Blind, Crossover, Comparative Study with Oxycodone Controlled Release." <i>Amer. Acad. Pain Management</i> , Mar. 2004, Poster Presentation.
WO	01/00181 A2	1/2001	Gabrail et al., "Establishing the dosage equivalency of oxymorphone extended release and oxycodone controlled release in patients with cancer pain: A randomized controlled study." <i>Current Medical Research Opin.</i> , vol. 20(6), pp. 911-918 (2004).
WO	01/08661 A2	2/2001	Galer et al., "Safety of New Oral Formulations of the Opioid Oxymorphone." <i>Int'l Assoc. for the Study of Pain</i> , May 2004, Poster Presentation.
WO	01/12230 A1	2/2001	Gallagher et al., "Assessment of dosing frequency of sustained-release opioid preparations in patients with chronic nonmalignant pain." <i>Pain Medicine</i> , vol. 8(1), pp. 71-74 (2004).
WO	01/15699 A1	3/2001	
WO	01/32148 A1	5/2001	
WO	01/15699 A1	7/2001	
WO	01/52813 A1	7/2001	
WO	01/58447 A1	8/2001	
WO	01/58451 A1	8/2001	
WO	02/05647 A1	1/2002	
WO	02/13886 A2	2/2002	
WO	02/087558 A1	11/2002	
WO	02/092059 A1	11/2002	
WO	02/092060 A1	11/2002	
WO	02/094172 A2	11/2002	
WO	02/094254 A2	11/2002	
WO	03/004029 A1	1/2003	
WO	03/004030 A1	1/2003	
WO	03/007802 A2	1/2003	
WO	03/013433 A2	2/2003	
WO	03/013476 A1	2/2003	
WO	03/013479 A1	2/2003	
WO	03/013525 A1	2/2003	
WO	03/013538 A1	2/2003	
WO	03/015531 A2	2/2003	
WO	03/026743 A	4/2003	
WO	03/039561 A1	5/2003	
WO	03/072106 A2	9/2003	
WO	2006/094083 A1	9/2006	
WO	2007/053698 A2	5/2007	
WO	2007/078895 A2	7/2007	

OTHER PUBLICATIONS

- U.S. Appl. No. 11/766,748, Ahdieh.
- U.S. Appl. No. 11/742,956, GerritsenvanderHoop.
- U.S. Appl. No. 12/203,758, Ahdieh.
- Abbott Laboratories, Package Insert for VICODIN®, Mar. 2007.
- Adams & Ahdieh, "Single- and Multiple-Dose Pharmacokinetic and Dose-Proportionality Study of Oxymorphone Immediate Release Tablets." *Drugs R D*, vol. 6(2), pp. 91-99 (2005).
- Adams & Ahdieh, "Single- and Multiple-Dose Pharmacokinetic and Dose-Proportionality Study of Oxymorphone Immediate Release Tablets." *Amer. Pharmacists Assoc.*, Mar. 2004, Poster Presentation.
- Adams et al., "Oxymorphone Extended Release Does Not Affect CYP2C9 or CYP2C9 or CYP3A4 Metabolic Pathways." *J Clinical Pharmacology*, vol. 45, pp. 337-345 (2005).
- Adams et al., "Oxymorphone Extended Release Does Not Affect CYP2C9 or CYP3A4 Metabolic Pathways." *Amer. Acad. Pain Management* Mar. 2004, Poster Presentation.
- Adams et al., "A New Oral Opioid, Oxymorphone Extended Release, Does Not Affect Human Metabolic Enzymes CYP2C9 or CYP3A4." *Amer. Pharmacists Assoc.*, Mar. 2004, Poster Presentation.
- Adams & Ahdieh, "Pharmacokinetics and dose-proportionality of oxymorphone extended release and its metabolites: Results of a randomized crossover study." *Pharmacotherapy*, vol. 24(4), pp. 468-476 (2004).

US 8,329,216 B2

Page 4

- Gibofsky & Barkin, "Chronic Pain of Osteoarthritis: Considerations for Selecting an Extended Release Opioid Analgesic." Amer. J Therapeutics, vol. 15, pp. 241-255 (2008).
- Gimbel & Adams, "Oxymorphone Immediate Release for Postsurgical Pain: Translating Pharmacokinetic Drug Properties Into Clinical Efficacy." Amer. Cancer Soc. (Abstract). 2004.
- Gimbel & Ahdieh, "The Efficacy and Safety of Oral Immediate-Release Oxymorphone for Postsurgical Pain." Anesth. Analg., vol. 99, pp. 1472-1477 (2004).
- Gimbel & Walker, "A Randomized Double-Blind Trial of Low-Dose Oxymorphone Immediate Release (5 mg) for Mild to Moderate Pain in Ambulatory Patients." Amer. Physiological Soc., May 2004, Poster Presentation.
- Gimbel et al., "Analgesic Efficacy of Oxymorphone Immediate Release in Postsurgical Orthopedic Pain: Results of a Randomized, Double-Blind, Placebo-Controlled, Dose-Ranging Comparison with Oxycodone." Amer. Pharmacists Assoc., Mar. 2004, Poster Presentation.
- Gimbel et al., "Low-Dose Oxymorphone Immediate Release (5mg) for Mild to Moderate Pain Following Arthroscopic Knee Surgery in Ambulatory Patients: A Randomized Double-Blind Trial." Amer. Acad. Nurse Practitioners, Jun.-Jul. 2004, Poster Presentation.
- Gimbel et al., "Efficacy and safety of oxymorphone immediate release for the treatment of mild to moderate pain after ambulatory orthopedic surgery: results of a randomized, double-blind, placebo-controlled trial." Archives of Physical Medicine Rehabilitation, vol. 86(12), pp. 2284-2289 (2005).
- Gould et al., "Retrospective and Prospective Analyses of Oxymorphone Extended Release for Neuropathic Pain." Neuropathic Pain, Nov. 2004 (Abstract).
- Gould et al., "Effective Titration With Oxymorphone Extended Release in Opioid-Naïve Patients with Low Back Pain." Amer. Acad. Pain Management, Feb. 2005 (Abstract).
- Gould et al., "Effective long-term management of opioid-naïve patients with oxymorphone extended release." Amer. Physiologic Soc., Mar. 2005 (Abstract).
- Gould et al., "Oxymorphone extended release for effective long-term treatment of elderly opioid-naïve patients with moderate to severe nonmalignant pain." Amer. Physiologic Soc., Mar. 2005, (Abstract).
- Gould & Ahdieh, "Case Studies of Opioid Treatments for Neuropathic Pain." Neuropathic Pain, Nov. 2004 (Abstract).
- Hale & Drass, "Safety, Tolerability, and Effectiveness of Oxymorphone Extended Release in Opioid-Naïve Patients: An Interim Analysis." Amer. Orthopedic Assoc., Nov. 2004, Poster Presentation.
- Hale et al., "Long-Term Safety and Efficacy of Oxymorphone Extended Release in Opioid-Naïve Patients with Chronic Pain." Amer. Acad. Pain Management, Mar. 2004, Poster Presentation.
- Hale et al., "Efficacy and safety of oxymorphone extended release in chronic low back pain: results of a randomized, double-blind, placebo- and active-controlled phase III study." J Pain, vol. 6(1), pp. 21-28 (2005).
- Hale et al., "Open-Label Long-Term Assessment of Tolerability, Safety, and Effectiveness of Oxymorphone Extended Release for Low Back Pain." Amer. Acad. Pain Management, Mar. 2004, Poster Presentation.
- Hale et al., "Tolerability and Effectiveness of Oxymorphone Extended Release in Opioid-Naïve Patients with Chronic Pain." Amer. Pharmacists Assoc., Mar. 2004, Poster Presentation.
- Hale et al., "Low-Dose Titration of Oxymorphone Extended Release in Opioid-Naïve Patients With Chronic Pain: Short- and Long-Term Results." Amer. Acad. Of Physical Medicine and Rehabilitation, Oct. 2004, Poster Presentation.
- Hinz et al., "Bioavailability of diclofenac potassium at low doses", British Journal of Clinical Pharmacology, vol. 59, No. 1, pp. 80-84 (2005).
- International Search Report issued in PCT/US02/21403, mailed Oct. 31, 2002.
- International Search Report issued in PCT/US02/21396, mailed Nov. 6, 2002.
- International Search Report issued in PCT/US02/21398, mailed Nov. 6, 2002.
- International Search Report issued in PCT/US02/21400, mailed Oct. 31, 2002.
- International Search Report issued in PCT/US02/21354, mailed Nov. 6, 2002.
- Kafka et al., "Effective Titration With Oxymorphone Extended Release for Opioid-Naïve Osteoarthritis Patients with Moderate to Severe Pain." Amer. Acad. Pain Management, Feb. 2005 (Abstract).
- King Pharmaceuticals, Package Insert for AVINZA®, Oct. 2006.
- Kivitz et al., "A 2-week, multicenter, randomized, double-blind, placebo-controlled, dose-ranging, phase III trial comparing the efficacy of oxymorphone extended release and placebo in adults with pain associated with osteoarthritis of the hip or knee." Clinical Therapeutics, vol. 28(3), pp. 352-364 (2006).
- Loan et al., "Studies of drugs given before anaesthesia, XVII: The natural and semi-synthetic opiates." Brit. J. Anaesth., vol. 41, pp. 57-63 (1969).
- Matsumoto et al., "Oxymorphone extended-release tablets relieve moderate to severe pain and improve physical function in osteoarthritis: Results of randomized, double-blind, placebo- and active-controlled phase III trial." Pain Medicine, vol. 6(5), pp. 357-366 (2005).
- McIlwain, "Safety and Tolerability of Oxymorphone Extended Release During Long-Term Treatment of Moderate to Severe Pain From Osteoarthritis." ASCP, Nov. 2004, Poster Presentation.
- McIlwain, "Safety and Effectiveness of Oxymorphone Extended Release During Long-Term Treatment of Moderate to Severe Pain from Osteoarthritis: One-Year Results." Amer. Osteopathic Assoc., Nov. 2004, Poster Presentation.
- McIlwain et al., "Oxymorphone Extended Release Maintains Effectiveness and Is Well Tolerated During Long-Term Treatment of Moderate to Severe Osteoarthritis Pain." Amer. Acad. Pain Management, Mar. 2004, Poster Presentation.
- McIlwain & Ahdieh, "Long-Term Effectiveness and Safety of a New Oral Opioid, Oxymorphone Extended Release, for Moderate to Severe Osteoarthritis Pain." Amer. Pharmacists Assoc., Mar. 2004, Poster Presentation.
- McIlwain & Ahdieh, "Safety, Tolerability, and Effectiveness of Oxymorphone Extended release for moderate to severe osteoarthritis pain: a one-year study." Amer. J Therapeutics, vol. 11(5), pp. 1-7 (2004).
- Numorphan Oral Advertisement, British Journal of Anaesthesia, vol. 34, No. 8 (Aug. 1962).
- News of Products and Services, Products for Dispensing: Numorphan Oral. The Pharmaceutical Journal (Jul. 7, 1962).
- Ossipov & Porreca, "Challenges in the Development of Novel Treatment Strategies for Neuropathic Pain." J. Amer. Society for Experimental Neurotherapeutics, vol. 2, pp. 650-661 (2005).
- PFORMULATE: <http://www.pformulate.com/disintegrals.html> (published on May 5, 2000).
- Pieniaszek et al., "Oxymorphone Does Not Affect CYP450 Enzymes 2C9 or 3A4: Positive Implications for Pain Management." Amer. Physiological Soc., May 2004, Poster Presentation.
- Pieniaszek et al., "Oxymorphone Exhibits a Low Potential for Drug Interactions Through CYP450 Isozyme 3A4: In Vitro and Clinical Studies." Amer. Acad. Physical Medicine and Rehabilitation, Oct. 2004 (Abstract).
- Plummer et al., "Influence of polarity on dose-response relationships of intrathecal opioids in rats." Pain, vol. 40, pp. 339-347 (1989).
- Prager & Rauck, "Oxymorphone Extended Release for Moderate to Severe Neuropathic Pain: Open-Label, Long-Term Study of Safety and Effectiveness." Amer. Physiological Soc., May 2004, Poster Presentation.
- Purdue Pharma L.P., Package Insert for MS CONTIN®, 2007.
- Rauck, "Oxymorphone Extended Release for Moderate to Severe Neuropathic Pain." ASCP, Nov. 2004, Poster Presentation.
- Rhiner et al., "Long-Term Safety, Effectiveness, and Dose Stabilization of Oxymorphone Extended Release in Cancer Pain." Amer. Acad. Nurse Practitioners, Jun. 2004, Poster Presentation.
- Rowland & Tozer *Clinical Pharmacokinetics: Concepts and Applications*, pp. 152-160, 392-393 (2d ed. 1989).
- Sargent et al., "Hydroxylated Codeine Derivatives." J. Org. Chem., vol. 23, pp. 1247-1251 (Sep. 1958).

US 8,329,216 B2

Page 5

- Shuey et al., "Reproductive and Developmental Toxicity Studies with Oxymorphone: A Potent Opioid Analgesic." Teratology Soc., Jun. 2004, Poster Presentation.
- Slatkin et al., "Oxymorphone Maintains Effectiveness and Is Well Tolerated During Long-Term Treatment of Moderate to Severe Cancer Pain." Amer. Acad. Pain Management, Mar. 2004, Poster Presentation.
- Slatkin et al., "Long-Term Effectiveness and Safety of a New Oral Opioid, Gxmorphone Extended Release, for Moderate to Severe Cancer Pain." Amer. Pharmacists Assoc., Mar. 2004, Poster Presentation.
- Slatkin et al., "Case Studies of Cancer Patients with Neuropathic Pain Treated with Oxymorphone." Int'l Assoc. for the Study of Pain, May 2004, Poster Presentation.
- Slatkin et al., "Long-Term Treatment of Moderate to Severe Cancer Pain: A 2-Year Study." Amer. Physiological Soc., May 2004, Poster Presentation.
- Slatkin et al., "Effectiveness, safety, and tolerability of oxymorphone extended release for moderate to severe cancer pain." Amer. Soc. Clinical Oncology, Jun. 2004 (Abstract).
- Sloan et al., "Effectiveness and safety of oral extended-release oxymorphone for the treatment of cancer pain: a pilot study." Supportive Care Cancer, vol. 13(1), pp. 57-65 (2005).
- Swerdlow & Brown, "Numorphan: A new supplement to anaesthesia." Brit. J. Anaesth., vol. 33, pp. 126-129 (1961).
- Tark et al., "Safety, Tolerability, and Effectiveness of Oxymorphone Extended Release During Long-Term Treatment of Cancer Pain: Results of a 12-month Open-Label Study." Multi-National Assoc. Supportive Cancer Care, Jun. 2004, Poster Presentation.
- Vashi et al., "Oral and I.V. oxymorphone clinical pharmacokinetics compared to those of oral oxycodone pharmacokinetics." ASHP Mid-year Clinical Meeting, vol. 39, pp. P435E (2004) (abstract of meeting presentation).
- Walker et al., "A Randomized Double-Blind Trial of Low-Dose Oxymorphone Immediate Release (5 mg) for Mild to Moderate Pain in Ambulatory Patients." Amer. Pharmacists Assoc., Mar. 2004, Poster Presentation.
- Weiss, "Derivatives of Morphine. IV. 14-Hydroxymorphone and 14-Hydroxydihydromorphone." J. Med. Chem., vol. 8, pp. 123-125 (Jan. 1965).
- White et al., "Application of Propensity Scores to Claims Data: Correcting Biases in Drug-Prescribing Patterns of Long-Acting Opioids." Amer. Soc. Health-System Pharmacists, Dec. 2004, Poster Presentation.
- White et al., "Improved quality of life during long-term treatment of moderate to severe pain with oxymorphone ER." Amer. Physiologic Soc., Mar. 2005 (Abstract).
- White, "Comment: therapy switching in patients receiving long-acting opioids." Annals of Pharmacotherapy, vol. 38(10), pp. 1752-1752 (2004).
- Wikipedia entry for Oxymorphone, 3 pages, last modified on Dec. 20, 2007.
- Zeller et al., "Acute Pain Treatment." JAMA vol. 299(1) (2008).
- McConville et al., "Use of a Novel Modified TSI for the Evaluation of Controlled-Release Aerosol Formulations. I," Drug Dev Ind Pharmacy, vol. 26, No. 11, pp. 1191-1198 (2000).
- Complaint, Endo Pharmaceuticals, Inc. And Penwest Pharmaceuticals Inc. v. Impax Laboratories, Inc., Docket No. 1:07cv731, entered Nov. 16, 2007.
- Answer to Complaint and Counterclaim filed by Impax Laboratories, Inc., Docket No. 1:07cv731, Entered Dec. 20, 2007.
- Answer to Counterclaim by Endo Pharmaceuticals Inc. and Penwest Pharmaceuticals Co., Docket No. 1:07cv731, entered Jan. 14, 2008.
- Complaint, Endo Pharmaceuticals, Inc. and Penwest Pharmaceuticals, Inc. v. Impax Laboratories, Inc., Docket No. 1:08-CV-00057-UNA filed Jan. 25, 2008.
- Complaint, Endo Pharmaceuticals Inc. and Penwest Pharmaceuticals Co. v. Actavis South Atlantic LLC, Docket No. 2:08-cv-01563-KSH-PS, United States District Court, District of New Jersey, filed Mar. 28, 2008.
- International Search Report for PCT/US2007/005560, Sep. 17, 2007.
- United States Patent and Trademark Office, Before the Board of Appeals and Interference, "Decision on Appeal, Appeal No. 2005-0416, U.S. Appl. No. 09/970,020." Apr. 28, 2005.
- Staniforth and Baichwal, "Synergistically Interacting Heterodisperse Polysaccharides—Function in Achieving Controllable Drug Delivery." American Chemical Society, pp. 327-350 (1993).
- Mandema et al., "Characterization and validation of a pharmacokinetic model for controlled-release oxycodone." British J Clinical Pharmacology, vol. 42(6), pp. 747-756 (1996).
- Benzinger et al., "A Pharmacokinetic/Pharmacodynamic Study of Controlled-Release Oxycodone." J Pain and Symptom Management, vol. 13(2), pp. 75-82 (1997).
- Sathyam et al., "Pharmacokinetic profile of a 24-hour controlled-release OROS® formulation of hydromorphone in the presence and absence of food." BMC Clinical Pharmacology, vol. 7(2) (2007).
- Johnson et al., "Effect of Concomitant Ingestion of Alcohol on the in Vivo Pharmacokinetics of Kadian (Morphine Sulfate Extended-Release) Capsules." J of Pain, vol. 9(4), pp. 330-336 (2008).
- Cappola, "A Better Dissolution Method for Ranitidine Tablets USP." Pharmaceutical Development and Technology, vol. 6(1), pp. 11-17 (2001).
- De Haan & Lerk, "Studies on different dissolution models." Pharmaceutisch Weekblad Scientific Edition, vol. 4, pp. 191-196 (1982).
- Karasulu & Ertan, "Different geometric shaped hydrogel theophylline tablets: statistical approach for estimating drug release." II Farmaco, vol. 57, pp. 939-945 (2002).
- United States Food and Drug Administration Alert Alcohol and Palladone Interaction available at www.fda.gov/CDER/Drug/infopage/palladone/default.htm, updated Jul. 2005.
- "Don't mix alcohol & pain killers" American Running & Fitness Association, available at findarticles/p/articles/mi_m0NHF/is_6_17/ai_8649661/print?tag=artBody:coll (1999).
- McNicol et al., "Management of opioids side effects in cancer related and chronic non cancer pain: a systematic review." J of Pain 4(5), pp. 231-256 (2003).
- Weiss, "Derivatives of Morphine. I. 14-Hydroxydihydromorphone." J American Chemical Society, 77(22), p. 5891-5892 (1955).
- U.S. Appl. No. 12/426,112, Kao et al.
- Adams et al., "Oxymorphone Extended Release Does Not Affect CYP2C9 or CYP3A4 Metabolic Pathways." J Clinical Pharmacology, vol. 45, pp. 337-345 (2005).
- Baichwal et al., "Gamma Scintigraphy Imaging and Pharmacokinetic Analysis of Extended-Release TIMERx Technology." Amer. Physiological Soc., May 2004, Poster Presentation.
- Chiao et al., Sustained-Released Drug Delivery Systems, Chapter 94, Remington, 1995.
- Gimbel & Adams, "Oxymorphone Immediate Release for Postsurgical Pain: Translating Pharmacokinetic Drug Properties Into Clinical Efficacy." Amer. Cancer Soc. (Abstract).
- Gould et al., "Oxymorphone extended release for effective long-term treatment of elderly opioid-naïve patients with moderate to severe nonmalignant pain." Amer. Physiologic Soc., Mar. 2005, (Abstract).
- Hale & Drass, "Safety, Tolerability, and Effectiveness of Oxymorphone Extended Release in Opioid-Naïve Patients: An Interim Analysis." Amer. Orthopedic Assoc., Nov. 2004, Poster Presentation.
- Hale et al., "Long-Term Safety and Efficacy of Oxymorphone Extended Release in Opioid-Naïve Patients with Chronic Pain." Amer. Acad. Pain Management, Mar. 2004, Poster Presentation.
- Matsumoto et al., "Oxymorphone extended-release tablets relieve moderate to severe pain and improve physical function in osteoarthritis: Results of randomized, double-blind, placebo- and active-controlled phase III trial." Pain Medicine, vol. 6(5), pp. 357-366 (2005).
- McIlwain, "Safety and Tolerability of Oxymorphone Extended Release During Long-Term Treatment of Moderate to Severe Pain From Osteoarthritis." ASCP, Nov. 2004, Poster Presentation.
- McIlwain, "Safety and Effectiveness of Oxymorphone Extended Release During Long-Term Treatment of Moderate to Severe Pain from Osteoarthritis: One-Year Results." Amer. Osteopathic Assoc., Nov. 2004, Poster Presentation.
- Rowland & Tozer, *Clinical Pharmacokinetics: Concepts and Applications*, pp. 152-160, 392-393 (2d ed. 1989).
- Sathyam et al., "Pharmacokinetic profile of a 24-hour controlled-release OROS® formulation of hydromorphone in the presence and absence of food." BMC Clinical Pharmacology, vol. 7(2) (2007).

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PK Profile for 6-OH-Oxymorphone with PID Scores

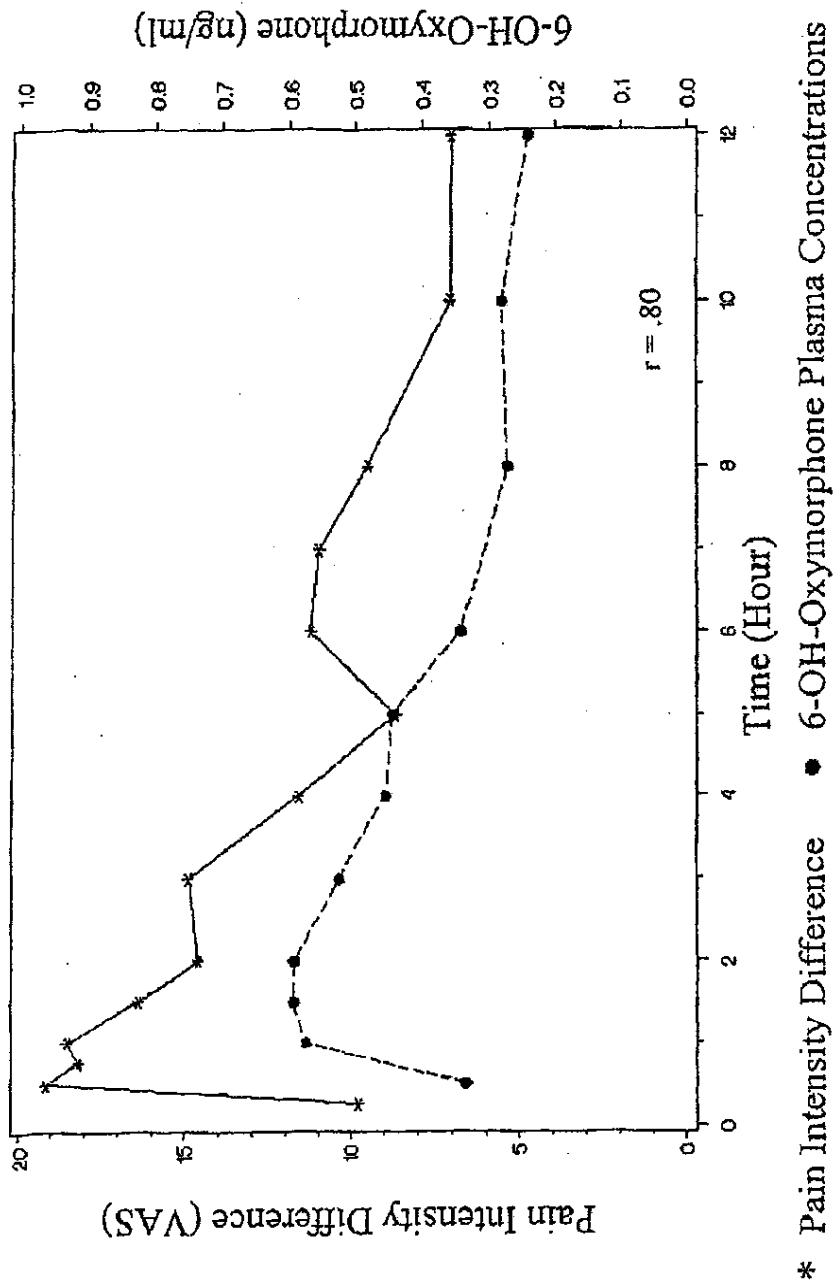


Fig. 1

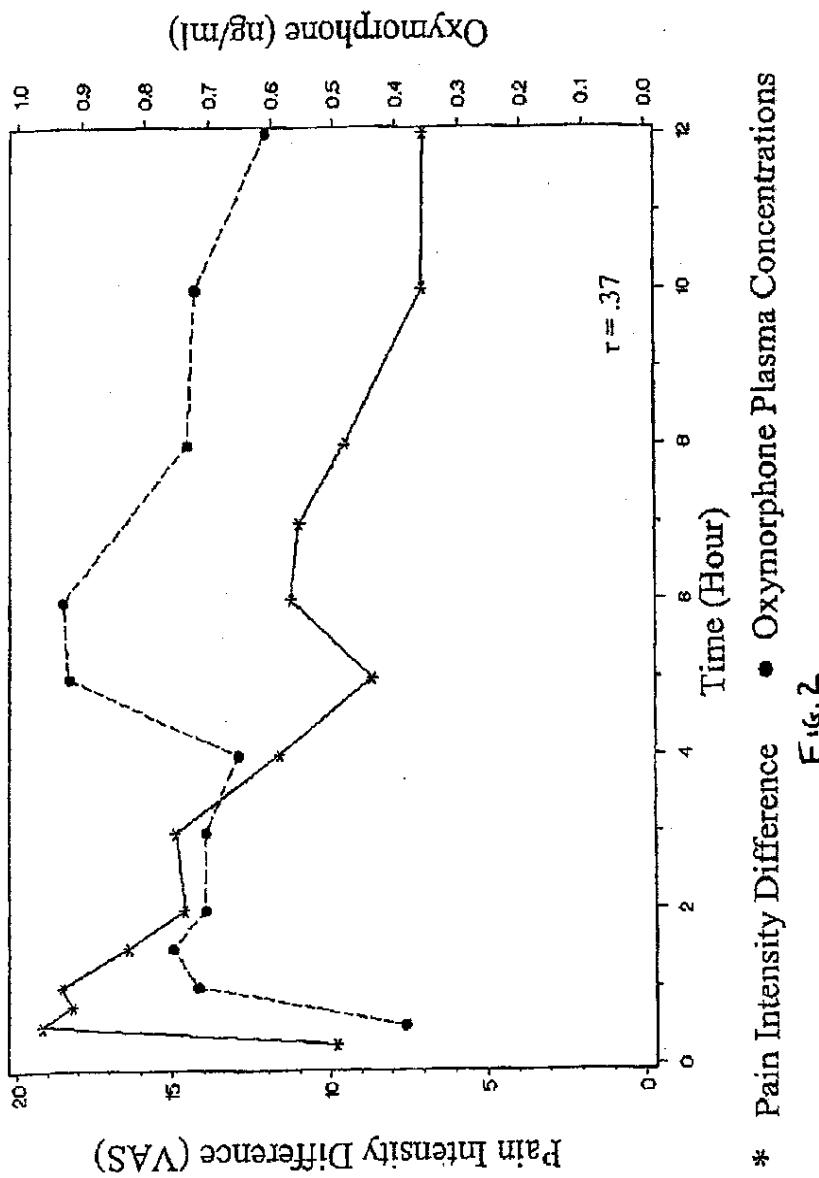
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PK Profile for Oxymorphone with PID Scores



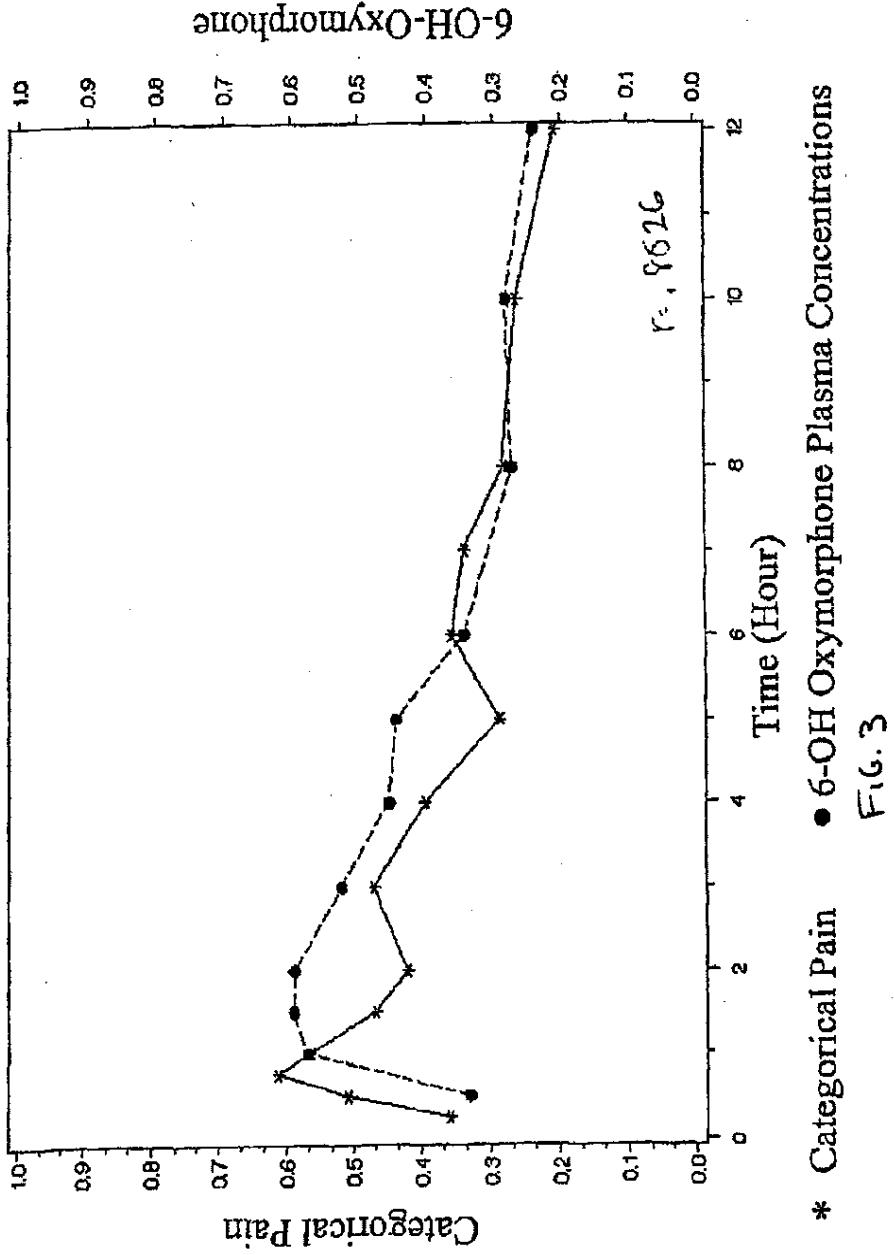
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PK Profile for 6-OH-Oxymorphone with Categorical Pain Scores



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PK Profile for Oxymorphone with Categorical Pain Scores

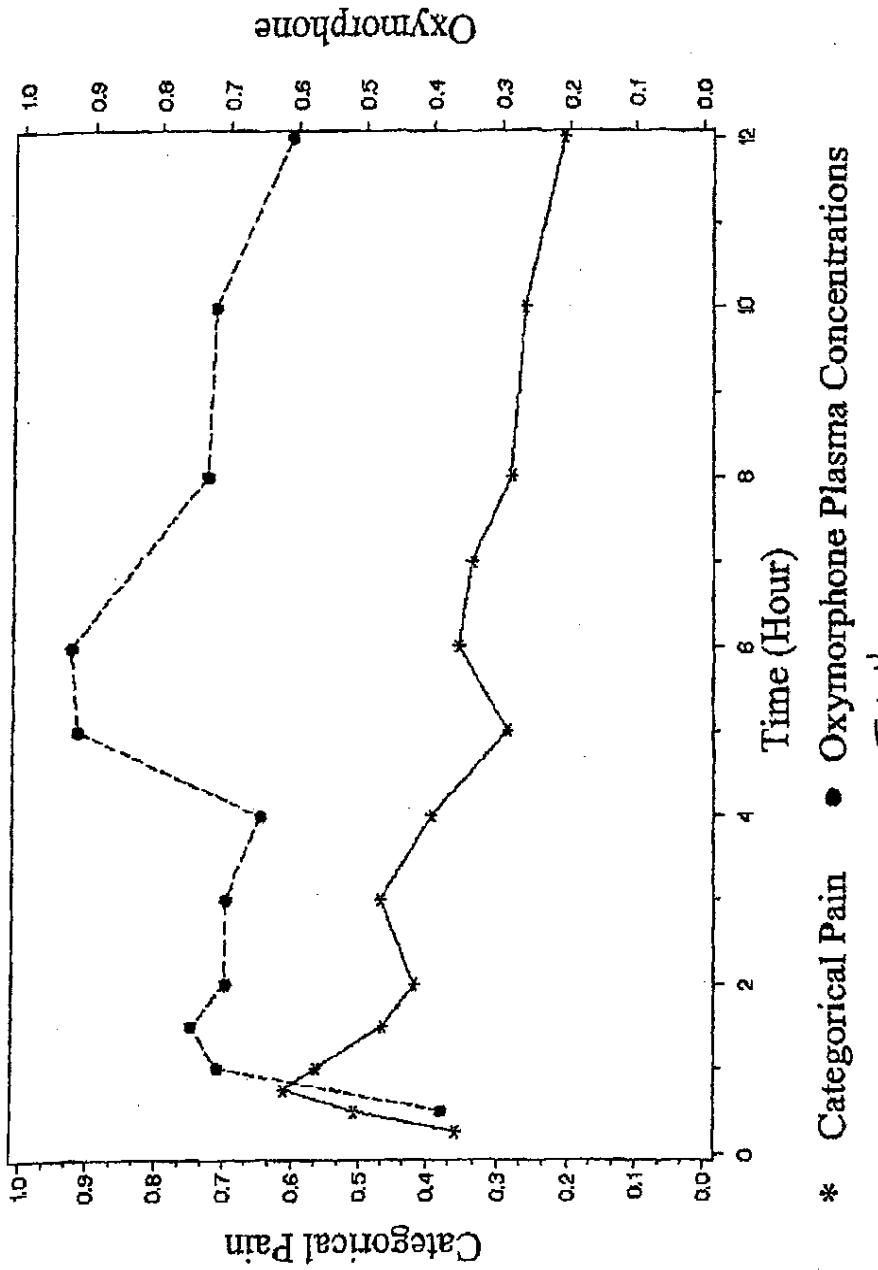


Fig. 4

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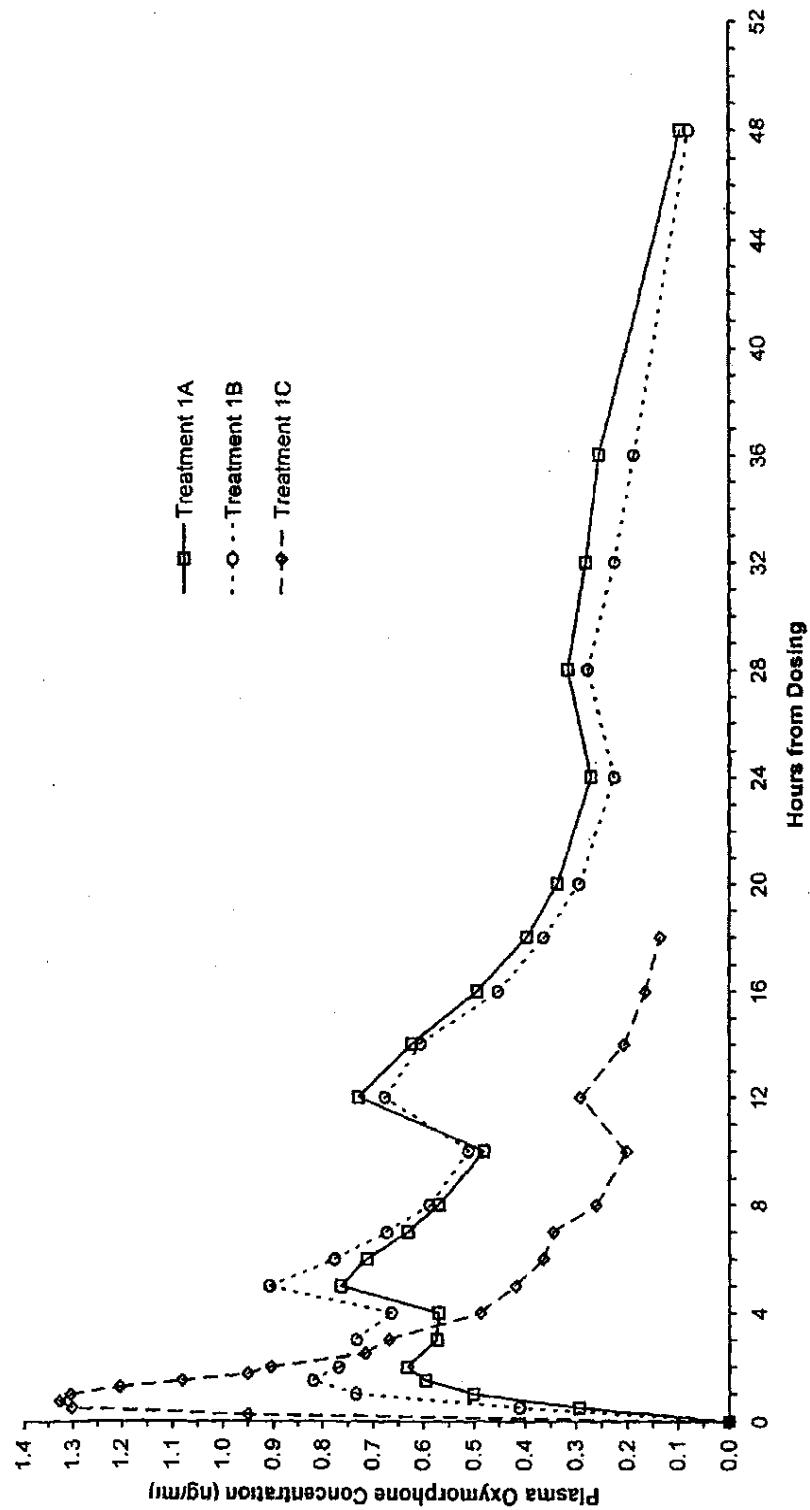


Figure 5

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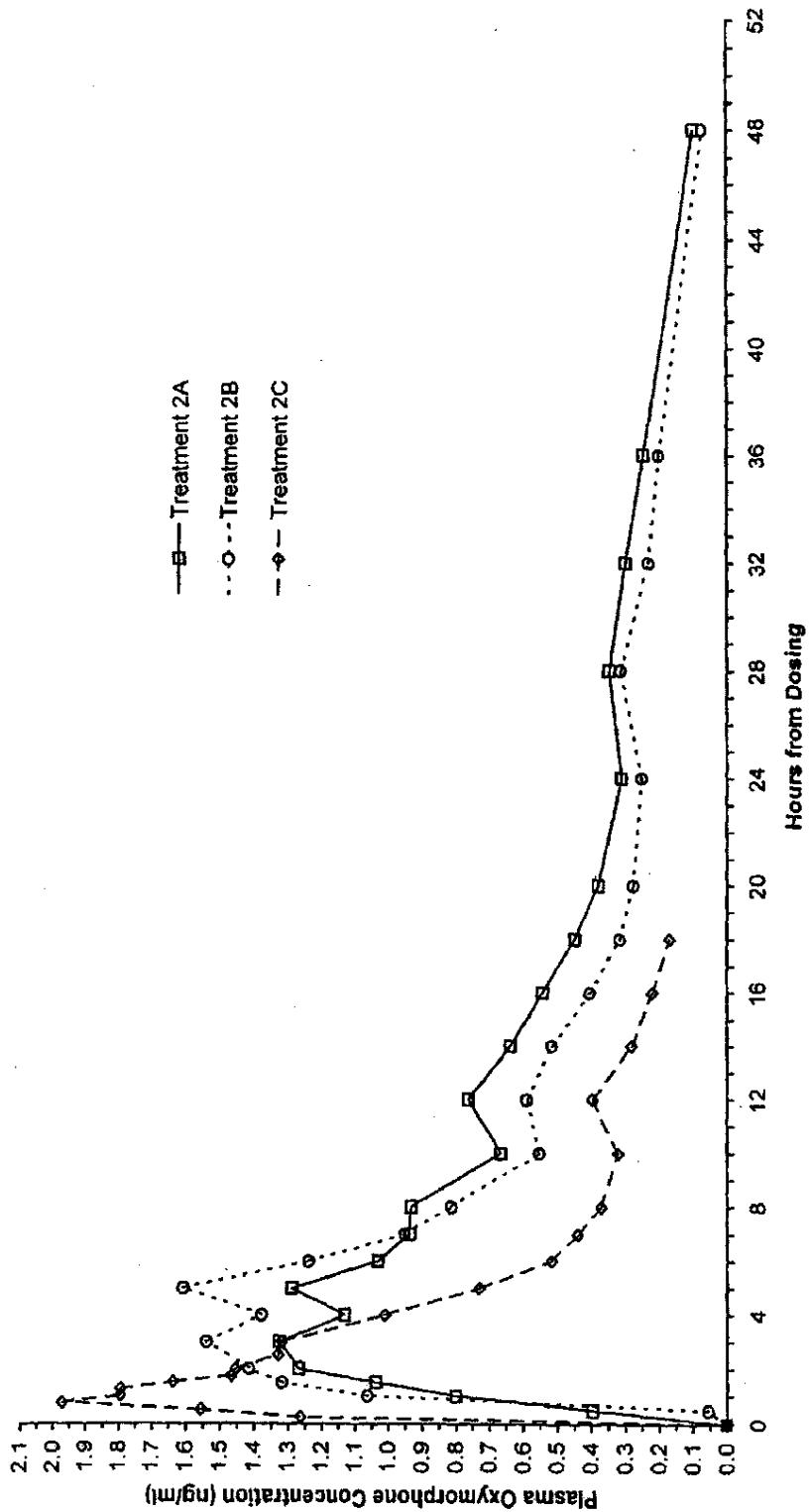


Figure 6

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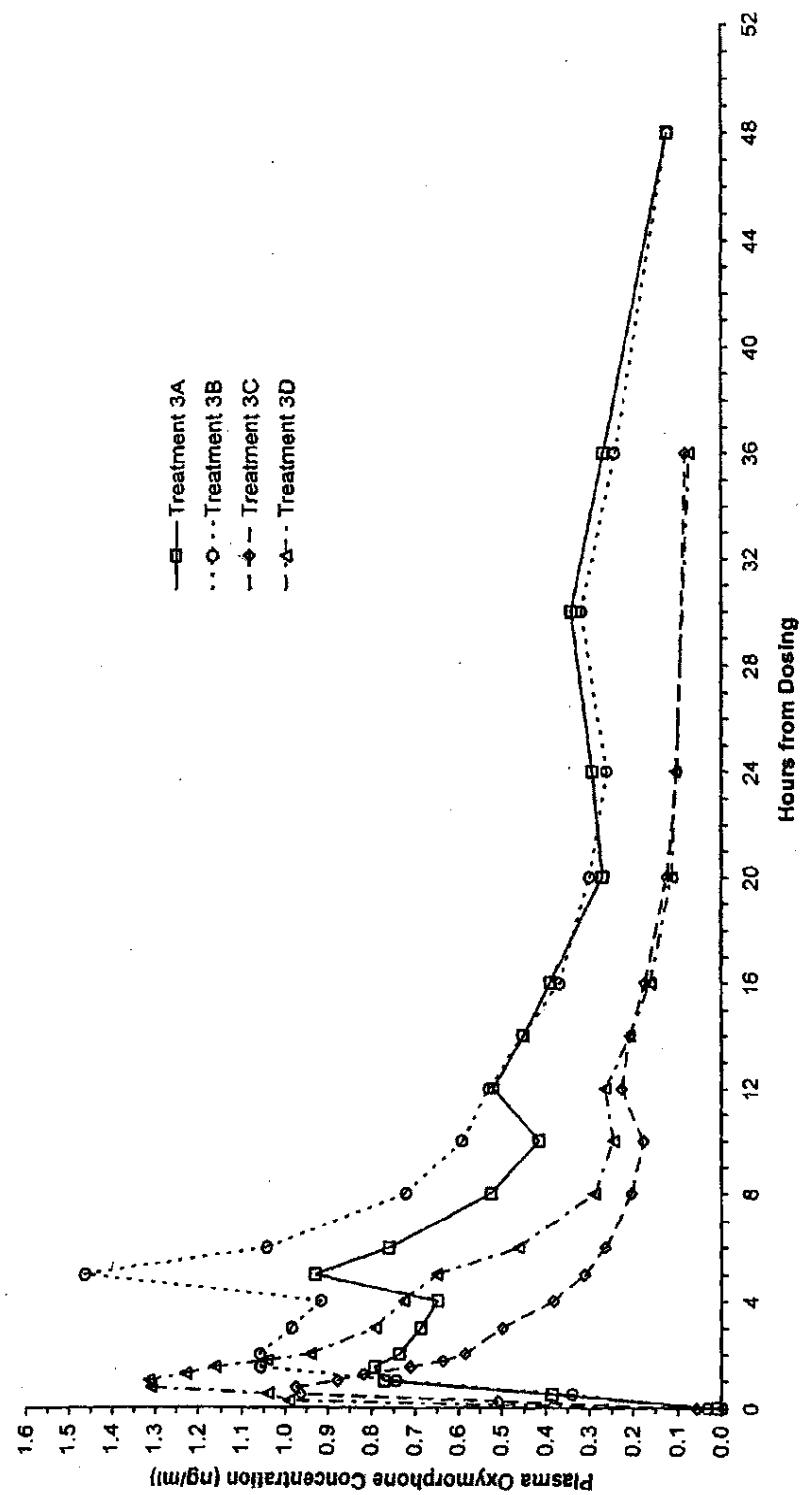


Figure 7

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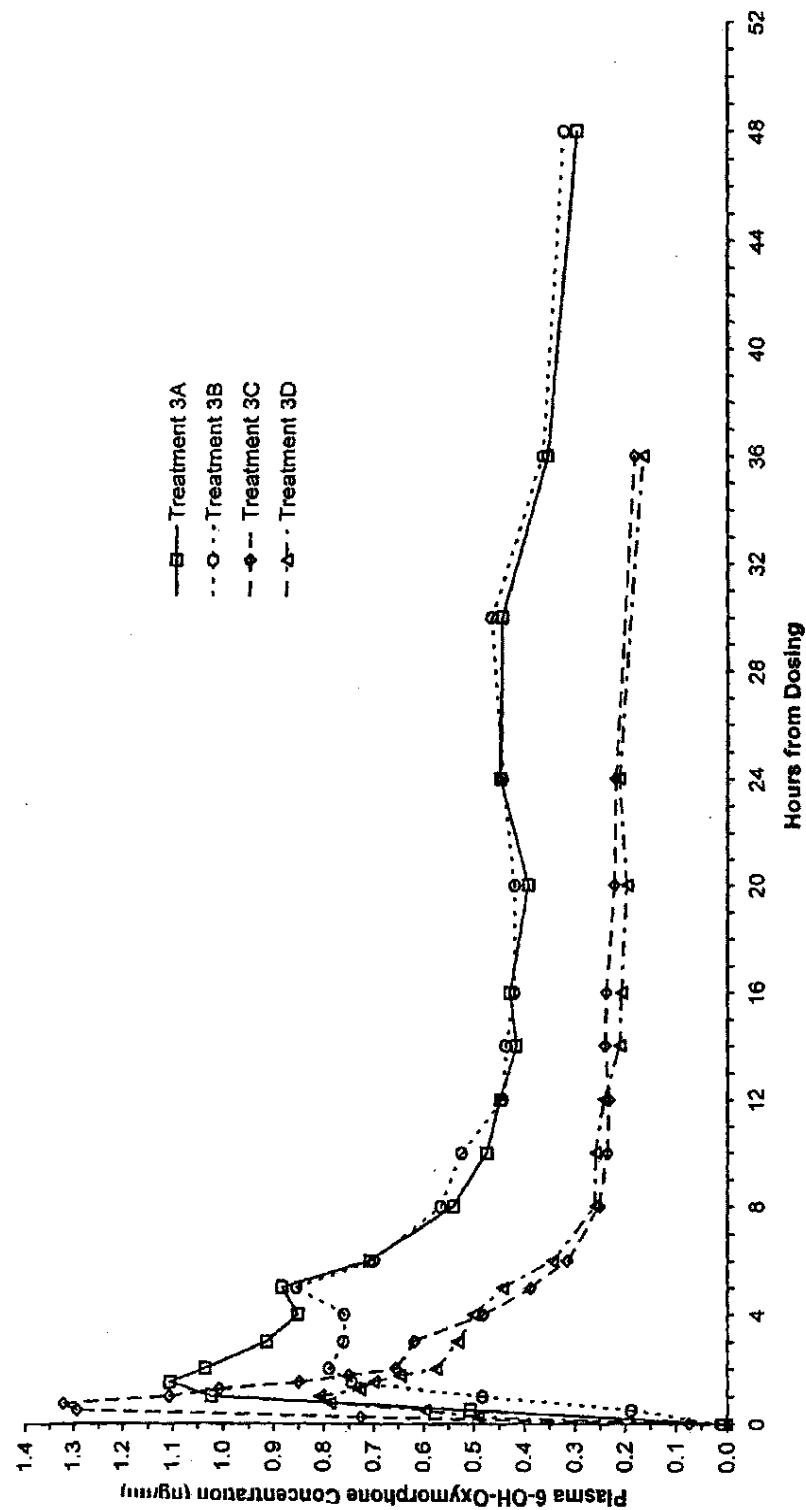


Figure 8

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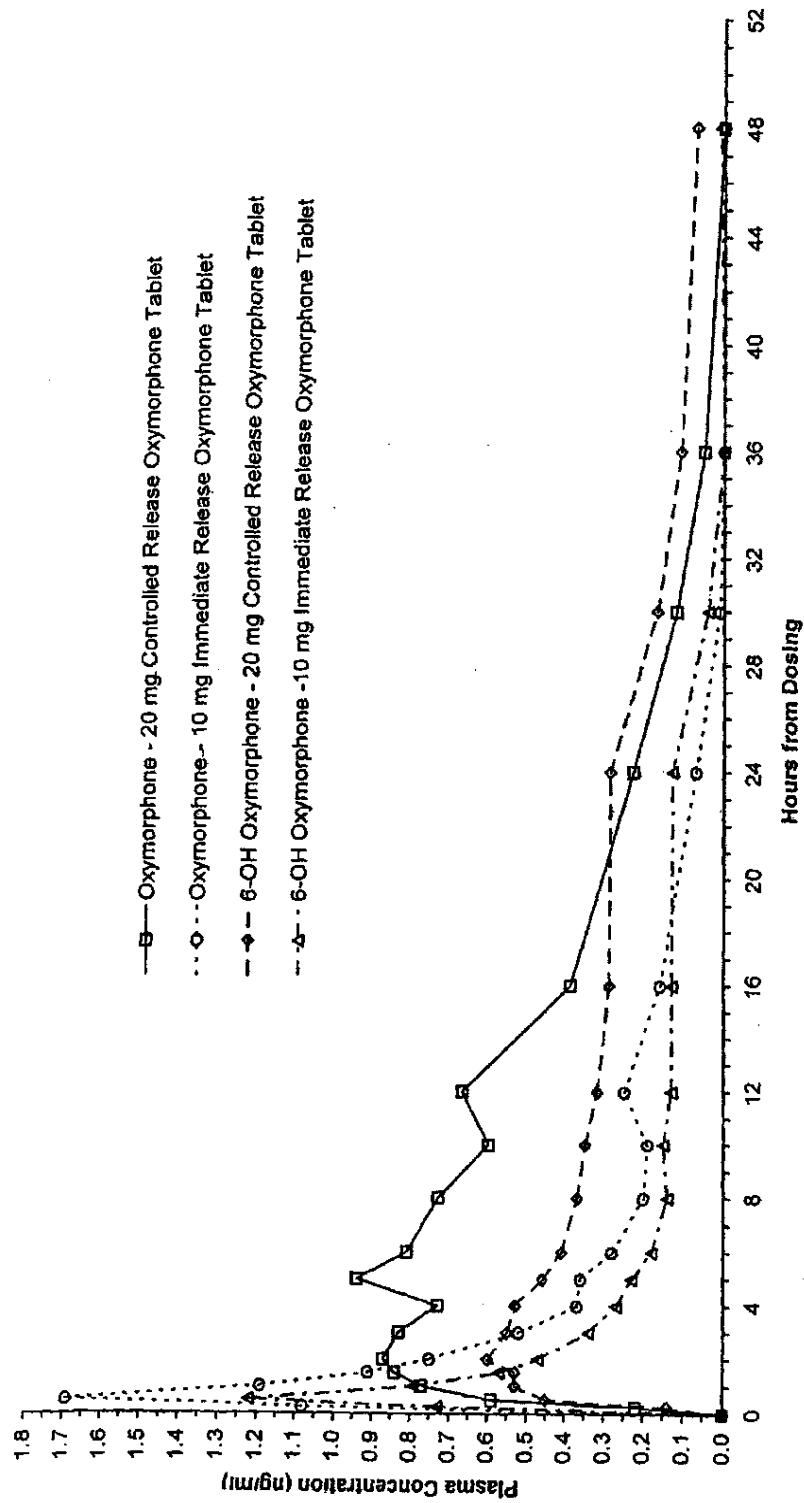


Figure 9

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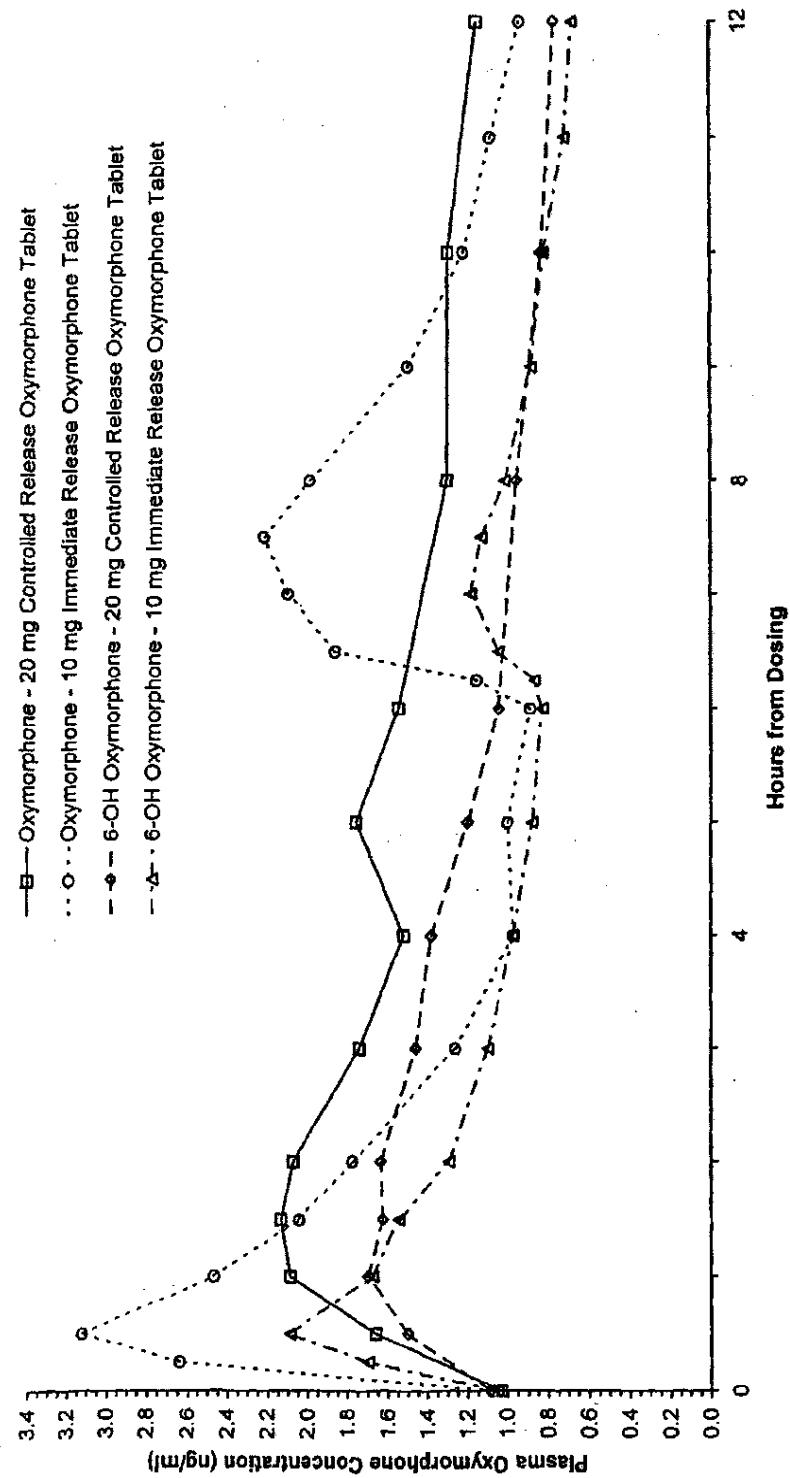


Figure 10

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OXYMORPHONE CONTROLLED RELEASE FORMULATIONS**RELATED APPLICATIONS**

This application is a continuation of U.S. patent application Ser. No. 10/190,192 filed Jul. 3, 2002 and claims priority to U.S. Provisional Patent Application Ser. Nos. 60/329,445 filed Oct. 15, 2001, 60/329,432 filed Oct. 15, 2001, 60/303, 357 filed Jul. 6, 2001, and 60/329,444 filed Oct. 15, 2001, which are incorporated herein by reference to the extent permitted by law.

BACKGROUND OF THE INVENTION

Pain is the most frequently reported symptom and it is a common clinical problem which confronts the clinician. Many millions of people in the USA suffer from severe pain that, according to numerous recent reports, is chronically undertreated or inappropriately managed. The clinical usefulness of the analgesic properties of opioids has been recognized for centuries, and morphine and its derivatives have been widely employed for analgesia for decades in a variety of clinical pain states.

Oxymorphone HCl (14-hydroxydihydromorphinone hydrochloride) is a semi-synthetic phenanthrene-derivative opioid agonist, widely used in the treatment of acute and chronic pain, with analgesic efficacy comparable to other opioid analgesics. Oxymorphone is currently marketed as an injection (1 mg/ml in 1 ml ampules; 1.5 mg/ml in 1 ml ampules; 1.5 mg/ml in 10 ml multiple dose vials) for intramuscular, subcutaneous, and intravenous administration, and as 5 mg rectal suppositories. At one time, 2 mg, 5 mg and 10 mg oral immediate release (IR) tablet formulations of oxymorphone HCl were marketed. Oxymorphone HCl is metabolized principally in the liver and undergoes conjugation with glucuronic acid and reduction to 6-alpha- and beta-hydroxy epimers.

An important goal of analgesic therapy is to achieve continuous relief of chronic pain. Regular administration of an analgesic is generally required to ensure that the next dose is given before the effects of the previous dose have worn off. Compliance with opioids increases as the required dosing frequency decreases. Non-compliance results in suboptimal pain control and poor quality of life outcomes. (Ferrell B et al. Effects of controlled-release morphine on quality of life for cancer pain. *Oncol. Nur. Forum* 1989; 4:521-26). Scheduled, rather than "as needed" administration of opioids is currently recommended in guidelines for their use in chronic non-malignant pain. Unfortunately, evidence from prior clinical trials and clinical experience suggests that the short duration of action of immediate release oxymorphone would necessitate administration every 4-6 hours in order to maintain optimal levels of analgesia in chronic pain. A controlled release formulation which would allow less frequent dosing of oxymorphone would be useful in pain management.

For instance, a controlled release formulation of morphine has been demonstrated to provide patients fewer interruptions in sleep, reduced dependence on caregivers, improved compliance, enhanced quality of life outcomes, and increased control over the management of pain. In addition, the controlled release formulation of morphine was reported to provide more constant plasma concentration and clinical effects, less frequent peak to trough fluctuations, reduced dosing frequency, and possibly fewer side effects. (Thirlwell M P et al., Pharmacokinetics and clinical efficacy of oral morphine solution and controlled-release morphine tablets in cancer

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patients. *Cancer* 1989; 63:2275-83; Goughnour B R et al., Analgesic response to single and multiple doses of controlled-release morphine tablets and morphine oral solution in cancer patients. *Cancer* 1989; 63:2294-97; Ferrell B. et al., Effects of controlled-release morphine on quality of life for cancer pain. *Oncol. Nur. Forum* 1989; 4:521-26.

There are two factors associated with the metabolism of some drugs that may present problems for their use in controlled release systems. One is the ability of the drug to induce or inhibit enzyme synthesis, which may result in a fluctuating drug blood plasma level with chronic dosing. The other is a fluctuating drug blood level due to intestinal (or other tissue) metabolism or through a hepatic first-pass effect.

Oxymorphone is metabolized principally in the liver, resulting in an oral bioavailability of about 10%. Evidence from clinical experience suggests that the short duration of action of immediate release oxymorphone necessitates a four hour dosing schedule to maintain optimal levels of analgesia. It would be useful to clinicians and patients alike to have controlled release dosage forms of oxymorphone to use to treat pain and a method of treating pain using the dosage forms.

SUMMARY OF THE INVENTION

The present invention provides methods for relieving pain by administering a controlled release pharmaceutical tablet containing oxymorphone which produces at least a predetermined minimum blood plasma level for at least 12 hours after dosing, as well as tablets that produce the sustained pain relief over this time period.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 is a pharmacokinetic profile for 6-hydroxy oxymorphone with PID scores.

FIG. 2 is a pharmacokinetic profile for oxymorphone with PID scores.

FIG. 3 is a pharmacokinetic profile for 6-hydroxy oxymorphone with categorical pain scores.

FIG. 4 is a pharmacokinetic profile for oxymorphone with categorical pain scores.

FIG. 5 is a graph of the mean blood plasma concentration of oxymorphone versus time for clinical study 1.

FIG. 6 is a graph of the mean blood plasma concentration of oxymorphone versus time for clinical study 2.

FIG. 7 is a graph of the mean blood plasma concentration of oxymorphone versus time for clinical study 3.

FIG. 8 is a graph of the mean blood plasma concentration of 6-hydroxy oxymorphone versus time for clinical study 3.

FIG. 9 is a graph of the mean blood plasma concentration of oxymorphone for immediate and controlled release tablets from a single dose study.

FIG. 10 is a graph of the mean blood plasma concentration of oxymorphone for immediate and controlled release tablets from a steady state study.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides methods for alleviating pain for 12 to 24 hours using a single dose of a pharmaceutical composition by producing a blood plasma level of oxymorphone and/or 6-OH oxymorphone of at least a minimum value for at least 12 hours or more. As used herein, the terms "6-OH oxymorphone" and "6-hydroxy oxymorphone" are interchangeable and refer to the analog of oxymorphone hav-

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ing an alcohol (hydroxy) moiety that replaces the carboxy moiety found on oxymorphone at the 6-position.

To overcome the difficulties associated with a 4-6 hourly dosing frequency of oxymorphone, this invention provides an oxymorphone controlled release oral solid dosage form, comprising a therapeutically effective amount of oxymorphone or a pharmaceutically acceptable salt of oxymorphone. It has been found that the decreased rate of release of oxymorphone from the oral controlled release formulation of this invention does not substantially decrease the bioavailability of the drug as compared to the same dose of a solution of oxymorphone administered orally. The bioavailability is sufficiently high and the release rate is such that a sufficient plasma level of oxymorphone and/or 6-OH oxymorphone is maintained to allow the controlled release dosage to be used to treat patients suffering moderate to severe pain with once or twice daily dosing. The dosing form of the present invention can also be used with thrice daily dosing.

It is critical when considering the present invention that the difference between a controlled release tablet and an immediate release formulation be fully understood. In classical terms, an immediate release formulation releases at least 80% of its active pharmaceutical ingredient within 30 minutes. With reference to the present invention, the definition of an immediate release formulation will be broadened further to include a formulation which releases more than about 80% of its active pharmaceutical ingredient within 60 minutes in a standard USP Paddle Method dissolution test at 50 rpm in 500 ml media having a pH of between 1.2 and 6.8 at 37° C. "Controlled release" formulations, as referred to herein, will then encompass any formulations which release no more than about 80% of their active pharmaceutical ingredients within 60 minutes under the same conditions.

The controlled release dosage form of this invention exhibits a dissolution rate in vitro, when measured by USP Paddle Method at 50 rpm in 500 ml media having a pH between 1.2 and 6.8 at 37° C., of about 15% to about 50% by weight oxymorphone released after 1 hour, about 45% to about 80% by weight oxymorphone released after 4 hours, and at least about 80% by weight oxymorphone released after 10 hours.

When administered orally to humans, an effective controlled release dosage form of oxymorphone should exhibit the following in vivo characteristics: (a) peak plasma level of oxymorphone occurs within about 1 to about 8 hours after administration; (b) peak plasma level of 6-OH oxymorphone occurs within about 1 to about 8 hours after administration; (c) duration of analgesic effect is through about 8 to about 24 hours after administration; (d) relative oxymorphone bioavailability is in the range of about 0.5 to about 1.5 compared to an orally-administered aqueous solution of oxymorphone; and (e) the ratio of the area under the curve of blood plasma level vs. time for 6-OH oxymorphone compared to oxymorphone is in the range of about 0.5 to about 1.5. Of course, there is variation of these parameters among subjects, depending on the size and weight of the individual subject, the subject's age, individual metabolism differences, and other factors. Indeed, the parameters may vary in an individual from day to day. Accordingly, the parameters set forth above are intended to be mean values from a sufficiently large study so as to minimize the effect of individual variation in arriving at the values. A convenient method for arriving at such values is by conducting a study in accordance with standard FDA procedures such as those employed in producing results for use in a new drug application (or abbreviated new drug application) before the FDA. Any reference to mean values herein, in conjunction with desired results, refer to results from such a study, or some comparable study. Reference to mean values

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reported herein for studies actually conducted are arrived at using standard statistical methods as would be employed by one skilled in the art of pharmaceutical formulation and testing for regulatory approval.

In one specific embodiment of the controlled release matrix form of the invention, the oxymorphone or salt of oxymorphone is dispersed in a controlled release delivery system that comprises a hydrophilic material which, upon exposure to gastrointestinal fluid, forms a gel matrix that releases oxymorphone at a controlled rate. The rate of release of oxymorphone from the matrix depends on the drug's partition coefficient between components of the matrix and the aqueous phase within the gastrointestinal tract. In a preferred form of this embodiment, the hydrophilic material of the controlled release delivery system comprises a mixture of a heteropolysaccharide gum and an agent capable of cross-linking the heteropolysaccharide in presence of gastrointestinal fluid. The controlled release delivery system may also comprise a water-soluble pharmaceutical diluent mixed with the hydrophilic material. Preferably, the cross-linking agent is a homopolysaccharide gum and the inert pharmaceutical diluent is a monosaccharide, a disaccharide, or a polyhydric alcohol, or a mixture thereof.

In a specific preferred embodiment, the appropriate blood plasma levels of oxymorphone and 6-hydroxy oxymorphone are achieved using oxymorphone in the form of oxymorphone hydrochloride, wherein the weight ratio of heteropolysaccharide to homopolysaccharide is in the range of about 1:3 to about 3:1, the weight ratio of heteropolysaccharide to diluent is in the range of about 1:8 to about 8:1, and the weight ratio of heteropolysaccharide to oxymorphone hydrochloride is in the range of about 10:1 to about 1:10. A preferred heteropolysaccharide is xanthan gum and a preferred homopolysaccharide is locust bean gum. The dosage form also comprises a cationic cross-linking agent and a hydrophobic polymer. In the preferred embodiment, the dosage form is a tablet containing about 5 mg to about 80 mg of oxymorphone hydrochloride. In a most preferred embodiment, the tablet contains about 20 mg oxymorphone hydrochloride.

The invention includes a method which comprises achieving appropriate blood plasma levels of drug while providing extended pain relief by administering one to three times per day to a patient suffering moderate to severe, acute or chronic pain, an oxymorphone controlled release oral solid dosage form of the invention in an amount sufficient to alleviate the pain for a period of about 8 hours to about 24 hours. This type and intensity of pain is often associated with cancer, autoimmune diseases, infections, surgical and accidental traumas and osteoarthritis.

The invention also includes a method of making an oxymorphone controlled release oral solid dosage form of the invention which comprises mixing particles of oxymorphone or a pharmaceutically acceptable salt of oxymorphone with granules comprising the controlled release delivery system, preferably followed by directly compressing the mixture to form tablets.

Pharmaceutically acceptable salts of oxymorphone which can be used in this invention include salts with the inorganic and organic acids which are commonly used to produce non-toxic salts of medicinal agents. Illustrative examples would be those salts formed by mixing oxymorphone with hydrochloric, sulfuric, nitric, phosphoric, phosphorous, hydrobromic, malic, malic, ascorbic, citric or tartaric, pamoic, lauric, stearic, palmitic, oleic, myristic, lauryl sulfuric, naphthalene sulfonic, linoleic or linolenic acid, and the like. The hydrochloride salt is preferred.

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It has now been found that 6-OH oxymorphone, which is one of the metabolites of oxymorphone, may play a role in alleviating pain. When oxymorphone is ingested, part of the dosage gets into the bloodstream to provide pain relief, while another part is metabolized to 6-OH oxymorphone. This metabolite then enters the bloodstream to provide further pain relief. Thus it is believed that both the oxymorphone and 6-hydroxyoxymorphone levels are important to pain relief.

The effectiveness of oxymorphone and 6-hydroxyoxymorphone at relieving pain and the pharmacokinetics of a single dose of oxymorphone were studied. The blood plasma levels of both oxymorphone and 6-hydroxyoxymorphone were measured in patients after a single dose of oxymorphone was administered. Similarly, the pain levels in patients were measured after a single administration of oxymorphone to determine the effective duration of pain relief from a single dose. FIGS. 1-2 show the results of these tests, comparing pain levels to oxymorphone and 6-hydroxy oxymorphone levels.

For these tests, pain was measured using a Visual Analog Scale (VAS) or a Categorical Scale. The VAS scales consisted of a horizontal line, 100 mm in length. The left-hand end of the scale (0 mm) was marked with the descriptor "No Pain" and the right-hand end of the scale (100 mm) was marked with the descriptor "Extreme Pain". Patients indicated their level of pain by making a vertical mark on the line. The VAS score was equal to the distance (in mm) from the left-hand end of the scale to the patient's mark. For the categorical scale, patients completed the following statement, "My pain at this time is" using the scale None=0, Mild=1, Moderate=2, or Severe=3.

As can be seen from these figures, there is a correlation between pain relief and both oxymorphone and 6-hydroxyoxymorphone levels. As the blood plasma levels of oxymorphone and 6-hydroxyoxymorphone increase, pain decreases (and pain intensity difference and pain relief increases). Thus, to the patient, it is the level of oxymorphone and 6-hydroxyoxymorphone in the blood plasma which is most important. Further it is these levels which dictate the efficacy of the dosage form. A dosage form which maintains a sufficiently high level of oxymorphone or 6-hydroxyoxymorphone for a longer period need not be administered frequently. Such a result is accomplished by embodiments of the present invention.

The oxymorphone controlled release oral solid dosage form of this invention can be made using any of several different techniques for producing controlled release oral solid dosage forms of opioid analgesics.

In one embodiment, a core comprising oxymorphone or oxymorphone salt is coated with a controlled release film which comprises a water insoluble material and which upon exposure to gastrointestinal fluid releases oxymorphone from the core at a controlled rate. In a second embodiment, the oxymorphone or oxymorphone salt is dispersed in a controlled release delivery system that comprises a hydrophilic material which upon exposure to gastrointestinal fluid forms a gel matrix that releases oxymorphone at a controlled rate. A third embodiment is a combination of the first two: a controlled release matrix coated with a controlled release film. In a fourth embodiment the oxymorphone is incorporated into an osmotic pump. In any of these embodiments, the dosage form can be a tablet, a plurality of granules in a capsule, or other suitable form, and can contain lubricants, colorants, diluents, and other conventional ingredients.

Osmotic Pump

An osmotic pump comprises a shell defining an interior compartment and having an outlet passing through the shell. The interior compartment contains the active pharmaceutical

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ingredient. Generally the active pharmaceutical ingredient is mixed with excipients or other compositions such as a polyalkylene. The shell is generally made, at least in part, from a material (such as cellulose acetate) permeable to the liquid of the environment where the pump will be used, usually stomach acid. Once ingested, the pump operates when liquid diffuses through the shell of the pump. The liquid dissolves the composition to produce a saturated situation. As more liquid diffuses into the pump, the saturated solution containing the pharmaceutical is expelled from the pump through the outlet. This produces a nearly constant release of active ingredient, in the present case, oxymorphone.

Controlled Release Coating

In this embodiment, a core comprising oxymorphone or oxymorphone salt is coated with a controlled release film which comprises a water insoluble material. The film can be applied by spraying an aqueous dispersion of the water insoluble materials include alkyl celluloses, acrylic polymers, waxes (alone or in admixture with fatty alcohols), shellac and zein. The aqueous dispersions of alkyl celluloses and acrylic polymers preferably contain a plasticizer such as triethyl citrate, dibutyl phthalate, propylene glycol, and polyethylene glycol. The film coat can contain a water-soluble material such as polyvinylpyrrolidone (PVP) or hydroxypropylmethylcellulose (HPMC).

The core can be a granule made, for example, by wet granulation of mixed powders of oxymorphone or oxymorphone salt and a binding agent such as HPMC, or by coating an inert bead with oxymorphone or oxymorphone salt and a binding agent such as HPMC, or by spheronising mixed powders of oxymorphone or oxymorphone salt and a spheronising agent such as microcrystalline cellulose. The core can be a tablet made by compressing such granules or by compressing a powder comprising oxymorphone or oxymorphone salt.

The in vitro and in vivo release characteristics of this controlled release dosage form can be modified by using mixtures of different water insoluble and water soluble materials, using different plasticizers, varying the thickness of the controlled release film, including release-modifying agents in the coating, or by providing passageways through the coating.

Controlled Release Matrix

It is important in the present invention that appropriate blood plasma levels of oxymorphone and 6-hydroxy oxymorphone be achieved and maintained for sufficient time to provide pain relief to a patient for a period of 12 to 24 hours. The preferred composition for achieving and maintaining the proper blood plasma levels is a controlled-release matrix. In this embodiment, the oxymorphone or oxymorphone salt is dispersed in a controlled release delivery system that comprises a hydrophilic material (gelling agent) which upon exposure to gastrointestinal fluid forms a gel matrix that releases oxymorphone at a controlled rate. Such hydrophilic materials include gums, cellulose ethers, acrylic resins, and protein-derived materials. Suitable cellulose ethers include hydroxylalkyl celluloses and carboxyalkyl celluloses, especially hydroxyethyl cellulose (HEC), hydroxypropyl cellulose (HPC), HPMC, and carboxy methylcellulose (CMC). Suitable acrylic resins include polymers and copolymers of acrylic acid, methacrylic acid, methyl acrylate and methyl methacrylate. Suitable gums include heteropolysaccharide and homopolysaccharide gums, e.g., xanthan, tragacanth, acacia, karaya, alginates, agar, guar, hydroxypropyl guar, carageenan, and locust bean gums.

Preferably, the controlled release tablet of the present invention is formed from (I) a hydrophilic material comprising (a) a heteropolysaccharide; or (b) a heteropolysaccharide

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and a cross-linking agent capable of cross-linking said heteropolysaccharide; or (c) a mixture of (a), (b) and a polysaccharide gum; and (II) an inert pharmaceutical filler comprising up to about 80% by weight of the tablet; and (III) oxymorphone.

The term "heteropolysaccharide" as used herein is defined as a water-soluble polysaccharide containing two or more kinds of sugar units, the heteropolysaccharide having a branched or helical configuration, and having excellent water-wicking properties and immense thickening properties.

A preferred heteropolysaccharide is xanthan gum, which is a high molecular weight ($>10^6$) heteropolysaccharide. Other preferred heteropolysaccharides include derivatives of xanthan gum, such as deacylated xanthan gum, the carboxymethyl ether, and the propylene glycol ester.

The cross linking agents used in the controlled release embodiment of the present invention which are capable of cross-linking with the heteropolysaccharide include homopolysaccharide gums such as the galactomannans, i.e., polysaccharides which are composed solely of mannose and galactose. Galactomannans which have higher proportions of unsubstituted mannose regions have been found to achieve more interaction with the heteropolysaccharide. Locust bean gum, which has a higher ratio of mannose to the galactose, is especially preferred as compared to other galactomannans such as guar and hydroxypropyl guar.

Preferably, the ratio of heteropolysaccharide to homopolysaccharide is in the range of about 1:9 to about 9:1, preferably about 1:3 to about 3:1. Most preferably, the ratio of xanthan gum to polysaccharide material (i.e., locust bean gum, etc.) is preferably about 1:1.

In addition to the hydrophilic material, the controlled release delivery system can also contain an inert pharmaceutical diluent such as a monosaccharide, a disaccharide, a polyhydric alcohol and mixtures thereof. The ratio of diluent to hydrophilic matrix-forming material is generally in the range of about 1:3 to about 3:1.

The controlled release properties of the controlled release embodiment of the present invention may be optimized when the ratio of heteropolysaccharide gum to homopolysaccharide material is about 1:1, although heteropolysaccharide gum in an amount of from about 20 to about 80% or more by weight of the heterodisperse polysaccharide material provides an acceptable slow release product. The combination of any homopolysaccharide gums known to produce a synergistic effect when exposed to aqueous solutions may be used in accordance with the present invention. It is also possible that the type of synergism which is present with regard to the gum combination of the present invention could also occur between two homogeneous or two heteropolysaccharides. Other acceptable gelling agents which may be used in the present invention include those gelling agents well-known in the art. Examples include vegetable gums such as alginates, carrageenan, pectin, guar gum, xanthan gum, modified starch, hydroxypropylmethylcellulose, methylcellulose, and other cellulosic materials such as sodium carboxymethylcellulose and hydroxypropyl cellulose. This list is not meant to be exclusive.

The combination of xanthan gum with locust bean gum with or without the other homopolysaccharide gums is an especially preferred gelling agent. The chemistry of certain of the ingredients comprising the excipients of the present invention such as xanthan gum is such that the excipients are considered to be self-buffering agents which are substantially

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insensitive to the solubility of the medicament and likewise insensitive to the pH changes along the length of the gastrointestinal tract.

The inert filler of the sustained release excipient preferably comprises a pharmaceutically acceptable saccharide, including a monosaccharide, a disaccharide, or a polyhydric alcohol, and/or mixtures of any of the foregoing. Examples of suitable inert pharmaceutical fillers include sucrose, dextrose, lactose, microcrystalline cellulose, fructose, xylitol, sorbitol, mixtures thereof and the like. However, it is preferred that a soluble pharmaceutical filler such as lactose, dextrose, sucrose, or mixtures thereof be used.

The cationic cross-linking agent which is optionally used in conjunction with the controlled release embodiment of the present invention may be monovalent or multivalent metal cations. The preferred salts are the inorganic salts, including various alkali metal and/or alkaline earth metal sulfates, chlorides, borates, bromides, citrates, acetates, lactates, etc. Specific examples of suitable cationic cross-linking agents include calcium sulfate, sodium chloride, potassium sulfate, sodium carbonate, lithium chloride, tripotassium phosphate, sodium borate, potassium bromide, potassium fluoride, sodium bicarbonate, calcium chloride, magnesium chloride, sodium citrate, sodium acetate, calcium lactate, magnesium sulfate and sodium fluoride. Multivalent metal cations may also be utilized. However, the preferred cationic cross-linking agents are bivalent. Particularly preferred salts are calcium sulfate and sodium chloride. The cationic cross-linking agents of the present invention are added in an amount effective to obtain a desirable increased gel strength due to the cross-linking of the gelling agent (e.g., the heteropolysaccharide and homopolysaccharide gums). In preferred embodiments, the cationic cross-linking agent is included in the sustained release excipient of the present invention in an amount from about 1 to about 20% by weight of the sustained release excipient, and in an amount about 0.5% to about 16% by weight of the final dosage form.

In the controlled release embodiments of the present invention, the sustained release excipient comprises from about 10 to about 99% by weight of a gelling agent comprising a heteropolysaccharide gum and a homopolysaccharide gum, from about 1 to about 20% by weight of a cationic crosslinking agent, and from about 0 to about 89% by weight of an inert pharmaceutical diluent. In other embodiments, the sustained release excipient comprises from about 10 to about 75% gelling agent, from about 2 to about 15% cationic crosslinking agent, and from about 30 to about 75% inert diluent. In yet other embodiments, the sustained release excipient comprises from about 30 to about 75% gelling agent, from about 5 to about 10% cationic cross-linking agent, and from about 15 to about 65% inert diluent.

The sustained release excipient used in this embodiment of the present invention (with or without the optional cationic cross-linking agent) may be further modified by incorporation of a hydrophobic material which slows the hydration of the gums without disrupting the hydrophilic matrix. This is accomplished in preferred embodiments of the present invention by granulating the sustained release excipient with the solution or dispersion of a hydrophobic material prior to the incorporation of the medicament. The hydrophobic polymer may be selected from an alkylcellulose such as ethylcellulose, other hydrophobic cellulosic materials, polymers or copolymers derived from acrylic or methacrylic acid esters, copolymers of acrylic and methacrylic acid esters, zein, waxes, shellac, hydrogenated vegetable oils, and any other pharmaceutically acceptable hydrophobic material known to those skilled in the art. The amount of hydrophobic material incor-

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porated into the sustained release excipient is that which is effective to slow the hydration of the gums without disrupting the hydrophilic matrix formed upon exposure to an environmental fluid. In certain preferred embodiments of the present invention, the hydrophobic material is included in the sustained release excipient in an amount from about 1 to about 20% by weight. The solvent for the hydrophobic material may be an aqueous or organic solvent, or mixtures thereof.

Examples of commercially available alkylcelluloses are Aquacoat coating (aqueous dispersion of ethylcellulose available from FMC of Philadelphia, Pa.) and Surelease coating (aqueous dispersion of ethylcellulose available from Colordon of West Point, Pa.). Examples of commercially available acrylic polymers suitable for use as the hydrophobic material include Eudragit RS and RL polymers (copolymers of acrylic and methacrylic acid esters having a low content (e.g., 1:20 or 1:40) of quaternary ammonium compounds available from Rohm America of Piscataway, N.J.).

The controlled release matrix useful in the present invention may also contain a cationic cross-linking agent such as calcium sulfate in an amount sufficient to cross-link the gelling agent and increase the gel strength, and an inert hydrophobic material such as ethyl cellulose in an amount sufficient to slow the hydration of the hydrophilic material without disrupting it. Preferably, the controlled release delivery system is prepared as a pre-manufactured granulation.

EXAMPLES

Example 1

Two controlled release delivery systems are prepared by dry blending xanthan gum, locust bean gum, calcium sulfate dehydrate, and dextrose in a high speed mixed/granulator for 3 minutes. A slurry is prepared by mixing ethyl cellulose with alcohol. While running choppers/impellers, the slurry is added to the dry blended mixture, and granulated for another 3 minutes. The granulation is then dried to a LOD (loss on drying) of less than about 10% by weight. The granulation is then milled using 20 mesh screen. The relative quantities of the ingredients are listed in the table below.

TABLE 1

Excipient	Controlled Release Delivery System	
	Formulation 1 (%)	Formulation 2 (%)
Locust Bean Gum, FCC	25.0	30.0
Xanthan Gum, NF	25.0	30.0
Dextrose, USP	35.0	40.0
Calcium Sulfate Dihydrate, NF	10.0	0.0
Ethylcellulose, NF	5.0	0.0
Alcohol, SD3A (Anhydrous)	(10) ¹	(20.0) ¹
Total	100.0	100.0

A series of tablets containing different amounts of oxymorphone hydrochloride were prepared using the controlled release delivery Formulation 1 shown in Table 1. The quantities of ingredients per tablet are as listed in the following table.

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TABLE 2

Component	Sample Tablets of Differing Strengths				
	Amounts in Tablet (mg)				
Oxymorphone HCl, USP (mg)	5	10	20	40	80
Controlled release delivery system	160	160	160	160	160
Silicified microcrystalline cellulose, NF	20	20	20	20	20
Sodium stearyl fumarate, NF	2	2	2	2	2
Total weight	187	192	202	222	262
Opadry (colored)	7.48	7.68	8.08	8.88	10.48
Opadry (clear)	0.94	0.96	1.01	1.11	1.31

Examples 2 and 3

20 Two batches of 20 mg tablets were prepared as described above, using the controlled release delivery system of Formulation 1. One batch was formulated to provide relatively fast controlled release, the other batch was formulated to provide relatively slow controlled release. Compositions of the tablets are shown in the following table.

TABLE 3

Ingredients	Slow and Fast Release Compositions		
	Example 2 Slow (mg)	Example 3 Fast (mg)	Example 4 Fast (mg)
Oxymorphone HCl, USP	20	20	20
Controlled Release Delivery System	360	160	160
Silicified Microcrystalline Cellulose, NF	20	20	20
Sodium stearyl fumarate, NF	4	2	2
Total weight	404	202	202
Coating (color or clear)	12	12	9

30 The tablets of Examples 2, 3, and 4 were tested for in vitro release rate according to USP Procedure Drug Release U.S. Pat. No. 23. Release rate is a critical variable in attempting to control the blood plasma levels of oxymorphone and 6-hydroxyoxymorphone in a patient. Results are shown in the following Table 4.

TABLE 4

Time (hr)	Release Rates of Slow and Fast Release Tablets		
	Example 2 (Slow Release)	Example 3 (Fast Release)	Example 4 (Fast Release)
0.5	18.8	21.3	20.1
1	27.8	32.3	31.7
2	40.5	47.4	46.9
3	50.2	58.5	57.9
4	58.1	66.9	66.3
5	64.7	73.5	74.0
6	70.2	78.6	83.1
8	79.0	86.0	92.0
10	85.3	90.6	95.8
12	89.8	93.4	97.3

Clinical Studies

50 65 Three clinical studies were conducted to assess the bioavailability (rate and extent of absorption) of oxymorphone. Study 1 addressed the relative rates of absorption of con-

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trolled release (CR) oxymorphone tablets (of Examples 2 and 3) and oral oxymorphone solution in fasted patients. Study 2 addressed the relative rates of absorption of CR oxymorphone tablets (of Examples 2 and 3) and oral oxymorphone solution in fed patients. Study 3 addressed the relative rates of absorption of CR oxymorphone tablets (of Example 4) and oral oxymorphone solution in fed and fasted patients.

The blood plasma levels set forth herein as appropriate to achieve the objects of the present invention are mean blood plasma levels. As an example, if the blood plasma level of oxymorphone in a patient 12 hours after administration of a tablet is said to be at least 0.5 ng/ml, any particular individual may have lower blood plasma levels after 12 hours. However, the mean minimum concentration should meet the limitation set forth. To determine mean parameters, a study should be performed with a minimum of 8 adult subjects, in a manner acceptable for filing an application for drug approval with the US Food and Drug Administration. In cases where large fluctuations are found among patients, further testing may be necessary to accurately determine mean values.

For all studies, the following procedures were followed, unless otherwise specified for a particular study.

The subjects were not to consume any alcohol-, caffeine-, or xanthine-containing foods or beverages for 24 hours prior to receiving study medication for each study period. Subjects were to be nicotine and tobacco free for at least 6 months prior to enrolling in the study. In addition, over-the-counter medications were prohibited 7 days prior to dosing and during the study. Prescription medications were not allowed 14 days prior to dosing and during the study.

Pharmacokinetic and Statistical Methods

The following pharmacokinetic parameters were computed from the plasma oxymorphone concentration-time data:

$AUC_{(0-t)}$ Area under the drug concentration-time curve from time zero to the time of the last quantifiable concentration (C_t), calculated using linear trapezoidal summation.

$AUC_{(0-\infty)}$ Area under the drug concentration-time curve from time zero to infinity. $AUC_{(0-\infty)} = AUC_{(0-t)} + C_t/K_{el}$, where K_{el} is the terminal elimination rate constant.

$AUC_{(0-24)}$ Partial area under the drug concentration-time curve from time zero to 24 hours.

C_{max} Maximum observed drug concentration.

T_{max} Time of the observed maximum drug concentration.

K_{el} Elimination rate constant based on the linear regression of the terminal linear portion of the LN (concentration) time curve.

Terminal elimination rate constants for use in the above calculations were in turn computed using linear regression of a minimum of three time points, at least two of which were consecutive. K_{el} values for which correlation coefficients were less than or equal to 0.8 were not reported in the pharmacokinetic parameter tables or included in the statistical analysis. Thus $AUC_{(0-\infty)}$ was also not reported in these cases.

A parametric (normal-theory) general linear model was applied to each of the above parameters (excluding T_{max}), and the LN-transformed parameters C_{max} , $AUC_{(0-24)}$, $AUC_{(0-t)}$, and $AUC_{(0-\infty)}$. Initially, the analysis of variance (ANOVA) model included the following factors: treatment, sequence, subject within sequence, period, and carryover effect. If carryover effect was not significant, it was dropped from the model. The sequence effect was tested using the subject within sequence mean square, and all other main effects were tested using the residual error (error mean square).

Plasma oxymorphone concentrations were listed by subject at each collection time and summarized using descriptive

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statistics. Pharmacokinetic parameters were also listed by subject and summarized using descriptive statistics.

Study 1-Two Controlled Release Formulations; Fasted Patients

Healthy volunteers received a single oral dose of 20 mg CR oxymorphone taken with 240 ml water after a 10-hour fast. Subjects received the tablets of Example 2 (Treatment 1A) or Example 3 (Treatment 1B). Further subjects were given a single oral dose of 10 mg/10 ml oxymorphone solution in 180 ml apple juice followed with 60 ml water (Treatment 1C). The orally dosed solution was used to simulate an immediate release (IR) dose.

This study had a single-center, open-label, randomized, three-way crossover design using fifteen subjects. Subjects were in a fasted state following a 10-hour overnight fast. There was a 14-day washout interval between the three dose administrations. The subjects were confined to the clinic during each study period. Subjects receiving Treatment 1C were confined for 18 hours and subjects receiving Treatments 1A or 1B were confined for 48 hours after dosing. Ten-milliliter blood samples were collected during each study period at the 0 hour (predose), and at 0.5, 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 10, 12, 14, 16, 18, 20, 24, 28, 32, 36, and 48 hours postdose for subjects receiving Treatment 1A or 1B and 0, 0.25, 0.5, 0.75, 1, 1.25, 1.5, 1.75, 2, 2.5, 3, 4, 5, 6, 7, 8, 10, 12, 14, 16, and 18 hours post-dose. The mean plasma concentration of oxymorphone versus time for each treatment across all subjects is shown in table 5.

TABLE 5

Mean Plasma Concentration vs. Time (ng/ml)			
Time (hr)	Treatment 1A	Treatment 1B	Treatment 1C
35 0	0.000	0.000	0.0000
0.25			0.9489
0.5	0.2941	0.4104	1.3016
0.75			1.3264
1	0.5016	0.7334	1.3046
1.25			1.2041
1.5	0.5951	0.8192	1.0813
1.75			0.9502
2	0.6328	0.7689	0.9055
2.5			0.7161
3	0.5743	0.7341	0.6689
4	0.5709	0.6647	0.4879
5	0.7656	0.9089	0.4184
6	0.7149	0.7782	0.3658
7	0.6334	0.6748	0.3464
8	0.5716	0.5890	0.2610
10	0.4834	0.5144	0.2028
12	0.7333	0.6801	0.2936
14	0.6271	0.6089	0.2083
16	0.4986	0.4567	0.1661
18	0.4008	0.3674	0.1368
20	0.3405	0.2970	
24	0.2736	0.2270	
28	0.3209	0.2805	
32	0.2846	0.2272	
36	0.2583	0.1903	
48	0.0975	0.0792	

The results are shown graphically in FIG. 5. In both Table 5 and FIG. 5, the results are normalized to a 20 mg dosage. The immediate release liquid of Treatment 1C shows a classical curve, with a high and relatively narrow peak, followed by an exponential drop in plasma concentration. However, the controlled release oxymorphone tablets exhibit triple peaks in blood plasma concentration. The first peak occurs (on average) at around 3 hours. The second peak of the mean blood plasma concentration is higher than the first, occurring around 6-7 hours, on average).

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Occasionally, in an individual, the first peak is higher than the second, although generally this is not the case. This makes it difficult to determine the time to maximum blood plasma concentration (T_{max}) because if the first peak is higher than the second, maximum blood plasma concentration (C_{max}) occurs much earlier (at around 3 hours) than in the usual case where the second peak is highest. Therefore, when we refer to the time to peak plasma concentration (T_{max}) unless otherwise specified, we refer to the time to the second peak. Further, when reference is made to the second peak, we refer to the time or blood plasma concentration at the point where the blood plasma concentration begins to drop the second time. Generally, where the first peak is higher than the second, the difference in the maximum blood plasma concentration at the two peaks is small. Therefore, this difference (if any) was ignored and the reported C_{max} was the true maximum blood plasma concentration and not the concentration at the second peak.

TABLE 6

Pharmacokinetic Parameters of Plasma Oxymorphone for Study 1						
	Treatment 1A		Treatment 1B		Treatment 1C	
	Mean	SD	Mean	SD	Mean	SD
C_{max}	0.8956	0.2983	1.0362	0.3080	2.9622	1.0999
T_{max}	7.03	4.10	4.89	3.44	0.928	0.398
$AUC_{(0-t)}$	17.87	6.140	17.16	6.395	14.24	5.003
$AUC_{(0-\infty)}$	19.87	6.382	18.96	6.908	16.99	5.830
$T_{1/2el}$	10.9	2.68	11.4	2.88	6.96	4.61

Units:
 C_{max} in ng/ml;
 T_{max} in hours;
 AUC in ng * hr/ml;
 $T_{1/2el}$ in hours.

Relative bioavailability determinations are set forth in Tables 7 and 8. For these calculations, AUC was normalized for all treatments to a 20 mg dose.

TABLE 7

Relative Bioavailability (F_{rel}) Determination Based on $AUC_{(0-\infty)}$		
F_{rel} (1A vs. 1C)	F_{rel} (1B vs. 1C)	F_{rel} (1A vs. 1B)
1.193 ± 0.203	1.121 ± 0.211	1.108 ± 0.152

TABLE 8

Relative Bioavailability Determination Based on $AUC_{(0-t)}$		
F_{rel} (1A vs. 1C)	F_{rel} (1B vs. 1C)	F_{rel} (1A vs. 1B)
0.733 ± 0.098	0.783 ± 0.117	0.944 ± 0.110

Study 2-Two CR Formulations; Fed Patients

Healthy volunteers received a single oral dose of 20 mg CR oxymorphone taken with 240 ml water in a fed state. Subjects received the tablets of Example 2 (Treatment 2A) or Example 3 (Treatment 2B). Further subjects were given a single oral dose of 10 mg/10 ml oxymorphone solution in 180 ml apple juice followed with 60 ml water (Treatment 2C). The orally dosed solution was used to simulate an immediate release (IR) dose.

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This study had a single-center, open-label, randomized, three-way crossover design using fifteen subjects. The subjects were in a fed state, after a 10-hour overnight fast followed by a standardized FDA high-fat breakfast. There was a 14-day washout interval between the three dose administrations. The subjects were confined to the clinic during each study period. Subjects receiving Treatment 2C were confined for 18 hours and subjects receiving Treatments 2A or 2B were confined for 48 hours after dosing. Ten-milliliter blood samples were collected during each study period at the 0 hour (predose), and at 0.5, 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 10, 12, 14, 16, 18, 20, 24, 28, 32, 36, and 48 hours postdose for subjects receiving Treatment 2A or 2B and 0, 0.25, 0.5, 0.75, 1, 1.25, 1.5, 1.75, 2, 2.5, 3, 4, 5, 6, 7, 8, 10, 12, 14, 16, and 18 hours postdose. The mean plasma concentration of oxymorphone versus time for each treatment across all subjects is shown in table 9.

TABLE 9

Mean Plasma Concentration vs. Time (ng/ml)			
Time (hr)	Treatment 2A	Treatment 2B	Treatment 2C
0	0.000	0.000	0.0000
0.25			1.263
0.5	0.396	.0553	1.556
0.75			1.972
1	0.800	1.063	1.796
1.25			1.795
1.5	1.038	1.319	1.637
1.75			1.467
2	1.269	1.414	1.454
2.5			1.331
3	1.328	1.540	1.320
4	1.132	1.378	1.011
5	1.291	1.609	0.731
6	1.033	1.242	0.518
7	0.941	0.955	0.442
8	0.936	0.817	0.372
10	0.669	0.555	0.323
12	0.766	0.592	0.398
14	0.641	0.519	0.284
16	0.547	0.407	0.223
18	0.453	0.320	0.173
20	0.382	0.280	
24	0.315	0.254	
28	0.352	0.319	
32	0.304	0.237	
36	0.252	0.207	
48	0.104	0.077	

The results are shown graphically in FIG. 6. Again, the results have been normalized to a 20 mg dosage. As with Study 1, the immediate release liquid of Treatment 2C shows a classical curve, with a high and relatively narrow peak, followed by an exponential drop in plasma concentration, while the controlled release oxymorphone tablets exhibit triple peaks in blood plasma concentration. Thus, again when we refer to the time to peak plasma concentration (T_{max}) unless otherwise specified, we refer to the time to the second peak.

TABLE 10

Pharmacokinetic Parameters of Plasma Oxymorphone for Study 2						
	Treatment 2A		Treatment 2B		Treatment 2C	
	Mean	SD	Mean	SD	Mean	SD
C_{max}	1.644	0.365	1.944	0.465	4.134	0.897
T_{max}	3.07	1.58	2.93	1.64	0.947	0.313
$AUC_{(0-t)}$	22.89	5.486	21.34	5.528	21.93	5.044

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TABLE 10-continued

Pharmacokinetic Parameters of Plasma Oxymorphone for Study 2						
Treatment 2A		Treatment 2B	Treatment 2C			
Mean	SD	Mean	SD	Mean	SD	
AUC _(0-∞)	25.28	5.736	23.62	5.202	24.73	6.616
T _{1/2el}	12.8	3.87	11.0	3.51	5.01	2.02

Units:

 C_{max} in ng/ml, T_{max} in hours,

AUC in ng * hr/ml,

T_{1/2el} in hours.

In Table 10, the T_{max} has a large standard deviation due to the two comparable peaks in blood plasma concentration. Relative bioavailability determinations are set forth in Tables 11 and 12.

TABLE 11

Relative Bioavailability Determination Based on AUC _(0-∞)		
F _{rel} (2A vs. 2C)	F _{rel} (2B vs. 2C)	F _{rel} (2A vs. 2B)
1.052 ± 0.187	0.949 ± 0.154	1.148 ± 0.250

TABLE 12

Relative bioavailability Determination Based on AUC _(0-∞)		
F _{rel} (2A vs. 2C)	F _{rel} (2B vs. 2C)	F _{rel} (2A vs. 2B)
0.690 ± 0.105	0.694 ± 0.124	1.012 ± 0.175

As may be seen from tables 5 and 10 and FIGS. 1 and 2, the C_{max} for the CR tablets (treatments 1A, 1B, 2A and 2B) is considerably lower, and the T_{max} much higher than for the immediate release oxymorphone. The blood plasma level of oxymorphone remains high well past the 8 (or even the 12) hour dosing interval desired for an effective controlled release tablet.

Study 3-One Controlled Release Formulation; Fed and Fasted Patients

This study had a single-center, open-label, analytically blinded, randomized, four-way crossover design. Subjects randomized to Treatment 3A and Treatment 3C, as described below, were in a fasted state following a 10-hour overnight fast. Subjects randomized to Treatment 3B and Treatment 3D, as described below, were in the fed state, having had a high fat meal, completed ten minutes prior to dosing. There was a 14-day washout interval between the four dose administrations. The subjects were confined to the clinic during each study period. Subjects assigned to receive Treatment 3A and Treatment 3B were discharged from the clinic on Day 3 following the 48-hour procedures, and subjects assigned to receive Treatment 3C and Treatment 3D were discharged from the clinic on Day 2 following the 36-hour procedures. On Day 1 of each study period the subjects received one of four treatments:

Treatments 3A and 3B: Oxymorphone controlled release 60 mg tablets from Example 3. Subjects randomized to Treatment 3A received a single oral dose of one 20 mg oxymorphone controlled release tablet taken with 240 ml of water after a 10-hour fasting period. Subjects randomized to Treatment 3B received a single oral dose of one 20 mg oxymorphone controlled release tablet taken with 240 ml of water 10 minutes after a standardized high fat meal.

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Treatments 3C and 3D: oxymorphone HCl solution, USP, 1.5 mg/ml 10 ml vials. Subjects randomized to Treatment 3C received a single oral dose of 10 mg (6.7 ml) oxymorphone solution taken with 240 ml of water after a 10-hour fasting period. Subjects randomized to Treatment 3D received a single oral dose of 10 mg (6.7 ml) oxymorphone solution taken with 240 ml of water 10 minutes after a standardized high-fat meal.

A total of 28 male subjects were enrolled in the study, and 24 subjects completed the study. The mean age of the subjects was 27 years (range of 19 through 38 years), the mean height of the subjects was 69.6 inches (range of 64.0 through 75.0 inches), and the mean weight of the subjects was 169.0 pounds (range 117.0 through 202.0 pounds).

A total of 28 subjects received at least one treatment. Only subjects who completed all 4 treatments were included in the summary statistics and statistical analysis.

Blood samples (7 ml) were collected during each study period at the 0 hour (predose), and at 0.5, 1, 1.5, 2, 3, 4, 5, 6, 8, 10, 12, 14, 16, 20, 24, 30, 36, and 48 hours post-dose (19 samples) for subjects randomized to Treatment 3A and Treatment 3B. Blood samples (7 ml) were collected during each study period at the 0 hour (predose), and at 0.25, 0.5, 0.75, 1, 1.25, 1.5, 1.75, 2, 3, 4, 5, 6, 8, 10, 12, 14, 16, 20, and 36 hours post-dose (21 samples) for subjects randomized to Treatment 3C and Treatment 3D.

The mean oxymorphone plasma concentration versus time curves for Treatments 3A, 3B, 3C, and 3D are presented in FIG. 7. The results have been normalized to a 20 mg dosage. The data is contained in Table 13. The arithmetic means of the plasma oxymorphone pharmacokinetic parameters and the statistics for all Treatments are summarized in Table 14.

TABLE 13

Time (hr)	Mean Plasma Concentration vs. Time (ng/ml)			
	Treatment 3A	Treatment 3B	Treatment 3C	Treatment 3D
0	0.0084	0.0309	0.0558	0.0000
0.25			0.5074	0.9905
0.5	0.3853	0.3380	0.9634	1.0392
0.75			0.9753	1.3089
1	0.7710	0.7428	0.8777	1.3150
1.25			0.8171	1.2274
1.5	0.7931	1.0558	0.7109	1.1638
1.75			0.6357	1.0428
2	0.7370	1.0591	0.5851	0.9424
3	0.6879	0.9858	0.4991	0.7924
4	0.6491	0.9171	0.3830	0.7277
5	0.9312	1.4633	0.3111	0.6512
6	0.7613	1.0441	0.2650	0.4625
8	0.5259	0.7228	0.2038	0.2895
10	0.4161	0.5934	0.1768	0.2470
12	0.5212	0.5320	0.2275	0.2660
14	0.4527	0.4562	0.2081	0.2093
16	0.3924	0.3712	0.1747	0.1623
20	0.2736	0.3021	0.1246	0.1144
24	0.2966	0.2636	0.1022	0.1065
30	0.3460	0.3231		
36	0.2728	0.2456	0.0841	0.0743
48	0.1263	0.1241		

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TABLE 14

Pharmacokinetic Parameters of Plasma Oxymorphone for Study 3								
	Treatment 3B		Treatment 3A		Treatment 3C		Treatment 3D	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
C_{max}	1.7895	0.6531	1.1410	0.4537	2.2635	1.0008	3.2733	1.3169
T_{max}	5.65	9.39	5.57	7.14	0.978	1.14	1.11	0.768
$AUC_{(0-24)}$	14.27	4.976	11.64	3.869	12.39	4.116	17.30	5.259
$AUC_{(0-t)}$	19.89	6.408	17.71	8.471	14.53	4.909	19.20	6.030
$AUC_{(0-infty)}$	21.29	6.559	19.29	5.028	18.70	6.618	25.86	10.03
$T_{1/2rel}$	12.0	3.64	12.3	3.99	16.2	11.4	20.6	19.3

The relative bioavailability calculations are summarized in tables 15 and 16.

TABLE 15

Relative Bioavailability Determination Based on $AUC_{(0-24)}$			
F_{rel} (3A vs. 3C)	F_{rel} (3B vs. 3D)	F_{rel} (3D vs. 3C)	F_{rel} (3A vs. 3B)
1.040 ± 0.1874	0.8863 ± 0.2569	1.368 ± 0.4328	1.169 ± 0.2041

TABLE 16

Relative bioavailability Determination Based on $AUC_{(0-24)}$			
F_{rel} (3A vs. 2C)	F_{rel} (3B vs. 3D)	F_{rel} (3D vs. 3C)	F_{rel} (3A vs. 3B)
0.9598 ± 0.2151	0.8344 ± 0.100	1.470 ± 0.3922	1.299 ± 0.4638

The objectives of this study were to assess the relative bioavailability of oxymorphone from oxymorphone controlled release (20 mg) compared to oxymorphone oral solution (10 mg) under both fasted and fed conditions, and to determine the effect of food on the bioavailability of oxymorphone from the controlled release formulation, oxymorphone CR, and from the oral solution.

The presence of a high fat meal had a substantial effect on the oxymorphone C_{max} , but less of an effect on oxymorphone AUC from oxymorphone controlled release tablets. Least Squares (LS) mean C_{max} was 58% higher and LS mean $AUC_{(0-t)}$ and $AUC_{(0-infinity)}$ were 18% higher for the fed condition (Treatment B) compared to the fasted condition (Treatment A) based on LN-transformed data. This was consistent with the relative bioavailability determination from $AUC_{(0-infinity)}$ since mean F_{rel} was 1.17. Mean T_{max} values were similar (approximately 5.6 hours), and no significant difference in T_{max} was shown using nonparametric analysis. Half value durations were significantly different between the two treatments.

The effect of food on oxymorphone bioavailability from the oral solution was more pronounced, particularly in terms of AUC. LS mean C_{max} was 50% higher and LS mean $AUC_{(0-t)}$ and $AUC_{(0-infinity)}$ were 32-34% higher for the fed condition (Treatment D) compared to the fasted condition (Treatment C) based on LN-transformed data. This was consistent with the relative bioavailability determination from

$AUC_{(0-infinity)}$ since mean F_{rel} was 1.37. Mean T_{max} (approximately 1 hour) was similar for the two treatments and no significant difference was shown.

Under fasted conditions, oxymorphone controlled release 20 mg tablets exhibited similar extent of oxymorphone availability compared to 10 mg oxymorphone oral solution normalized to a 20 mg dose (Treatment A versus Treatment C). From LN-transformed data, LS mean $AUC_{(0-t)}$ was 17% higher for oxymorphone CR, whereas LS mean $AUC_{(0-infinity)}$ values were nearly equal (mean ratio=99%). Mean F_{rel} values calculated from $AUC_{(0-infinity)}$ and $AUC_{(0-24)}$, (1.0 and 0.96, respectively) also showed similar extent of oxymorphone availability between the two treatments.

As expected, there were differences in parameters reflecting rate of absorption. LS mean C_{max} was 49% lower for oxymorphone controlled release tablets compared to the dose-normalized oral solution, based on LN-transformed data. Half-value duration was significantly longer for the controlled release formulation (means, 12 hours versus 2.5 hours).

Under fed conditions, oxymorphone availability from oxymorphone controlled release 20 mg was similar compared to 10 mg oxymorphone oral solution normalized to a 20 mg dose (Treatment B versus Treatment D). From LN-transformed data, LS mean $AUC_{(0-infinity)}$ was 12% lower for oxymorphone CR. Mean F_{rel} values calculated from $AUC_{(0-infinity)}$ and $AUC_{(0-24)}$, (0.89 and 0.83 respectively) also showed similar extent of oxymorphone availability from the tablet. As expected, there were differences in parameters reflecting rate of absorption. LS mean C_{max} was 46% lower for oxymorphone controlled release tablets compared to the dose-normalized oral solution, based on LN-transformed data. Mean T_{max} was 5.7 hours for the tablet compared to 1.1 hours for the oral solution. Half-value duration was significantly longer for the controlled release formulation (means, 7.8 hours versus 3.1 hours).

The presence of a high fat meal did not appear to substantially affect the availability of 6-hydroxymorphone following administration of oxymorphone controlled release tablets. LS mean ratios were 97% for $AUC_{(0-t)}$ and 91% for C_{max} (Treatment B versus A), based on LN-transformed data. This was consistent with the relative bioavailability determination from $AUC_{(0-24)}$, since mean F_{rel} was 0.97. Mean T_{max} was

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later for the fed treatment compared to the fasted treatment (5.2 and 3.6 hours, respectively), and difference was significant.

Under fasted conditions, oxymorphone controlled release 20 mg tablets exhibited similar availability of 6-hydroxymorphone compared to 10 mg oxymorphone oral solution normalized to a 20 mg dose (Treatment A versus Treatment C). From LN-transformed data, LS mean ratio for $AUC_{(0-t)}$ was 104.5%. Mean F_{rel} (0.83) calculated from $AUC_{(0-24)}$ also showed similar extent of oxymorphone availability between the two treatments. Mean T_{max} was 3.6 hours for the tablet compared to 0.88 for the oral solution. Half-value duration was significantly longer for the controlled release formulation (means, 11 hours versus 2.2 hours).

Under fed conditions, availability of 6-hydroxymorphone from oxymorphone controlled release 20 mg was similar compared to 10 mg oxymorphone oral solution normalized to a 20 mg dose (Treatment B versus Treatment D). From LN-transformed data, LS mean $AUC_{(0-t)}$ was 14% higher for oxymorphone CR. Mean F_{rel} (0.87) calculated from $AUC_{(0-24)}$ also indicated similar extent of availability between the treatments. Mean T_{max} was 5.2 hours for the tablet compared to 1.3 hour for the oral solution. Half-value duration was significantly longer for the controlled release formulation (means, 14 hours versus 3.9 hours).

The extent of oxymorphone availability from oxymorphone controlled release 20 mg tablets was similar under fed and fasted conditions since there was less than a 20% difference in LS mean $AUC_{(0-t)}$ and $AUC_{(0-inf)}$ values for each treatment, based on LN-transformed data. T_{max} was unaffected by food; however, LS mean C_{max} was increased 58% in the presence of the high fat meal. Both rate and extent of oxymorphone absorption from the oxymorphone oral solution were affected by food since LS mean C_{max} and AUC values were increased approximately 50 and 30%, respectively. T_{max} was unaffected by food. Under both fed and fasted conditions, oxymorphone controlled release tablets exhibited similar extent of oxymorphone availability compared to oxy-

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ment. T_{max} was later for the fed condition. The presence of food did not affect the extent of availability from oxymorphone oral solution since LS mean AUC values were less than 20% different. However, C_{max} was decreased 35% in the presence of food. T_{max} was unaffected by food. Under both fed and fasted conditions, oxymorphone controlled release tablets exhibited similar extent of availability compared to oxymorphone oral solution since there was less than a 20% difference in LS mean AUC values for each treatment.

The mean 6-OH oxymorphone plasma concentration versus time curves for Treatments 3A, 3B, 3C, and 3D are presented in FIG. 8. The data is contained in Table 17.

TABLE 17

Time (hr)	Mean Plasma Concentration vs. Time (ng/ml) 6-Hydroxyoxymorphone			
	Treatment 3A	Treatment 3B	Treatment 3C	Treatment 3D
0	0.0069	0.0125	0.0741	0.0000
0.25			0.7258	0.4918
0.5	0.5080	0.1879	1.2933	0.5972
0.75			1.3217	0.7877
1	1.0233	0.4830	1.1072	0.8080
1.25			1.0069	0.7266
1.5	1.1062	0.7456	0.8494	0.7001
1.75			0.7511	0.6472
2	1.0351	0.7898	0.6554	0.5758
3	0.9143	0.7619	0.6196	0.5319
4	0.8522	0.7607	0.4822	0.5013
5	0.8848	0.8548	0.3875	0.4448
6	0.7101	0.7006	0.3160	0.3451
8	0.5421	0.5681	0.2525	0.2616
10	0.4770	0.5262	0.2361	0.2600
12	0.4509	0.4454	0.2329	0.2431
14	0.4190	0.4399	0.2411	0.2113
16	0.4321	0.4230	0.2385	0.2086
20	0.3956	0.4240	0.2234	0.1984
24	0.4526	0.4482	0.2210	0.2135
30	0.4499	0.4708		
36	0.3587	0.3697	0.1834	0.1672
48	0.3023	0.3279		

TABLE 18

Pharmacokinetic Parameters of Plasma Oxymorphone for Study 3								
Treatment 3A		Treatment 3B		Treatment 3C		Treatment 3D		
Mean	SD	Mean	SD	Mean	SD	Mean	SD	
C_{max}	1.2687	0.5792	1.1559	0.4848	1.5139	0.7616	0.9748	0.5160
T_{max}	3.61	7.17	5.20	9.52	0.880	0.738	1.30	1.04
$AUC_{(0-t)}$	22.47	10.16	22.01	10.77	10.52	4.117	9.550	4.281
$AUC_{(0-inf)}$	38.39	23.02	42.37	31.57	20.50	7.988	23.84	11.37
$T_{1/2el}$	39.1	36.9	39.8	32.6	29.3	12.0	44.0	35.00

morphine oral solution since there was less than a 20% difference in LS mean $AUC(0-t)$ and $AUC(0-inf)$ values for each treatment.

Bioavailability of 6-hydroxymorphone following oxymorphone controlled release 20 mg tablets was also similar under fed and fasted conditions since there was less than a 20% difference in LS mean C_{max} and AUC values for each treat-

60 Study 4-Controlled Release 20 mg vs Immediate Release 10 mg

A study was conducted to compare the bioavailability and pharmacokinetics of controlled release and immediate release oxymorphone tablets under single-dose and multiple-dose (steady state) conditions. For the controlled release study, healthy volunteers received a single dose of a 20 mg

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controlled release oxymorphone tablet on the morning of Day 1. Beginning on the morning of Day 3, the volunteers were administered a 20 mg controlled release oxymorphone tablet every 12 hours through the morning dose of Day 9. For the immediate release study, healthy volunteers received a single 10 mg dose of an immediate release oxymorphone tablet on the morning of Day 1. On the morning of Day 3, additional 10 mg immediate release tablets were administered every six hours through the first two doses on Day 9.

FIG. 9 shows the average plasma concentrations of oxymorphone and 6-hydroxyoxymorphone for all subjects after a single dose either controlled release (CR) 20 mg or immediate release (IR) 10 mg oxymorphone. The data in the figure (as with the other relative experimental data herein) is normalized to a 20 mg dose. The immediate release tablet shows a classical curve, with a high, relatively narrow peak followed by an exponential drop in plasma concentration. The controlled release oxymorphone tablets show a lower peak with extended moderate levels of oxymorphone and 6-hydroxy oxymorphone. Table 19 shows the levels of oxymorphone and 6-hydroxy oxymorphone from FIG. 9 in tabular form.

TABLE 19

Mean Plasma Concentration (ng/ml)												
Hour	Oxymorphone		6-Hydroxyoxymorphone		Oxymorphone				6-Hydroxyoxymorphone			
	Controlled Release 20 mg	Immediate Release 10 mg										
0.00	0.00	0.00	0.00	0.00	1.10	0.75	0.89	0.72				
0.25	0.22	1.08	0.14	0.73	1.12	0.84	1.15	0.88				
0.50	0.59	1.69	0.45	1.22	1.20	0.92	1.15	0.87				
1.00	0.77	1.19	0.53	0.79	1.19	0.91	1.27	1.00				
1.50	0.84	0.91	0.53	0.57	1.19	0.86	1.29	0.98				
2.00	0.87	0.75	0.60	0.47	1.03	1.07	1.09	1.05				
3.00	0.83	0.52	0.55	0.34	0.25	2.64	1.70					
4.00	0.73	0.37	0.53	0.27	0.50	3.12	1.50	2.09				
5.00	0.94	0.36	0.46	0.23	1.00	2.47	1.70	1.68				
6.00	0.81	0.28	0.41	0.18	1.50	2.05	1.63	1.55				
8.00	0.73	0.20	0.37	0.14	2.00	1.78	1.64	1.30				
10.0	0.60	0.19	0.35	0.15	3.00	1.27	1.47	1.11				
12.0	0.67	0.25	0.32	0.13	4.00	0.98	1.39	0.98				
16.0	0.39	0.16	0.29	0.13	5.00	1.01	1.21	0.89				
24.0	0.23	0.07	0.29	0.13	6.00	0.90	1.06	0.84				
30.0	0.12	0.01	0.17	0.04	6.25	1.17						
36.0	0.05	0.00	0.11	0.00	7.00	1.88						
48.0	0.00	0.00	0.07	0.01	7.50	2.12						
					8.00	1.32	2.24	1.15				
					9.00	2.01	0.97	1.03				
					10.0	1.52						
					11.0	1.24	0.85	0.84				
					12.0	0.96	1.11	0.74				
						1.18	0.79	0.70				

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FIG. 10 shows the average plasma concentrations of oxymorphone and 6-hydroxyoxymorphone for all subjects in the steady state test, for doses of controlled release 20 mg tablets and immediate release 10 mg tablets of oxymorphone. The figure shows the plasma concentrations after the final controlled release tablet is given on Day 9, and the final immediate release tablet is given 12 hours thereafter. The steady state administration of the controlled release tablets clearly shows a steady moderate level of oxymorphone ranging from just over 1 ng/ml to almost 1.75 ng/ml over the course of a twelve hour period, where the immediate release tablet shows wide variations in blood plasma concentration. Table 20 shows the levels of oxymorphone and 6-hydroxyoxymorphone from FIG. 10 in tabular form.

TABLE 20

Summary of Mean Plasma Concentration (ng/ml)							
Day	Hour	Oxymorphone		6-Hydroxyoxymorphone			
		Controlled Release 20 mg	Immediate Release 10 mg	Controlled Release 20 mg	Immediate Release 10 mg	Controlled Release 20 mg	Immediate Release 10 mg
4	0.00	1.10	0.75	0.89	0.72		
5	0.00	1.12	0.84	1.15	0.88		
6	0.00	1.20	0.92	1.15	0.87		
7	0.00	1.19	0.91	1.27	1.00		
8	0.00	1.19	0.86	1.29	0.98		
9	0.00	1.03	1.07	1.09	1.05		
		0.25	2.64				
		0.50	3.12	1.50	2.09		
		1.00	2.47	1.70	1.68		
		1.50	2.05	1.63	1.55		
		2.00	1.78	1.64	1.30		
		3.00	1.27	1.47	1.11		
		4.00	0.98	1.39	0.98		
		5.00	1.01	1.21	0.89		
		6.00	0.90	1.06	0.84		
		6.25	1.17				
		6.50	1.88				
		7.00	2.12				
		7.50	2.24				
		8.00	1.32	2.01	1.03		
		9.00		1.52	0.90		
		10.0	1.32	1.24	0.85	0.84	
		11.0		1.11		0.74	
		12.0	1.18	0.96	0.79	0.70	

TABLE 21

Mean Single-Dose Pharmacokinetic Results				
	Controlled Release 20 mg		Immediate Release 10 mg	
	6-OH-oxymorphone	oxymorphone	6-OH-oxymorphone	oxymorphone
AUC _(0->t)	14.74	11.54	7.10	5.66
AUC _(0->48h)	15.33	16.40	7.73	8.45
C _{max} (ng/ml)	1.12	0.68	1.98	1.40
T _{max} (hr)	5.00	2.00	0.50	0.50
T _{1/2} (hr)	9.25	26.09	10.29	29.48

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Parent 6-OH oxymorphone AUC_(0-t) values were lower than the parent compound after administration of either dosage form, but the AUC_(0-∞) values are slightly higher due to the longer half-life for the metabolite. This relationship was similar for both the immediate-release (IR) and controlled release (CR) dosage forms. As represented by the average plasma concentration graph, the CR dosage form has a significantly longer time to peak oxymorphone concentration and a lower peak oxymorphone concentration. The 6-OH oxymorphone peak occurred sooner than the parent peak following the CR dosage form, and simultaneously with the parent peak following the IR dosage form.

It is important to note that while the present invention is described and exemplified using 20 mg tablets, the invention may also be used with other strengths of tablets. In each strength, it is important to note how a 20 mg tablet of the same composition (except for the change in strength) would act. The blood plasma levels and pain intensity information are provided for 20 mg tablets, however the present invention is also intended to encompass 5 to 80 mg controlled release tablets. For this reason, the blood plasma level of oxymorphone or 6-hydroxyoxymorphone in nanograms per milliliter of blood, per mg oxymorphone (ng/mg·ml) administered is measured. Thus at 0.02 ng/mg·ml, a 5 mg tablet should produce a minimum blood plasma concentration of 0.1 ng/ml. A stronger tablet will produce a higher blood plasma concentration of active molecule, generally proportionally. Upon administration of a higher dose tablet, for example 80 mg, the blood plasma level of oxymorphone and 6-OH oxymorphone may more than quadruple compared to a 20 mg dose, although conventional treatment of low bioavailability substances would lead away from this conclusion. If this is the case, it may be because the body can only process a limited amount oxymorphone at one time. Once the bolus is processed, the blood level of oxymorphone returns to a proportional level.

It is the knowledge that controlled release oxymorphone tablets are possible to produce and effective to use, which is most important, made possible with the high bioavailability of oxymorphone in a controlled release tablet. This also holds true for continuous periodic administration of controlled release formulations. The intent of a controlled release opioid formulation is the long-term management of pain. Therefore, the performance of a composition when administered periodically (one to three times per day) over several days is important. In such a regime, the patient reaches a "steady state" where continued administration will produce the same results, when measured by duration of pain relief and blood plasma levels of pharmaceutical. Such a test is referred to as a "steady state" test and may require periodic administration over an extended time period ranging from several days to a week or more. Of course, since a patient reaches steady state in such a test, continuing the test for a longer time period should not affect the results. Further, when testing blood plasma levels in such a test, if the time period for testing exceeds the interval between doses, it is important the regimen be stopped after the test is begun so that observations of change in blood level and pain relief may be made without a further dose affecting these parameters.

Study 5-Controlled Release 40 mg vs Immediate Release 4.Times.10 mg under Fed and Fasting Conditions

The objectives of this study were to assess the relative bioavailability of oxymorphone from oxymorphone controlled release (40 mg) compared to oxymorphone immediate release (4.Times.10 mg) under both fasted and fed conditions, and to determine the effect of food on the bioavailability of

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oxymorphone from the controlled release formulation, oxymorphone CR, and from the immediate release formulation, oxymorphone IR.

This study had a single-center, open-label, analytically blinded, randomized, four-way crossover design. Subjects randomized to Treatment 5A and Treatment 5C, as described below, were in a fasted state following a 10-hour overnight fast. Subjects randomized to Treatment 5B and Treatment 5D, as described below, were in the fed state, having had a high fat meal, completed ten minutes prior to dosing. There was a 14-day washout interval between the four dose administrations. The subjects were confined to the clinic during each study period. Subject assigned to receive Treatment 5A and Treatment 5B were discharged from the clinic on Day 3 following the 48-hour procedures, and subjects assigned to receive Treatment 5C and Treatment 5D were discharged from the clinic on Day 2 following the 36-hour procedures. On Day 1 of each study period the subjects received one of four treatments:

Treatments 5A and 5B: Oxymorphone controlled release 40 mg tablets from Table 2. Subjects randomized to Treatment 5A received a single oral dose of one 40 mg oxymorphone controlled release tablet taken with 240 ml of water after a 10-hour fasting period. Subjects randomized to Treatment 5B received a single oral dose of one 40 mg oxymorphone controlled release tablet taken with 240 ml of water 10 minutes after a standardized high fat meal.

Treatments 5C and 5D: Immediate release tablet (IR) 4.times.10 mg Oxymorphone. Subjects randomized to Treatment 5C received a single oral dose of 4.times.10 mg oxymorphone IR tablet taken with 240 ml of water after a 10-hour fasting period. Subjects randomized to Treatment 5D received a single oral dose of 4.times.10 mg oxymorphone IR tablet taken with 240 ml of water 10 minutes after a standardized high-fat meal.

A total of 28 male subjects were enrolled in the study, and 25 subjects completed the study. A total of 28 subjects received at least one treatment. Only subjects who completed all 4 treatments were included in the summary statistics and statistical analysis.

Blood samples (7 ml) were collected during each study period at the 0 hour (predose), and at 0.25, 0.5, 0.75, 1.0, 1.5, 2, 3, 4, 5, 6, 8, 10, 12, 24, 36, 48, 60, and 72 hours post-dose (19 samples) for subjects randomized to all Treatments.

The mean oxymorphone plasma concentration versus time is presented in Table 22. The arithmetic means of the plasma oxymorphone pharmacokinetic parameters and the statistics for all Treatments are summarized in Table 23.

TABLE 22

Time (hr)	Mean Plasma Concentration vs. Time (ng/ml)			
	Treatment 5A	Treatment 5B	Treatment 5C	Treatment 5D
0	0.00	0.00	0.00	0.00
0.25	0.47	0.22	3.34	1.79
0.50	1.68	0.97	7.28	6.59
0.75	1.92	1.90	6.60	9.49
1	2.09	2.61	6.03	9.91
1.5	2.18	3.48	4.57	8.76
2	2.18	3.65	3.68	7.29
3	2.00	2.86	2.34	4.93
4	1.78	2.45	1.65	3.11
5	1.86	2.37	1.48	2.19
6	1.67	2.02	1.28	1.71
8	1.25	1.46	0.92	1.28
10	1.11	1.17	0.78	1.09
12	1.34	1.21	1.04	1.24

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TABLE 22-continued

Time (hr)	Mean Plasma Concentration vs. Time (ng/ml)			
	Treatment 5A	Treatment 5B	Treatment 5C	Treatment 5D
24	0.55	0.47	0.40	0.44
36	0.21	0.20	0.16	0.18
48	0.06	0.05	0.04	0.05
60	0.03	0.01	0.01	0.01
72	0.00	0.00	0.00	0.00

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TABLE 26-continued

Time (hr)	Mean Plasma Concentration vs. Time (ng/ml)			
	Treatment 5A	Treatment 5B	Treatment 5C	Treatment 5D
48	0.18	0.20	0.20	0.19
60	0.09	0.10	0.09	0.09
72	0.06	0.06	0.04	0.05

TABLE 23

Pharmacokinetic Parameters of Plasma Oxymorphone for Study 5									
	Treatment 5A		Treatment 5B		Treatment 5C		Treatment 5D		
	Mean	SD	Mean	SD	Mean	SD	Mean	SD	
C_{max}	2.79	0.84	4.25	1.21	9.07	4.09	12.09	5.42	
T_{max}	2.26	2.52	1.96	1.06	0.69	0.43	1.19	0.62	
$AUC_{(0-t)}$	35.70	10.58	38.20	11.04	36.00	12.52	51.35	20.20	
$AUC_{(0-\infty)}$	40.62	11.38	41.17	10.46	39.04	12.44	54.10	20.26	
$T_{1/2\alpha}$	12.17	7.57	10.46	5.45	11.65	6.18	9.58	3.63	

The relative bioavailability calculations are summarized in Tables 24 and 25.

TABLE 24

Relative Bioavailability Determination Based on $AUC_{(0-\infty)}$	
F_{rel} (5D vs. 5C)	F_{rel} (5B vs. 5A)
1.3775	1.0220

TABLE 25

Relative bioavailability Determination Based on $AUC_{(0-24)}$	
F_{rel} (5D vs. 5C)	F_{rel} (5B vs. 5A)
1.4681	1.0989

The mean 6-OH oxymorphone plasma concentration versus time is presented in Table 26.

TABLE 26

Time (hr)	Mean Plasma Concentration vs. Time (ng/ml)			
	Treatment 5A	Treatment 5B	Treatment 5C	Treatment 5D
0	0.00	0.00	0.00	0.00
0.25	0.27	0.05	2.36	0.50
0.50	1.32	0.31	5.35	1.98
0.75	1.37	0.59	4.53	2.97
1	1.44	0.82	3.81	2.87
1.5	1.46	1.09	2.93	2.58
2	1.46	1.28	2.37	2.29
3	1.39	1.14	1.69	1.72
4	1.25	1.14	1.33	1.26
5	1.02	1.00	1.14	1.01
6	0.93	0.86	0.94	0.86
8	0.69	0.72	0.73	0.77
10	0.68	0.67	0.66	0.75
12	0.74	0.66	0.70	0.77
24	0.55	0.52	0.54	0.61
36	0.23	0.30	0.28	0.27

TABLE 27

Time (hr)	Pharmacokinetic Parameters of Plasma 6-Hydroxyoxymorphone for Study 5			
	Treatment 5A	Treatment 5B	Treatment 5C	Treatment 5D
C_{max}	1.88	0.69	1.59	0.63
T_{max}	1.48	1.18	2.73	1.27
$AUC_{(0-t)}$	28.22	10.81	26.95	11.39
$AUC_{(0-\infty)}$	33.15	11.25	32.98	10.68
$T_{1/2\alpha}$	17.08	7.45	21.92	8.41
$T_{1/2\beta}$	16.01	6.68	16.01	7.42

The above description incorporates preferred embodiments and examples as a means of describing and enabling the invention to be practiced by one of skill in the art. It is imagined that changes can be made without departing from the spirit and scope of the invention described herein and defined in the appended claims.

We claim:

- An oral controlled release oxymorphone formulation, comprising:
 - about 5 mg to about 80 mg of oxymorphone or a pharmaceutically acceptable salt of oxymorphone; and
 - a hydrophilic material,
- wherein upon oral administration of the formulation to a subject in need of an analgesic effect:
 - the formulation provides detectable blood plasma levels of 6-OH oxymorphone and oxymorphone;
 - the blood plasma levels of 6-OH oxymorphone and oxymorphone peak within about 1 hour to about 8 hours after administration;
 - the blood plasma levels of 6-OH oxymorphone and oxymorphone exhibit a ratio of area under the curve ($AUC_{(0 \text{ to } t_{\infty})}$) of blood plasma level versus time for 6-OH oxymorphone compared to oxymorphone in a range of about 0.5 to about 1.5;
 - the duration of the analgesic effect is through at least about 12 hours after administration; and
 - the blood plasma levels of oxymorphone exhibit two or three peaks within about 12 hours after administration.
- The formulation of claim 1 wherein the hydrophilic material is selected from the group consisting of a gum, a cellulose ether, an acrylic resin, a protein-derived material, and mixtures thereof.
- The formulation of claim 1 wherein the hydrophilic material is a gum selected from the group consisting of a heteropolysaccharide gum, a homopolysaccharide gum, and mixtures thereof.
- The formulation of claim 3 wherein the gum is selected from the group consisting of xanthan, tragacanth, acacia, karaya, alginates, agar, guar, hydroxypropyl guar, carrageenan, locust bean, and mixtures thereof.

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5. The formulation of claim 1 wherein the hydrophilic material is a cellulose ether selected from the group consisting of a hydroxyalkyl cellulose, a carboxyalkyl cellulose, and mixtures thereof.

6. The formulation of claim 1 wherein the hydrophilic material is selected from the group consisting of hydroxyethyl cellulose, hydroxypropyl cellulose, hydroxypropyl methylcellulose, carboxymethylcellulose, and mixtures thereof.

7. The formulation of claim 1 wherein the hydrophilic material comprises at least one of:

- i. a heteropolysaccharide; or
- ii. a heteropolysaccharide and a cross-linking agent capable of cross-linking the heteropolysaccharide; or
- iii. a mixture of (i), (ii) and a polysaccharide gum.

8. The formulation of claim 7 wherein the heteropolysaccharide is a water soluble polysaccharide containing two or more kinds of sugar units and having a branched or helical configuration.

9. The formulation of claim 7 wherein the heteropolysaccharide is selected from the group consisting of xanthan gum, deacylated xanthan gum, carboxymethyl ether xanthan gum, propylene glycol ester xanthan gum and mixtures thereof.

10. The formulation of claim 7 wherein the cross-linking agent is a homopolysaccharide gum.

11. The formulation of claim 1 further comprising a hydrophobic polymer.

12. A method of treating pain in a subject in need thereof, the method comprising the step of administering to the subject the formulation of claim 1.

13. A pharmaceutical tablet prepared by:

- a. mixing oxymorphone or a pharmaceutically acceptable salt of oxymorphone and controlled release granules comprising a hydrophilic material and one or more optional excipients; and
- b. directly compressing the mixture of (a) to form the tablet,

wherein upon placement of the tablet in an in vitro dissolution test comprising USP Paddle Method at 50 rpm in 500 ml media having a pH of 1.2 to 6.8 at 37° C., about 15% to about 50%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 1 hour in the test.

14. The tablet preparation of claim 13 wherein the hydrophilic material is selected from the group consisting of a gum, a cellulose ether, an acrylic resin, a protein-derived material, and mixtures thereof.

15. The tablet preparation of claim 13 wherein the hydrophilic material is a gum selected from the group consisting of a heteropolysaccharide gum, a homopolysaccharide gum, and mixtures thereof.

16. The tablet preparation of claim 13 wherein the hydrophilic material is a cellulose ether selected from the group consisting of a hydroxyalkyl cellulose, a carboxyalkyl cellulose, and mixtures thereof.

17. The tablet preparation of claim 13 wherein the hydrophilic material is hydroxyethyl cellulose, hydroxypropyl cellulose, hydroxypropyl methylcellulose, carboxymethylcellulose, and mixtures thereof.

18. The tablet preparation of claim 13 wherein the hydrophilic material comprises at least one of:

- i. a heteropolysaccharide; or
- ii. a heteropolysaccharide and a cross-linking agent capable of cross-linking the heteropolysaccharide; or
- iii. a mixture of (i), (ii) and a polysaccharide gum.

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19. The tablet preparation of claim 18 wherein the heteropolysaccharide is a water soluble polysaccharide containing two or more kinds of sugar units and having a branched or helical configuration.

20. The tablet preparation of claim 19 wherein the heteropolysaccharide is selected from the group consisting of xanthan gum, deacylated xanthan gum, carboxymethyl ether xanthan gum, propylene glycol ester xanthan gum and mixtures thereof.

21. A pharmaceutical tablet prepared by:

- a. mixing oxymorphone or a pharmaceutically acceptable salt of oxymorphone and one or more controlled release excipients; and
- b. forming the tablet,

wherein upon placement of the tablet in an in vitro dissolution test comprising USP Paddle Method at 50 rpm in 500 ml media having a pH of 1.2 to 6.8 at 37° C., about 15% to about 50%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 1 hour in the test; and wherein upon oral administration to a human subject the tablet alleviates pain for 12 to 24 hours.

22. The tablet of claim 21 wherein about 45% to about 80%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 4 hours in the test, and at least about 80%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 10 hours in the test.

23. The tablet of claim 21 wherein at least 27%, by weight, of the oxymorphone or salt thereof is released from the tablet

at about 1 hour in the test, at least 40%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 2 hours in the test, at least 50%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 3 hours in the test, at least 64%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 5 hours in the test, at least 70%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 6 hours in the test, at least 79%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 8 hours in the test, at least 85%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 10 hours in the test, and at least 89%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 12 hours in the test.

24. The tablet of claim 21, wherein at least 27%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 1 hour in the test.

25. The tablet of claim 21, wherein at least 40%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 2 hours in the test.

26. The tablet of claim 21, wherein at least 50%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 3 hours in the test.

27. The tablet of claim 21, wherein at least 64%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 5 hours in the test.

28. The tablet of claim 21, wherein at least 70%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 6 hours in the test.

29. The tablet of claim 21, wherein at least 79%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 8 hours in the test.

30. The tablet of claim 21, wherein at least 85%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 10 hours in the test.

31. A method for treating pain in a human subject in need of acute or chronic pain relief, comprising the steps of:

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(a) Providing a solid oral dosage form of a controlled release oxymorphone formulation with a release rate profile designed to provide adequate blood plasma levels over at least 12 hours to provide sustained pain relief over this same period comprising about 5 mg to about 80 mg oxymorphone or a pharmaceutically acceptable salt thereof wherein oxymorphone is the sole active ingredient, and wherein upon placement of the composition in an *in vitro* dissolution test comprising USP Paddle Method at 50 rpm in 500 ml media having a pH of 1.2 to 6.8 at 37° C., about 15% to about 50%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 1 hour in the test; and

(b) administering a single dose of the dosage form to the subject,

wherein the oxymorphone C_{max} is at least 50% higher when the dosage form is administered to the subject under fed as compared to fasted conditions.

32. The method of claim 31 wherein the dosage form comprises about 40 mg oxymorphone or a pharmaceutically acceptable salt thereof, and wherein the oxymorphone C_{max} is about 58% higher when the dosage form is administered to the subject under fed as compared to fasted conditions.

33. The method of claim 31 wherein the dosage form comprises about 20 mg oxymorphone or a pharmaceutically acceptable salt thereof.

34. The method of claim 31 wherein the dosage form comprises about 20 mg to about 40 mg oxymorphone hydrochloride.

35. The method of claim 31 wherein the difference in the oxymorphone area under the curve ($AUC_{(0-\infty)}$) between fed and fasted conditions is less than 20%.

36. The method of claim 35 wherein the difference in $AUC_{(0-\infty)}$ between fed and fasted conditions is about 18%.

37. The method of claim 31 wherein upon oral administration of the dosage form to the subject under fed or fasting conditions:

- (i) the dosage form provides detectable blood plasma levels of 6-OH oxymorphone and oxymorphone;
- (ii) the blood plasma levels of 6-OH oxymorphone and oxymorphone peak within about 1 hour to about 8 hours after administration; and
- (iii) the blood plasma levels of 6-OH oxymorphone and oxymorphone exhibit a ratio of $AUC_{(0-\infty)}$ of blood plasma level versus time for 6-OH oxymorphone compared to oxymorphone in a range of about 0.5 to about 1.5.

38. A method for treating pain in a human subject in need of acute or chronic pain relief, comprising the steps of:

- (a) Providing a solid oral dosage form comprising about 5 mg to about 80 mg oxymorphone or a pharmaceutically acceptable salt thereof in a controlled release delivery system with a release rate profile designed to provide adequate blood plasma levels over at least 12 hours to provide sustained pain relief over this same period, wherein oxymorphone is the sole active ingredient, and wherein upon placement of the composition in an *in vitro* dissolution test comprising USP Paddle Method at 50 rpm in 500 ml media having a pH of 1.2 to 6.8 at 37° C., about 15% to about 50%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 1 hour in the test, about 45% to about 80%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 4 hours in the test, and at least about 80%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 10 hours in the test; and

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(b) administering a single dose of the dosage form to the subject, wherein the oxymorphone C_{max} is at least 50% higher when the dosage form is administered to the subject under fed versus fasted conditions.

39. The method of claim 38 wherein the oxymorphone C_{max} is at least about 58% higher when the dosage form is administered to the subject under fed as compared to fasted conditions.

40. The method of claim 38 wherein the difference in the oxymorphone area under the curve $AUC_{(0-\infty)}$ between fed and fasted conditions is less than 20%.

41. The method of claim 40 wherein the difference in $AUC_{(0-\infty)}$ between fed and fasted conditions is about 18%.

42. The method of claim 38 wherein upon oral administration of the dosage form to the subject under fed or fasting conditions:

- (i) the dosage form provides detectable blood plasma levels of 6-OH oxymorphone and oxymorphone;
- (ii) the blood plasma levels of 6-OH oxymorphone and oxymorphone peak within about 1 hour to about 8 hours after administration; and
- (iii) the blood plasma levels of 6-OH oxymorphone and oxymorphone exhibit a ratio of $AUC_{(0-\infty)}$ of blood plasma level versus time for 6-OH oxymorphone compared to oxymorphone in a range of about 0.5 to about 1.5.

43. The method of claim 38 wherein the system further comprises a hydrophilic material.

44. The method of claim 43 wherein the hydrophilic material is selected from the group consisting of a gum, a cellulose ether, an acrylic resin, a protein-derived material, and mixtures thereof.

45. The method of claim 44 wherein the hydrophilic material is a gum selected from the group consisting of xanthan, tragacanth, acacia, karaya, alginates, agar, guar, hydroxypropyl guar, carrageenan, locust bean, and mixtures thereof.

46. The method of claim 43 wherein the hydrophilic material is a cellulose ether selected from the group consisting of a hydroxyalkyl cellulose, a carboxyalkyl cellulose, and mixtures thereof.

47. The method of claim 43 wherein the hydrophilic material is selected from the group consisting of hydroxyethyl cellulose, hydroxypropyl cellulose, hydroxypropyl methylcellulose, carboxymethylcellulose, and mixtures thereof.

48. The method of claim 43 wherein the hydrophilic material comprises at least one of:

- a. a heteropolysaccharide; or
- b. a heteropolysaccharide and a cross-linking agent capable of cross-linking the heteropolysaccharide; or
- c. a mixture of (a), (b) and a polysaccharide gum.

49. An analgesically effective controlled release pharmaceutical composition for oral delivery, comprising:

- a. a controlled release delivery system with a release rate profile designed to provide adequate blood plasma levels over at least 12 hours to provide sustained pain relief over this same period; and
- b. about 5 mg to about 80 mg of oxymorphone or a pharmaceutically acceptable salt of oxymorphone, wherein oxymorphone is the sole active ingredient, wherein upon oral administration of a single dose of the composition to a human subject, the oxymorphone C_{max} is at least 50% higher when the dose is administered to the subject under fed as compared to fasted conditions, and wherein upon placement of the composition in an *in vitro* dissolution test comprising USP Paddle Method at 50 rpm in 500 ml media having a pH of 1.2 to 6.8 at 37°

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C., about 15% to about 50%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 1 hour in the test.

50. The composition of claim 49 wherein upon oral administration thereof the oxymorphone $AUC_{(0 \rightarrow \infty)}$ is no more than 20% higher when the dosage form is administered to the subject under fed as compared to fasted conditions.

51. The composition of claim 49 wherein the dosage form comprises about 40 mg oxymorphone, and wherein the oxymorphone C_{max} is about 58% higher when the dosage form is administered to the subject under fed as compared to fasted conditions.

52. The composition of claim 49 wherein the controlled release delivery system comprises a heteropolysaccharide and an agent capable of cross-linking the heteropolysaccharide in presence of gastrointestinal fluid.

53. The composition of claim 52 wherein the heteropolysaccharide and the agent capable of cross-linking the heteropolysaccharide are present in a weight ratio of about 1:3 to about 3:1.

54. The composition of claim 49 wherein about 45% to about 80%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 4 hours in the test, and at least about 80%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 10 hours in the test.

55. An analgesically effective controlled release pharmaceutical composition for oral delivery, comprising:

- a. a controlled release delivery system with a release rate profile designed to provide adequate blood plasma levels of oxymorphone and 6-hydroxy-oxymorphone over at least 12 hours to provide sustained pain relief over this same period; and
 - b. about 5 mg to about 80 mg of oxymorphone or a pharmaceutically acceptable salt of oxymorphone, wherein oxymorphone is the sole active ingredient,
- wherein upon placement of the composition in an in vitro dissolution test comprising USP Paddle Method at 50 rpm in 500 ml media having a pH of 1.2 to 6.8 at 37° C., about 15% to about 50%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 1 hour in the test.

56. The composition of claim 55, wherein upon oral administration of a single dose of the composition to a human subject, the oxymorphone C_{max} is at least 50% higher when the dose is administered to the subject under fed as compared to fasted conditions.

57. The composition of claim 55, wherein the composition is in the form of a tablet and wherein at least 27%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 1 hour in the test, at least 40%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 2 hours in the test, at least 50%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 3 hours in the test, at least 64%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 5 hours in the test, at least 70%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 6 hours in the test, at least 79%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 8 hours in the test, at least 85%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 10 hours in the test, and at least 89%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 12 hours in the test.

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58. The composition of claim 55, wherein the composition is in the form of a tablet and wherein at least 27%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 1 hour in the test.

59. The composition of claim 55, wherein the composition is in the form of a tablet and wherein at least 40%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 2 hours in the test.

60. The composition of claim 55, wherein the composition is in the form of a tablet and wherein at least 50%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 3 hours in the test.

61. The composition of claim 55, wherein the composition is in the form of a tablet and wherein at least 64%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 5 hours in the test.

62. The composition of claim 55, wherein the composition is in the form of a tablet and wherein at least 70%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 6 hours in the test.

63. The composition of claim 55, wherein the composition is in the form of a tablet and wherein at least 79%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 8 hours in the test.

64. The composition of claim 55, wherein the composition is in the form of a tablet and wherein at least 85%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 10 hours in the test.

65. The composition of claim 55, wherein the composition is in the form of a tablet and wherein at least 89%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 12 hours in the test.

66. An analgesically effective controlled release pharmaceutical composition for oral delivery, comprising:

- a. a controlled release delivery system with a release rate profile designed to provide adequate blood plasma levels over at least 12 hours to provide sustained pain relief over this same period; and
 - b. about 5 mg to about 80 mg of oxymorphone or a pharmaceutically acceptable salt of oxymorphone, wherein oxymorphone is the sole active ingredient,
- wherein upon placement of the composition in an in vitro dissolution test comprising USP Paddle Method at 50 rpm in 500 ml media having a pH of 1.2 to 6.8 at 37° C., about 15% to about 50%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 1 hour in the test, and wherein upon oral administration of the composition to a human subject, the blood plasma levels of oxymorphone comprise one or more peaks.

67. The composition of claim 66 wherein the blood plasma levels comprise two peaks.

68. The composition of claim 66 wherein upon oral administration of the composition to a subject in need of an analgesic effect:

- (i) the composition provides detectable blood plasma levels of 6-OH oxymorphone and oxymorphone;
- (ii) the blood plasma levels of 6-OH oxymorphone and oxymorphone peak within about 1 hour to about 8 hours after administration; and
- (iii) the blood plasma levels of 6-OH oxymorphone and oxymorphone exhibit a ratio of area under the curve ($AUC_{(0 \rightarrow \infty)}$) of blood plasma level versus time for 6-OH oxymorphone compared to oxymorphone in a range of about 0.5 to about 1.5.

69. The composition of claim 66 wherein upon oral administration of the composition to a subject in need of an anal-

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gesic effect the blood plasma levels of oxymorphone exhibit two or three peaks within about 12 hours after administration.

70. The composition of claim 66 wherein upon oral administration of the composition to a subject in need of an analgesic effect the blood plasma levels of oxymorphone comprise a first peak at about 3 hours after administration and a second peak at about 6-7 hours after administration.

71. The composition of claim 66 wherein the composition is in the form of a tablet and about 45% to about 80%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 4 hours in the test, and at least about 80%, by weight, of the oxymorphone or salt thereof is released from the tablet at about 10 hours in the test.

72. A controlled release pharmaceutical composition comprising oxymorphone or a pharmaceutically acceptable salt thereof as the sole active ingredient and a controlled release matrix, comprising about 10% to about 75% (by total weight of the controlled release matrix) of a gelling agent which forms a gel upon exposure to gastrointestinal fluid;

wherein upon placement of the composition in an in vitro dissolution test comprising USP paddle method at 50 rpm in 500 ml media having a pH of 1.2 to 6.8 at 37° C., about 15% to about 50%, by weight, of the oxymorphone or salt thereof is released from the composition after about 1 hour in the test.

73. The pharmaceutical composition of claim 72 wherein about 45% to about 80%, by weight, of the oxymorphone or salt thereof is released from the composition after about 4 hours in the test.

74. The pharmaceutical composition of claim 72 wherein at least 80%, by weight, of the oxymorphone or salt thereof is released from the composition after about 10 hours in the test.

75. The pharmaceutical composition of claim 72 wherein upon oral administration of the dosage form to a human subject in need of an analgesic effect, the blood plasma concentration of oxymorphone comprises one or peaks.

76. The pharmaceutical composition of claim 72 wherein upon oral administration of the dosage form to a human subject in need of an analgesic effect, the blood plasma concentration of oxymorphone comprises a first peak at about 3 hours after administration and a second peak at about 6-7 hours after administration; and wherein

- (i) the dosage form provides detectable blood plasma levels of 6-OH oxymorphone and oxymorphone;
- (ii) the blood plasma levels of 6-OH oxymorphone and oxymorphone peak within about 1 hour to about 8 hours after administration;
- (iii) the blood plasma levels of 6-OH oxymorphone and oxymorphone exhibit a ratio of area under the curve ($AUC_{(0 \text{ to } \infty)}$) of blood plasma level versus time for 6-OH oxymorphone compared to oxymorphone in a range of about 0.5 to about 1.5; and
- (iv) the duration of the analgesic effect is through at least about 12 hours after administration.

77. A controlled release pharmaceutical composition comprising oxymorphone or pharmaceutically acceptable salt thereof as the sole active ingredient, and a controlled release matrix comprising about 10% to about 75% (by total weight of the controlled release matrix) of a gelling agent which forms a gel upon exposure to gastrointestinal fluid;

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wherein upon placement of the composition in an in vitro dissolution test comprising USP paddle method at 50 rpm in 500 ml media having a pH of 1.2 to 6.8 at 37° C., about 15% to about 50%, by weight, of the oxymorphone or salt thereof is released from the composition after about 1 hour in the test, about 45% to about 80%, by weight, of the oxymorphone or salt thereof is released from the composition after about 4 hours in the test, and at least 80%, by weight, of the oxymorphone or salt thereof is released from the composition after about 10 hours in the test,

wherein upon oral administration of a single dose of the composition to a human subject, the composition provides an oxymorphone C_{max} of at least 50% higher when the dose is administered to the subject under fed as compared to fasted conditions and provides a difference in oxymorphone $AUC_{(0 \text{ to } \infty)}$ of less than 20% higher when the dose is administered to the subject under fed as compared to fasted conditions.

78. The pharmaceutical composition of claim 77 wherein upon oral administration of the dosage form to a human subject in need of an analgesic effect the blood plasma level of oxymorphone displays two or three peaks over about the first 12 hours after administration; and

- (i) the dosage form provides detectable blood plasma levels of 6-OH oxymorphone and oxymorphone;
- (ii) the blood plasma levels of 6-OH oxymorphone and oxymorphone peak within about 1 hour to about 8 hours after administration;
- (iii) the blood plasma levels of 6-OH oxymorphone and oxymorphone exhibit a ratio of area under the curve ($AUC_{(0 \text{ to } \infty)}$) of blood plasma level versus time for 6-OH oxymorphone compared to oxymorphone in a range of about 0.5 to about 1.5; and
- (iv) the duration of the analgesic effect is through at least about 12 hours after administration.

79. The pharmaceutical composition of claim 77 wherein about 58% to about 66%, by weight, of the oxymorphone or salt thereof is released from the composition after about 4 hours in the test.

80. The pharmaceutical composition of claim 77 wherein about 85% to about 96%, by weight, of the oxymorphone or salt thereof is released from the composition after about 10 hours in the test.

81. A method of treating pain in a subject in need thereof, the method comprising administering to the subject the pharmaceutical composition of claim 72 in an amount sufficient to provide the subject with about 5 mg to about 80 mg of oxymorphone or salt thereof, wherein upon oral administration of a single dose of the composition to a human subject, the composition provides an oxymorphone C_{max} of at least 50% higher when the dose is administered to the subject under fed as compared to fasted conditions and provides a difference in oxymorphone $AUC_{(0 \text{ to } \infty)}$ of less than 20% higher when the dose is administered to the subject under fed as compared to fasted conditions.

82. A method of treating pain in a subject in need thereof, the method comprising administering to the subject the pharmaceutical composition of claim 77 in an amount sufficient to provide the subject with about 5 mg to about 80 mg of oxymorphone or salt thereof.

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